St. Marys River Area of Concern CANADIAN SECTION

Status of the Degradation of Fish and Wildlife Populations Beneficial Use Impairment

October 2023

Executive Summary

In 1987, Canada and the United States designated the St. Marys River as one of 43 Areas of Concern (AOC) in the Great Lakes basin, and both countries committed to restoring it under the Canada-U.S. Great Lakes Water Quality Agreement. The fish component of the *Degradation of Fish and Wildlife Populations* was identified as one of ten beneficial use impairments (BUI) in the St. Marys River Stage 1 Remedial Action Plan (RAP) in 1992 and again in the Stage 2 RAP (2002). The wildlife component was never deemed impaired, but instead was determined to "Require Further Assessment".

Since the river was designated as an AOC, vast improvements have been made to help restore water quality and ecosystem health. This report provides a summary on these improvements that have helped to support healthy fish and wildlife populations in the St. Marys River.

This document serves as an official record to account and recommend a change in status, on the Canadian side of the St. Marys River AOC, of the *Degradation of Fish and Wildlife Populations* BUI to "Not Impaired". The re-designation is supported by studies that address the Stage 2 RAP recommended actions on fish and wildlife and that showcase how the delisting criteria have been met. Specifically:

- Fisheries and Oceans Canada completed an overall assessment of the AOC fish community using the index of biotic integrity (IBI) approach in two separate studies. Both studies conclude the St. Marys River is home to a relatively healthy fish community that is complex, diverse, and dominated by native species (Appendix 1a, 1b).
- Results from the binational St. Marys River Fisheries Task Group's fish community gillnet survey and the river-wide Creel survey were used to confirm the status of managed¹ fish populations, with key findings revealing that managed fish populations appear to be stable or increasing (Appendix 2a, 2b).
- The wildlife component of the *Degradation of Fish and Wildlife Populations* BUI was designated as "Requires Further Assessment" in the Stage 1 and Stage 2 RAP reports (never deemed Impaired), and there were specific monitoring actions recommended in the Stage 2 RAP as the means to study the matter (i.e., Actions FFM-2, FFM-5, FFM-6, and FFM-8). These actions were completed, and an assessment of wildlife populations completed in September 2018 (Derickx, 2018) found wildlife to be in good condition and conclude it is not impaired (Appendix 3a-c).

A community review and engagement period on the assessment results and proposed re-designation was completed. Feedback and suggestions were taken into consideration and incorporated within this document as appropriate. At the end of the engagement period there were no objections to the recommendation to redesignate the *Degradation of Fish and Wildlife Populations* BUI to 'not impaired". A detailed account of the engagement process can be found in Appendix 4.

¹ These studies helped to assess the sport fisheries and to supplement the Index of Biotic Integrity approach that looks at nearshore fish communities. All the studies combined help to complement each other and showcase a picture of the fish populations in the St. Marys River as a whole.

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1.0 Introduction

1.1 The St. Marys River Area of Concern

The St. Marys River is a 112km binational waterway that connects Lake Superior to the North Channel of Lake Huron. The St. Marys River is one of the 43 Great Lakes Areas of Concern originally identified under *the Canada-United States Great Lakes Water Quality Agreement* (GLWQA). An Area of Concern (AOC) is a location with historically significant environmental impairment resulting from human activities at the local level. The St. Marys River, as a connecting channel, is one of five AOCs jointly shared by Canada and the United States.

The Canadian portion of the AOC extends from its head at Gros Cap in Whitefish Bay downstream to St. Joseph Island via Lake George to Quebec Bay in the St. Joseph Channel and downstream to Hay Point on the western shore of St. Joseph Island (Figure 1).

The St. Marys River AOC has a Remedial Action Plan that identifies the original beneficial use impairments and guides efforts to restore beneficial uses. Beneficial Use Impairments (BUIs) are reductions in the chemical, physical, or biological integrity of the area sufficient to cause environmental issues. These environmental problems were first identified in the Stage 1 RAP report (1992). In 2002, a number of remedial actions and monitoring initiatives to address the problems were identified, and restoration efforts have been underway since.

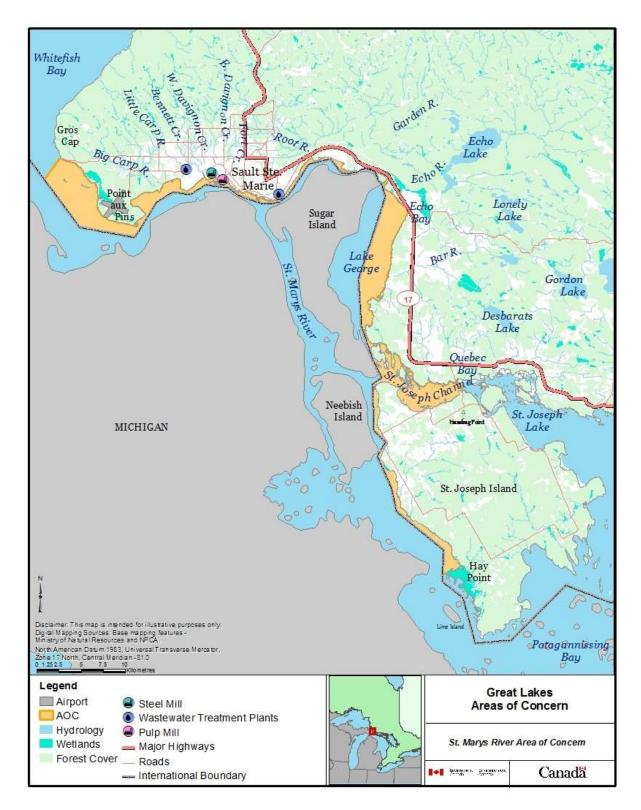


Figure 1: St. Marys River Area of Concern Map (Canadian Section).

1.2 Beneficial Use Impairment

The *Degradation of Fish and Wildlife Populations* BUI was listed as "Impaired" in the Stage 1 RAP report (1992) because native fish populations were being effected by habitat alteration, overfishing, pollution and invasive species. Although at the time wildlife populations appeared to be stable or increasing, it was noted that assessment criteria were needed to determine whether wildlife populations were impaired; and therefore further assessment was required.

The Stage 2 RAP report (2002) outlines a strategy to remediate the impaired beneficial uses identified in the St. Marys River AOC. It details 57 recommended actions to restore beneficial uses on both the Canadian and U.S. side of the AOC. The specific recommended actions for the *Degradation of Fish and Wildlife Populations* BUI on the Canadian side of the AOC are:

Fish Populations

Action FF-7: Develop a 10-year fisheries assessment program for the river Action FF-8: Continued support for Sea Lamprey control efforts Action FFM-3: The Fish Harvest Survey

Wildlife populations

Action FFM-2: Continued support for the Marsh Monitoring Program Action FFM-5: Complete an assessment of Common and Black Tern populations for the area

Action FFM-6: Analyze contaminant levels in eggs from Herring Gull, Black Tern, and Common Tern nests in the AOC

Action FFM-8: Assessments of Herring Gulls, Black Terns and Common Terns within the AOC

1.3 Additional Actions

In addition to the actions outlined in the Stage 2 RAP report, other actions have been implemented that have helped to improve conditions for fish and wildlife populations. These actions are outlined below and are documented in the St. Marys River Interactive Map². The online map highlights projects and progress made since the St. Marys River was first designated as an Area of Concern under the Great Lakes Water Quality Agreement.

• Fort Creek Clean-up:

 In 2009, the Sault Ste. Marie Region Conservation Authority (SSMRCA) completed improvements to Fort Creek south of Second Line by planting trees, improving flow through sediment and garbage removal, and by establishing a pond to attract wildlife.

² The interactive map can be accessed here: <u>http://bpac.algomau.ca/?page_id=76</u>

- Bar River Restoration Project:
 - The community-driven Bar River Habitat Project improved stream bank and riparian zone conditions through the planting of trees and the restriction of livestock to the St. Marys River. The project was formed in 1999 by a local chapter of Scouts Canada as well as teachers and students from Central Algoma Secondary School, who worked with Environment and Climate Change Canada and the Ministry of Natural Resources and Forestry to plant spruce, cedar and hardwood species along three areas in the upper reaches of the Bar River while reinforcing banks against erosion (CWS, 2013).
- Improvements to Water Quality:
 - The East End Waste Water Treatment Plant was upgraded in 2006, which has improved water quality by reducing suspended solids, phosphorus levels, biological oxygen demand, and significantly reducing nitrogen and ammonia levels in water discharged. These upgrades allowed for secondary treatment of water, including Ontario's first biological nutrient removal and ultraviolet light disinfection, which prevents chlorine from being released into the St. Marys River.
 - A study conducted by Algoma University from 2013-2015 found no large algal blooms or high concentrations of microscopic algae. There was also no evidence of unnatural colour, odour, or turbidity indicating resolved problems associated with degraded aesthetics (Ginou, 2016).
 - Starting the early 1980s, Algoma Steel has worked to improve its operations and to reduce the impact it has on the quality of the river and the surrounding environment. Algoma Steel has worked to improve sources of water pollution in many ways such as the creation of a main water filtration plant, decommissioning of settling ponds, installation of a biological treatment facility and toxicity control system, as well as improving overall water re-use efficiency.
 - Once a source of historical pollution for the river, the St. Marys Paper Mill was decommissioned in 2012. The site was dismantled, removing it as a pollution source for the river.

1.4 Delisting Criteria

In 2015, the suite of BUI delisting criteria were finalized for the Canadian side of the AOC, including that for the *Degradation of Fish and Wildlife Populations* BUI to account for local circumstances, link to relevant regulations or guidelines, and to be specific, measurable, achievable, relevant, and time-oriented ("SMART"). The delisting criteria for fish component of the *Degradation of Fish and Wildlife Populations* states that the BUI will no longer be impaired when:

The overall fish community health within the AOC is comparable to that of a suitable reference site, as assessed using an index of biotic

integrity through a minimum of two consecutive studies.

At the time, it was decided that other information/data sources would be used to supplement the IBI approach. Specifically, results from the binational St. Marys River Fisheries Task Group's fish surveys would be used to confirm the status of "managed" fish populations.

In the Stage 1 and 2 RAP reports, the wildlife portion of the BUI was never deemed "Impaired" but instead "Required Further Assessment" because assessment criteria were absent to help determine the status. Delisting criteria are only appropriate for when a beneficial use is impaired, but the assessment of the wildlife component follows established methods for evaluating wildlife health and ecosystem conditions so as to conclude if there is impairment. These studies are summarized in section 3.2 and included in full in Appendix 3a-c.

2.0 Degradation of Fish Populations

2.1 Summary of Impairment

At the time of the Stage 1 RAP report in the early 1990s, reductions in fish populations in the St. Marys River was suspected due to alteration of fish spawning habitat, overfishing, exotic species and decreases in benthos populations. In 2002, the Stage 2 RAP report recommended looking at fish population dynamics in order to assess whether local environmental conditions support healthy, self-sustaining communities of fish. This was not to be limited to population levels, but overall fish community health. Similar to concerns raised during in the Stage 1 RAP report, the Stage 2 RAP report also noted that native fish populations were being stressed by habitat alteration, over-fishing, pollution, and invasive species. Concerns with fish body burdens were also identified in the Stage 2 RAP report linked to polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), and resin acids. Since the impact of these contaminants on fish are addressed in other BUIs, namely *Restrictions on Fish Consumption* and *Fish Tumours and Other Deformities*, the focus of this BUI was kept to fish population dynamics when the delisting criteria were revised in 2015.

2.2 Stage 2 Recommended Remedial Actions Specific to Fish Population Dynamics

2.2.1 Develop a 10 year fisheries assessment program for the river (Action FF-7)

In 2002, the St. Marys River Fisheries Task Group completed a Fisheries Assessment Plan. This plan provides a standardized approach for regular assessment of the river's fishery and aquatic resources. Included are approaches for fish community assessment, fish harvest estimates and reporting, lower trophic level monitoring, and habitat mapping and data collection. The plan serves as a mutual and coordinated approach to assessment and ensures coordination of management actions for the St. Marys River Fishery through the Great Lakes Fishery Commission's Lake Huron Committee (Gerbhardt et al. 2002).

As part of the Fisheries Assessment Plan, the St. Marys Fishery Task Group has ongoing fish community surveys that occur every 3-5 years. These surveys provide managers with the necessary information and data to make informed decisions about the management of fish communities within the river. Fish community objectives for the St. Marys River are currently being developed (Godby et al. 2019).

This action is considered addressed because fisheries resource management is led by the binational St. Marys River Fishery Task Group under the Great Lakes Fisheries Commission, which functions independently of the RAP.

2.2.2 Continued support for Sea Lamprey control efforts (Action FF-8)

Although the production of Sea Lamprey from the St. Marys River has been a serious problem for salmonine management in the Great Lakes, the overall presence of migrating adults in the river is not believed to be a significant factor affecting the rest of the river's fish community (Gerbhardt et al. 2002).

Overall, Sea Lamprey wounding rates remain low in the St. Marys River with 0.2% of the total fish collected during a 2017 fish community survey exhibiting a Sea Lamprey wound. Sea Lamprey management within the St. Marys River encompasses both the assessment of adult and larval populations within the river as well as targeted treatment of larval populations. Sea Lamprey assessment and control within the St. Marys River is an ongoing priority operation of DFO's Sea Lamprey Control Centre in Sault Ste. Marie, Ontario (O'Connor et al. 2019).

This action is considered addressed because aquatic invasive species are a lake- and basin-wide management issue, and not a local AOC-specific issue. And as noted above, there are dedicated programs led by the Sea Lamprey Control Centre under DFO, which functions independently of the RAP.

2.2.3 The Fish Harvest Survey (Action FFM-3)

In order to quantify the sport fisheries, the Michigan Department of Natural resources (MDNR) and the Ontario Ministry of Natural Resources and Forestry (MNRF) undertook an open water access creel survey of sport anglers (Godby et al 2019). This was accompanied by a fish community gillnet survey of the river by the St. Marys River Fisheries Task Group and its member agencies. The general conclusion is that managed fish populations are stable or increasing, with key findings from Godby et al. that include:

- Northern Pike harvest and harvest rate are still lower than they were historically but are trending upwards.
- Smallmouth Bass are plentiful in the river and angler harvest and harvest rate remain substantially higher than in 1999.
- Walleye continue to be an important sport fish in the St. Marys River. Overall harvest and harvest rate of Walleye have declined since 2007, but both are higher than they were in 1999.
- Yellow Perch support a popular fishery in the river. Although overall harvest of Yellow Perch is down in 2017 compared to previous years, harvest rate has remained stable over the survey series (Godby et al. 2019).

And the key findings from the 2017 fish community gillnet survey (O'Connor et al. 2019) include:

- Walleye catch per unit effort (CPUE) has remained stable from 2006-2017, after increasing from a survey low in 2002. Overall in 2017, Walleye appear to be well distributed throughout the St. Marys River.
- In 2017, Northern Pike CPUE continued to improve, reversing the downward trend that began in 2002. Although the CPUE has not returned to pre-2002 levels, it has rebounded to more than double that of its lowest point, measured in 2009.
- Yellow Perch abundance decreased in 2017 but the mean CPUE is just below the survey average and the overall trend remains positive since the first survey in 1975.
- Cisco appear to be less abundant in recent years although the overall trends in gillnet CPUE were not significantly different (O'Connor et al. 2019).

2.2.4 Assessment of Fish Populations using Index of Biotic Integrity

In 2009, DFO (Pratt & O'Connor, 2011) compared the status of communities from four distinct areas of the river including the upper river above the compensating works, the main river, Lake George and the lower river. They provided an overall assessment of the fish community using an index of biotic integrity (IBI) approach. An IBI is a scientific tool used to identify and classify faunal communities. Several matrices are used to provide an overall assessment of fish community including mortality rates, age-class structure, survival to spawning age, reproductive success, total biomass, productivity, richness, assemblage, and abundance. The study concluded that the overall health of the St. Marys River fish community compared favourably with healthy reference sites from Lake Huron.

In 2014, DFO (O'Connor & Pratt, 2017) initiated a follow-up 2-year survey that again used the IBI approach called for in the BUI delisting criteria, and relied on the Mississagi River near the North Channel as the reference site. Key findings from this study include:

 Using the Great Lakes scoring system, the overall IBI score for the St. Marys River AOC is 60.3 which borders between fair (>41 -60) and good (>61-80).

- A small decline in IBI was observed between 2006-2009 and 2014-2015. The lower IBI scores can be attributed to more generalist species, lower percent piscivourous community, and lower biomass. However, these are expected and reflective of a cold-water, lower productivity, riverine environment such as the St. Marys River.
- The St. Marys River maintains a native fish community that is complex, diverse, and healthy.

Together, these studies directly answer the BUI delisting criteria and present a case for not impaired status, because: fish community health was assessed using an index of biotic integrity approach and in these two consecutive studies the health of the AOC fish community was found to be comparable to suitable reference sites.

2.3 Conclusions for Fish Populations

The delisting criteria for the *Degradation of Fish and Wildlife Populations* states that the BUI will no longer be impaired when *the overall fish community health within the AOC is comparable to that of a suitable reference site, as assessed using an index of biotic integrity through a minimum of two consecutive studies*. The two fish community studies by DFO that use an IBI approach show AOC fish populations "compare relatively well" to reference areas and the IBI score for the St. Marys River fish community borders between fair and good. In addition, supplemental assessments and work completed under Actions FF-7, FF-8, and FFM-3 (described above) all show that the St. Marys River is home to a relatively healthy fish community that is complex, diverse and dominated by native species, and has managed fish populations that are stable or increasing. It is therefore recommended a change in status from Impaired to Not Impaired.

3.0 Degradation of Wildlife Populations

3.1 Summary of Impairment

In the Stage RAP reports, the wildlife portion of the *Degradation of Fish and Wildlife Populations* BUI was designated as "Requires Further Assessment". Although wildlife populations appeared to be stable or increasing, it was noted assessment criteria were needed to evaluate their status. In the early 1990s, Common Tern (*Sterna hirundo*) populations were declining while Ring-billed Gull (*Larus delawarensis*) numbers were increasing. As stated in the Stage 1 RAP report, habitat loss and alteration were causing a reduction in nesting habitat and the larger, earlier nesting Ring-billed Gulls were displacing Common Terns along with other smaller species from nesting sites. In addition, both the Stage 1 and 2 RAP reports noted chemical contaminants within the tissues of waterfowl, particularly mercury and polychlorinated biphenyls (PCBs).

In response to this, the Stage 2 RAP report recommended an assessment of wildlife habitat conditions (Action FFM-2), an assessment of Common Tern and Black Tern (*Chlidonias niger*) populations for the entire St. Marys River (Action FFM-5) and a reproductive assessment of

Herring Gulls and Black and Common Terns (Action FFM-8, which fell under the *Bird and Animal Deformities or Reproductive Problems* BUI). The Stage 2 RAP report also recommended a full analysis of contaminant levels in the eggs of Herring Gulls and Black and Common Terns within the AOC (Action FFM-6). These actions and the associated studies are discussed below, because they serve to evaluate the health of wildlife populations in the AOC.

3.2 Stage 2 Recommended Remedial Actions Specific to Wildlife Populations

3.2.1 Continued Support for the Marsh Monitoring Program (Action FFM-2)

In 2011, the Canadian RAP agencies and Binational Public Advisory Council (BPAC) discussed the need for a comprehensive assessment of wildlife habitat conditions (specifically coastal wetlands) and associated wildlife populations, and to evaluate the degree of impairment, if any. A multi-year study by Environment and Climate Change Canada's (ECCC) Canadian Wildlife Service (Darwin, 2016) started in 2012, and in August 2016, the 5-year monitoring effort was completed. The study assessed baseline wildlife habitat conditions and evaluated coastal wetland water quality, and breeding bird, amphibian, aquatic macroinvertebrate and submerged vegetation communities within the AOC, concluding the wildlife habitat and populations are not impaired. The full report is provided in Appendix 3a, but key findings included:

- Water quality within the AOC's coastal wetlands is comparable to non-AOC reference sites; suggesting overall water quality can be considered not impaired.
 Algoma University's water quality survey (2013-15) supports this (Ginou 2016).
- Breeding marsh birds in the AOC are in relatively undisturbed condition, and sites inside and outside the AOC are in comparable condition; suggesting there is no impairment.
- There is no clear response disturbance within the amphibian and aquatic macroinvertebrate communities, suggesting those populations are not impaired.
- There are some differences between submerged aquatic vegetation communities in the AOC versus non-AOC reference sites, but the overall area is not impaired for this community type.

3.2.2 Complete an Assessment of Common and Black Tern Populations for the Area (Action FFM-5)

The Canadian RAP agencies and BPAC discussed the need to assess the populations of Common Terns and Black Terns within the AOC as per the recommendation in the Stage

2 RAP report. In 2014, ECCC completed the population assessment based on nest count surveys conducted between 2010 and 2013, which was supplemented with historical breeding data from 1978-80, 1989, 1999-00, and 2007-08 (Hughes et al. 2014b, see Appendix 3b). Population trends for colonial waterbirds breeding on the North Channel of Lake Huron were included to provide a broader context of trends in diversity and abundance within the AOC. The study concluded Commons Terns and Black Terns are breeding within the AOC, and that there is no evidence that breeding status within the AOC differs from that outside of AOC. Also, nesting and population patterns are influenced by life history strategies of the species and factors that are regional or basinwide in nature, and not specific to influences within the AOC. The full report is provided in Appendix 3b, but key findings include:

- Despite natural fluctuations, Common Tern populations have had no significant change over the past 30 years, with 70 nests found in 1978-80 versus 78 in 2007-08.
- Evidence from nest count surveys between 1980 and 2013 suggest trends in populations of nesting Common Terns in the AOC are likely related to factors consistent with the life history strategies of the species, and are not specific to influences in the AOC.
- Black Terns seem to be limited on the Ontario side of the river, with a breeding colony found only at Echo Bay. It is not possible to report on temporal trends in abundance of Black Terns in the AOC due to limited data. However, based on the data that is available, there is no evidence to suggest that breeding status of Black Terns nesting within the AOC differs from those nesting at sites downstream in the North Channel.
- The relative low population of nesting Black Terns is likely reflective of low densities reported throughout the region, Ontario and the Great Lakes basin and not due to AOC-specific conditions.
- Overall, the total number of colonial waterbird nests on the Ontario side of the St.
 Marys River increased by almost 23% between 1999 and 2008; largely driven by dramatic increases in Ring-billed Gulls.

3.2.3 Analyze Contaminant Levels in Eggs from Herring Gull, Black Tern, and Common Tern Nests in the AOC (Action FFM-6) and complete Reproductive Assessments of Herring Gulls, Black Terns and Common Terns within the AOC (Action FFM-8)

In parallel to the above-mentioned assessment of Common Tern and Black Tern populations within the AOC, in 2014, ECCC also completed a three-year Common Tern and Herring Gull study based on fieldwork and laboratory analysis to assess deformities, reproductive health, and chemical contamination in eggs of these indicator species (under the *Bird and Animal Deformities or Reproductive Problems BUI*). The study report (Hughes et al. 2014a) concludes that there is no evidence of impaired reproduction or deformities in colonial waterbirds attributable to local contamination effects within the AOC, and the reproductive success for birds studied within the AOC is similar to that from outside the AOC. The full report is provided in Appendix 3c, but the key findings include:

- Contaminant levels are low overall and not sufficiently elevated to have an adverse impact on reproductive success and development (which is an impact related to polychlorinated biphenyls (PCBs) and other organochlorines, dioxins/furans, heavy metals like mercury, and polybrominated diphenyl ethers (PBDEs)).
- No physical deformities have been detected within gull or tern chicks (the original issue identified by the RAP). There is a low incidence of embryonic deformities that cannot be linked to contaminant burdens or geographical area (i.e., there is no significant difference between AOC and non-AOC bird colonies); and the reproductive success for Herring Gulls within the AOC is high, and that of Common Terns is similar to the rest of the region.

The report supported the official re-designation of the *Bird and Animal Deformities or Reproductive Problems* BUI from "Requires Further Assessment", as identified under the Stage 2 RAP report (2002), to "Not Impaired" in January 2016 via a notification letter to the International Joint Commission. This is consistent with the same change in BUI status by the United States Environmental Protection Agency in 2014.

3.3 Conclusions for Wildlife Populations

All recommended remedial and monitoring actions pertaining to the wildlife component of the *Degradation of Fish and Wildlife* Populations BUI have been completed. The assessments and work completed under Actions FFM-2, FFM-5, and FFM-8 described above show that the overall health of wildlife in the St. Marys River is comparable to non-AOC areas. It is therefore recommended a change in status from Requires Further Assessment to Not Impaired.

4.0 Status of the Degradation of Fish and Wildlife Populations BUI in Sault Ste. Marie, Michigan

The *Degradation of Fish and Wildlife Populations* BUI removal on the U.S. side of the St. Marys River AOC became final on September 23, 2019. Subsequent to engaging BPAC on findings relating to both the *Degradation of Fish and Wildlife Populations* and *Loss of Fish and Wildlife Habitat BUIs*, the U.S. Environmental Protection Agency re-designated both BUIs to "Not Impaired" on the U.S. side of the AOC upon the recommendation of the Michigan Department of Environment, Great Lakes, and Energy. Michigan's delisting criteria for both BUIs state:

"The two fish and wildlife BUIs will be considered restored in the Michigan portion of the St. Marys River AOC upon completion of the Little Rapids project at Sugar Island, which would restore approximately 50 to 70 acres of fish and wildlife habitat." The Little Rapids restoration project was carried out between 2015 and 2016. More than 50 acres were positively impacted by the change in flow regime resulting from the removal of a causeway and construction of a bridge with 600 feet of free-flowing water beneath it. The project successfully achieved the target of restored habitat area acreage and the two BUIs were deemed "Not Impaired" in 2019 (Riley, 2019).

5.0 Conclusions and Recommendations Regarding Re-designation

The fish component of the *Degradation of Fish and Wildlife Populations* BUI was identified as "Impaired" in both the Stage 1 and Stage 2 RAP reports. The delisting criteria states that the BUI will no longer be impaired when: *the overall fish community health within the AOC is comparable to that of a suitable reference site, as assessed using an index of biotic integrity through a minimum of two consecutive studies*. Fisheries and Oceans Canada completed an assessment of the AOC fish community using the index of biotic integrity (IBI) approach in two separate studies, encompassing fieldwork and data analysis in 2006-08 and 2014-15. Both studies conclude the St. Marys River is home to a relatively healthy fish community that is complex, diverse, and dominated by native species. These results were shared and discussed with BPAC in October 2017. Other information and data sources have been used to supplement the IBI approach. Specifically, results from the binational St. Marys River Fisheries Task Group's fish community gillnet survey and the river-wide Creel survey was used to confirm the status of "managed" fish populations and show they are stable or increasing.

The wildlife component of the *Degradation of Fish and Wildlife Populations* BUI was designated as "Requires Further Assessment" in the Stage 2 RAP report, and wildlife populations have since been assessed through the completion of Actions FFM-2, FFM-5, FFM-6, and FFM-8 and three studies as discussed above. Wildlife populations are found to be in good condition.

Based on the lines-of-evidence presented in this report, it is recommended that the *Degradation of Fish and Wildlife Populations* BUI for the Canadian side of the St. Marys River AOC be re-designated to "Not Impaired" since the BUI delisting criteria, as endorsed by BPAC, has been fulfilled.

6.0 References

Canadian Wildlife Service (CWS). (2013). St. Marys River Area of Concern: Bar River Habitat Project: 2013 Follow-up. Environment and Climate Change Canada. Available here: <u>http://bpac.algomau.ca/wp-</u> <u>content/uploads/2018/01/BarRiverHabitatProject-Nov-2013.pdf</u>

Darwin, A. (2016). St. Marys River Area of Concern: Coastal Wetland Habitat Assessment Report. Environment and Climate Change Canada – Canadian Wildlife Service. See Appendix 3a.

Derickx, L. (2018). St. Marys River Area of Concern (Canadian Section); Beneficial Use Impairment Status Report: Degradation of Fish and Wildlife Populations (Wildlife Populations) and Loss of Fish and Wildlife Habitat (Wildlife Habitat). 26 pp. Available here: <u>http://bpac.algomau.ca/wp-</u> <u>content/uploads/2019/06/St-Marys-River-AOC-Wildlife-Redesignation-Report-2018.pdf</u>

Gebhardt, K.D., Fielder, D., Greenwood, S., Robbins, H., Sutton, T. [Editors]. (2002). St. Marys River Fisheries Assessment Plan. Great Lakes Fisheries Commission, Special Report. Ann Arbor. 46 pp. Available here: <u>http://bpac.algomau.ca/wp-content/uploads/2015/09/St.-Marys-River-Fisheries-Assessment-Plan-2002.pdf</u>

Ginou, C. (2016). Water Quality Monitoring and Analysis: An Investigation of the Eutrophication and Undesirable Algae, and the Degradation of Aesthetics Beneficial Use Impairments in the Canadian St. Marys River Area of Concern (2013-2015). Algoma University Remedial Action Plan Office. 94 pp. Available here: <u>http://bpac.algomau.ca/wp-content/uploads/2016/09/Water-Quality-Technical-Report-</u> 2013-15 Aug-2016.pdf

Godby, N., Claramunt, T., Fielder, D.G., Chong, S., Bowen, A., Morrow, E. (2019). A Synthesis of Sport Fishing Activity in the St. Marys River. 17 pp. See Appendix 2a.

Hughes, K.D., Crump, D., Williams, K., and Martin, P.A. (2014a). Assessment of the Wildlife Reproduction and Deformities Beneficial Use Impairment in the St. Marys River Area of Concern (Ontario). Environment and Climate Change Canada – Ecotoxicology and Wildlife Health Division. See Appendix 3c.

Hughes, K.D., Moore, D.J., and Martin P.A. (2014b). An Assessment of Breeding Populations of Common Terns and Black Terns in the St. Marys River Area of Concern (Ontario). Environment and Climate Change Canada – Ecotoxicology and Wildlife Health Division. See Appendix 3b.

O'Connor, L., Aikens, R., Bowen, A., Chong, S., Fielder, D.G., Godby, N. (2019). Population Dynamics of the St. Marys River Fish Community 1975-2017. See Appendix 2b.

Riley, J. (2019). Removal Recommendation: Degradation of Fish and Wildlife Populations and Loss of Fish and Wildlife Habitat Beneficial Use Impairments, St. Marys River Area of Concern. Michigan Ddpartment of Environment, Great Lakes, and Energy. Available here: <u>https://www.michigan.gov/documents/egle/egle-wrd-aoc-remrec-stmarys_661909_7.pdf</u>

St. Marys River Remedial Action Plan: Stage 1 Report (RAP 1). (1992). St. Marys River Area of Concern Environmental Conditions and Problem Definitions. Ontario Ministry of the Environment, Conservation and Parks and Michigan Department of Natural Resources. Available here: <u>http://bpac.algomau.ca/wp-content/uploads/2015/09/The-St.-Marys-River-Area-of-Concern-Stage-1-Remedial-Action-Plan-Report-on-Environmental-Conditions-and-Problem-Definitions-1992.pdf</u>

St. Marys River Remedial Action Plan: Stage 2 Report (RAP 2). (2002). St. Marys River Area of Concern Remedial Strategies for Ecosystem Restoration. Environment Canada, United States Environmental Protection Agency. Ontario Ministry of the Environment, Conservation and Parks, and Michigan Department of Environmental Quality. Available here: <u>http://bpac.algomau.ca/wp-</u> <u>content/uploads/2015/09/The-St.-Marys-River-Area-of-Concern-Stage-2-Remedial-Action-Plan-Reporton-Remedial-Strategies-for-Ecosystem-Restoration-2002.pdf</u> Appendix 1a: An assessment of the nearshore fish community of the St. Marys River, Ontario (O'Connor & Pratt, 2017)

An Assessment of the Nearshore Fish Community of the St. Marys River, Ontario

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2017

Canadian Manuscript Report of Fisheries and Aquatic Sciences

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An Assessment of the Nearshore Fish Community of the St. Marys River, Ontario

by

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ABSTRACT

The health of the nearshore fish community in the St. Marys River, a binational Area of Concern, was assessed via night boat electrofishing over six sampling seasons, in 2006-2009, and again in 2014-15. Our objectives were to (1) determine if changes had occurred in the fish community from the previous five years; (2) assess and contrast the status of the fish community within each of the four sampling areas; (3) compare the fish community of the St. Marys River with that of the Mississagi River, a relatively undisturbed river used as a reference site; and (4) assess the current overall health of the fish community in the St. Marys River. The St. Marys River contains a diverse and complex native fish community, with consistent differences in fish communities observed among broad habitat types. There have been recently declines in the fish community despite being present in low numbers. The overall health of the St. Marys River fish community is considered fair, and is poorer than relatively un-impacted sites assessed from Lake Huron primarily due to the low number of piscivores captured and low overall biomass.

INTRODUCTION

The St. Marys River, the connecting channel between Lake Superior and Lake Huron, is a 112 km long river bounded by Canada and the United States that encompasses a wide diversity of fish habitats (Bray 1996) and fish species (Duffy et al. 1987). Duffy et al. (1987) identified 75 fishes from 22 families. The greatest physical changes to the river occurred roughly between 1850 and 1930, which reduced the area of the St. Marys rapids to less than 20% of its original size through the construction of shipping locks and the development of multiple hydroelectric generating facilities (Duffy et al. 1987, Bray 1996, Ripley et al. 2011). Additional changes have included the building of a compensating works to control the outflow of Lake Superior, and ongoing dredging to support the commercial shipping industry.

The river was declared an Area of Concern (AOC) in 1987 by the International Joint Commission (IJC) as an amendment to the Great Lakes Water Quality Agreement (GLWQA 1987). The fish community was described as impaired due to the abundance of sea lamprey (*Petromyzon marinus*), the supporting of recreational and commercial fisheries through the use of stocked non-native salmonids, declining native species due to over-harvesting, the loss of fish habitat within the river, and concerns over aquatic invasive species (OMOE and MDNR 1992). As part of the AOC rehabilitation process, a Remedial Action Plan was developed for the St. Marys River which included actions to rehabilitate the fish community. The health of recreationally valuable sport fishes are determined tri-annually (since 1976) by the bi-national St. Marys River Fisheries Task Group using data from a graded mesh gillnet survey (Schaeffer et al. 2011). A baseline assessment of the broader fish community was undertaken by Liston et al. (1980; 1983) and Liston and McNabb (1986) using bottom trawls, and these data were used in conjunction with data from a recent standardized boat electrofishing survey using data from 2006-2008 to assess the health of the nearshore fish community (Pratt and O'Connor 2011).

A further update of the status of the Degradation of Fish and Wildlife Populations Beneficial Use Impairment was requested five years after the completion of the initial nearshore fish community surveys of Pratt and O'Connor (2011). Four distinct areas of the St. Marys River were sampled to: (1) determine if changes had occurred in the fish community from the previous five years; (2) assess and contrast the status of the fish community within each of the four sampling areas; (3) compare the fish community of the St. Marys River with that of the Mississagi River, a relatively undisturbed river used as a reference site for the St. Marys River fish community; and (4) assess the current overall health of the fish community in the St. Marys River. All objectives were addressed using an index of biotic integrity approach (Minns et al. 1994).

METHODS

STUDY SITE

The St. Marys River is the connecting channel between lakes Superior and Huron (Figure 1a). It is bounded on the upper end by Whitefish Bay (46°31'N; 84°36'W) and at the southeastern end below St. Joseph's Island (46°12'N; 83°40'W). Historically, the rapids at Sault Ste. Marie were the main feature of this river. However, modifications to the rapids area to provide for shipping through the construction of the locks, the construction of compensation gates to regulate the river

flow and the development hydroelectric generating facilities drastically reduced the rapids to about 7% of its original flow (Duffy et al. 1987; Edsall and Gannon 1993). The four survey sections of the river outlined in Pratt and O'Connor (2011) corresponded with the three distinct areas of the river described in Duffy et al. (1987); the upper river, the rapids, and the lower river. We sampled the Canadian nearshore habitats of the St. Marys River, with the exception of the main rapids area where the current was too strong and the river too shallow to sample safely in our 4.4 m electrofishing boat. In 2014 and 2015, we also sampled the lower reach of the Mississagi River, located approximately 50 km east of the outflow of the St. Marys River, as a non-AOC reference site (Figure 1b). While the size, flow and habitat complexity available in the St. Marys River means that there are no perfect candidate rivers for comparison, the Mississagi River is largest river on the north shore of Lake Huron without an AOC designation and was selected a reference site for this study. Both study sites are described in detail below and the sampling dates, the number of transects completed in each section and habitat descriptions are found in Table 1.

SITE DESCRIPTIONS

Upper river

The transect locations in the upper river were approximately 5 km above the compensating gates, in Marks Bay and Leigh Bay (46°29'N; 84°27'W). There is little structural habitat in the area, with predominantly sand substrates. The majority of the available structural habitat is provided by sunken logs, remaining from the days of holding log booms in the western edge of Marks Bay. Sparse aquatic vegetation (primarily quillwort (*Isoetes riparia*)) provided the only macrophyte cover. Water temperatures were cool in this section of the river, consistent with its proximity to Lake Superior (Table 1).

Main river

The main river extended from below the rapids (46° 30'N 84° 20' W) to Bells Point (46° 32'N 84° 13' W), a distance of approximately 12 km. This area of the river contained the greatest variation of habitat and the most flow of any of the sampled transects. The habitat below the main rapids and to Topsail Island is mainly cobble and boulder with some sand, and from east of Topsail Island to Bells Point the substrate is silt/sand and macrophytes (mainly *Potamogeton* spp. and quillwort). The area represented the open water main stem and embayment fish habitat described by Duffy et al. (1987). Water temperatures were coldest most years in this section of the river.

Lake George

Lake George is large, shallow embayment with low flow and sandy substrate area with little in the way of macrophytes or structure for fish habitat. This area represented the sand beach fish habitat described by Duffy et al. (1987). Transects in this section were concentrated in the Bar River mouth area (46° 26'N 84° 06' W). Water temperatures were warmer and conductivity higher in this section of the river.

Lower river

The lower river was sampled along of the northwest edge of St. Joseph's Island, in the Richards Landing area (46° 17'N 84° 02' W). Bottom substrate varied, ranging from sand and silt to

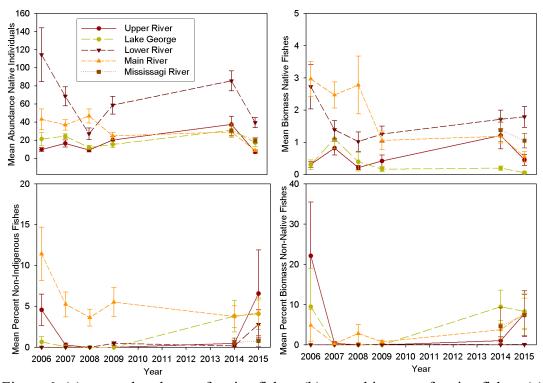


Figure 6: (a) mean abundance of native fishes, (b) mean biomass of native fishes, (c) mean percent non-native fish abundance, (d) mean percent non-native biomass by year for each of the four study areas in the St. Marys River and the Mississagi River. Data for 2006-2008 from Pratt and O'Connor (2011).

cobble and a wide range of macrophyte densities (sparse to dense). This area represented the emergent wetlands and protected reaches described by Duffy et al. (1987). Water temperatures and conductivity were similar to those of Lake George in this section of the river, and dissolved oxygen concentrations were lower than the upper and main river locations.

Mississagi River

The lower section of the Mississagi River (46° 11'N 83° 01 W) covers a delta area approximately 7 km in length. Bottom substrate in this stretch of river is similar to the Lake George and the lower St. Marys River areas; mainly sand, silt and sparse to moderate macrophytes. Water temperatures were warmer in the Mississagi River, likely as a consequence of later temporal sampling, and dissolved oxygen concentrations in the Mississagi River were the lowest of any location fished.

SAMPLING METHODS

The nearshore fish community was sampled using a boat electrofisher. The technique for sampling the nearshore community was developed Fisheries and Oceans Canada for use in the Laurentian Great Lakes (Valere 1996, Brousseau et al. 2005) and described in Pratt and O'Connor (2011). Surveys were conducted in late spring, with the earliest survey beginning May 25th and the latest June 20th in the St. Marys River (Table 1). Surveys in the Mississagi River were conducted after those in the St. Marys River, starting as early as June 26th and ending as late as July 19th (Table 1). The same sites were visited during each sampling period where possible; changing water depths or turbidity levels periodically resulted a slight shift in transect location.

Detailed descriptions of the sampling procedure are available (Valere 1996, Brousseau et al. 2005, Pratt and O'Connor 2011), but are further summarized here. In 2006 – 2009, 100 m transects were laid out prior to dusk using floats with reflective tape and marked with a GPS. Transects were placed along the 1.5 m depth contour. The physical habitat was sampled prior to dark; at mid-transect, dissolved oxygen (mg/L), conductivity (µS/cm) and water temperature were taken, and bottom substrate was sampled using an Ekman dredge at each transect marker. Substrate was classified as silt, sand or gravel texturally, while areas of larger substrate, cobble or boulder, were classified visually. Organic and woody debris were also identified as well as the dominant (>50%), sub-dominant (10 to 50%), and trace (<10%) amounts for each component. Macrophyte abundance was determined visually and assigned to one of 4 categories: none (0%), sparse (1 to 19% cover), moderate (20 to 70% cover) or dense (>70% cover). The dominant macrophytes were identified to genus. For the 2014 and 2015 surveys, a change occurred and the 100 m transects were laid out using GPS coordinates along the 1.5 m contour and habitat data was collected after the completion of the transect. Dredge sampling was no longer used to collect bottom substrate for classification purposes; bottom samples were taken via net probing or anchor drops, and followed the previous classification system. Collected water sampling parameters remained the same and were assessed mid-transect. We found we were able to sample significantly more transects over the same time period by adjusting the transect marking and habitat collection techniques as the crew did not start until dark, which allowed for more time to be devoted to the electrofishing survey effort.

Electrofishing surveys were conducted in a Smith-Root SR -14H electrofishing boat (4.4 m long, 2.0 m wide). Electrofishing was conducted parallel to shore along the 100 m transect after dark. The target electrofishing time was 300 sec per transect. All fish were targeted for collection, brought on board and held in an aerated live well until the transect was completed. All fish were identified to species by the crew and 20 of each species were measured (± 1 mm) and weighed (± 1 g) in the field. If over 20 individuals of a given species were collected, the remainder was counted and batch weighed. Fish were then returned to the water, close to their location of capture. For species that could not be identified in the field, a subsample were preserved in 10% formalin and returned to the lab for individual identification and length (± 1 mm) and weight (± 0.01 g) measures.

In 2014 and 2015 we also added an additional fish collection sampling method, the Herzog-Armadillo (Modified Missouri) trawl net (Herzog et al. 2009). We added the trawl net to determine if bottom fishes beyond the 1.5 m depth contour were not being sampled due to depth limitations of the boat electrofisher. The net (2.4 m x 0.6 m) was towed along 100 m transects in generally the same areas where boat electrofishing occurred, but beyond the 2 m depth contour. All trawling surveys were conducted during the day. Captured fishes were processed the same as those collected during the electrofishing surveys, with all fishes identified to species, 20 individuals of each species measured for length (± 1 mm) and weight (± 1 g), and the remainder counted and bulk weighed.

INDEX OF BIOTIC INTEGRITY (IBI) CALCULATION

All collected habitat and fish data were entered into a database developed specifically for Great Lakes nearshore fish communities (Moore et al. 1998). The database was developed for the comparison of the nearshore fish community, species richness and biomass with the habitat attributes of the surveyed transects. The database also calculates a Great Lakes nearshore index of biotic integrity (IBI) (Minns et al. 1994). Minns et al. (1994) developed the IBI specifically for the littoral habitats in the Great Lakes to provide a quantitative measure of impairment within an ecosystem and restoration measures for the fish community. The IBI uses 12 separate metrics based on the diversity and trophic metrics of the fish community to provide a score from 0 to 100 to determine the condition of the fish community, based on four broad classes of factors: non-native species, water quality, physical habitat, and piscivore abundance (Minns et al. 1994). Positive metrics include species richness (native, centrarchid, intolerant (intolerant of high turbidity) and native cyprinids), trophic structure (percent piscivore, specialist biomass), and abundance and condition (number and biomass of native species; Table 2). Negative metrics include non-native species (Randall and Minns 2002, Brousseau et al. 2005).

We also calculated a habitat-productivity index (HPI) (Randall and Minns 2002). The habitat productivity index Randall and Minns (2002). developed is a measure of habitat productive capacity for a given fish assemblage. For each sample, HPI was calculated as the product of fish biomass and the production to biomass ratio (P/B) for all species captured from a given transect. This index was used as an estimate of fish production. As detailed in Pratt and O'Connor (2011) for the St. Marys River, our total fish biomass from each transect was converted to kg/ha by multiplying the biomass by 40, as we estimated our electrofishing boat was effective over a 2.5

m strip along the 100 m transect. We also assumed our capture efficiency was 100% (Pratt and O'Connor 2011). Together, the IBI and the HPI provide indices of fish production and diversity (Randall and Minns 2002) and were used to examine the nearshore fish community in the St. Marys River.

DATA ANALYSIS

All data analysis completed using SPSS 18 (SPSS 18). One-way analysis of variance (ANOVA) was used to test macrophyte density among years at the five individual fishing locations (lower river, Lake George, main river, upper river, and Mississagi River), followed by Tukey highly significant difference (HSD) post-hoc analyses to separate locations after a significant ANOVA was identified. IBI, HPI and IBI metrics were compared among years at the five locations with a one-way ANOVA, with Tukey HSD post-hoc comparisons completed as required. Two-way ANOVAs were used to contrast among year and site comparisons, with Tukey HSD post-hoc comparisons completed when required; when the site*year interactions were significant, MANOVAs were used to compare the individual factors, with Tukey HSD post-hoc analysis used to separate factors after significant tests were identified.

RESULTS

HABITAT

Mean water temperature varied among years across all locations in the St. Marys River with the highest mean temperatures occurring in 2007 (range 14.2 - 22.3°C) and the lowest temperatures occurring in the 2015 (range 6.4 - 12.1°C; Table 1). In the Mississagi River, the highest mean water temperature during sampling occurred in 2014 (mean 19.4°C).

Mean dissolved oxygen (mg/L) levels varied among years at individual sites in the St. Marys River, ranging from 9.45 - 12.64 mg/L over the six years of sampling. Mean dissolved oxygen levels are inversely related to water temperature, as the lowest oxygen levels were observed in 2007 when the water temperatures in the St. Marys locations were the warmest. Mean dissolved oxygen levels were also lower in the Mississagi River, ranging from 9.10 to 9.26 mg/L, where the water was also relatively warmer than most years on the St. Marys River (Table 1).

In the St. Marys River, mean conductivity ranged from $70.8 - 141.75 \ \mu$ S in the five fishing locations and six years of sampling. Conductivity was lower in all fishing locations in 2015. Conductivity showed unusually high annual variation in the Mississagi River, with an average of 141 μ S in 2014 and an average of 71.8 μ S in 2015.

Bottom substrate in most transects consisted of sand and some silt with macrophytes ranging from zero to sparse in most locations. Lake George transects contained the least amount of macrophytes overall, while the main river had the most sites with sparse to moderate macrophytes (Figure 2). Macrophyte density among the years was not significantly different in either the main (ANOVA F $_{5,121} = 1.915$, p=0.097) or upper river (ANOVA F $_{5,89} = 0.203$, p = 0.961) survey locations. In the lower river sites macrophyte density was significantly different among years (ANOVA F $_{5,83} = 6.864$, p< 0.001), with the sites with the most macrophytes

(moderate) sampled in 2008 and the presence of macrophytes diminishing in transects sampled after 2009 (Figure 2). Macrophyte density was also significantly different in Lake George (ANOVA $F_{5,73} = 4.612$, p=0.001), with the majority of the macrophytes (sparse) sampled in 2008 compared to all other years, which were none to sparse. In the Mississagi River, the majority of sites contained zero to sparse macrophytes, with significantly fewer sites with macrophytes in the 2015 sampling (t test, p =0.010).

FISH COLLECTION

In total, 410 sites were electrofished in the St. Marys River over the six years and 45 sites were surveyed in two years of electrofishing in the Mississagi River (Table 3). The number of transects fished per year varied with weather conditions and available fishing time, though in general fewer transects were fished earlier in the time series. Fifty-two species were collected in the St. Marys River electrofishing surveys over the six years of sampling, and an additional three species were collected in the Mississagi River. In 2014, a daytime small boat trawl was added to the fish collection methods to supplement sampling in waters deeper than 1.5 m. A total of 86 trawls were completed over the two years, with the majority of trawl surveys occurring in the St. Marys River (Table 3). Only one additional species, lake sturgeon (*Acipenser fulvescens*) in the Mississagi River, was collected in with the addition of the trawl surveys. The cumulative number of species collected across all years in both sampling locations with the two gear types was 56 (Table 4).

On average, 33.5 species were collected per year across all four sampling sites in the St. Marys River. The highest species richness (36 species) was observed in 2006 and 2014, while the fewest species (29) were collected in 2015. The greatest number of species collected in a given sampling site was in the main river in 2007, with 30 different species captured, and the fewest species were observed in Lake George in 2008 with 8 (Table 3). The number of species/transect collected in lower river and the upper river were not significantly different among years (Table 5). In the lower river, the mean number of species collected was 7.2 per transect, with a range of 17 - 24 total species per year (Table 3). The upper river was less diverse with a mean of 3.5 species per transect collected and a range of 12-16 total species per year (Table 3). In Lake George and the main river, the number of species captured was significantly different among years (Table 5). In Lake George, an average of 4.3 species/transect, ranging from 8 to 19 total species per year (Table 3). Species captured/transect in 2015 (2.6 species/transect) and 2008 (4.0 species/transect) were significantly lower than all other years, with a peak of 5.4 species/transect in 2006 (Table 5). In the main river, a mean of 6.6 per species per transect were collected, with a range of 20 to 30 total species per year (Table 3). Capture rates in 2015 (3.8 species/transect), 2014 (5.7 species/transect) and 2009 (6.8 species/transect) were significantly lower than all other years (Table 5). There was no significant difference in species richness in the Mississagi River between years, with 25 species sampled in 2014 and 20 in 2015 (Table 5). On average, 6.4 species were collected per transect in the Mississagi River.

A total of 13,180 individual fish were sampled from the St. Marys River electrofishing transects (n=410), while 1,110 fish were sampled from the Mississagi River transects (n=45). An additional 4,095 fishes were collected in the trawl nets, with 3,850 captured in the St. Marys River and 245 caught in the Mississagi River. Abundance/transect in the Mississagi River and in

Lake George was not significantly different among years (Table 5). Abundance/transect was significantly different among years in the lower, main and upper river sections of the St. Marys River (Table 5). In the lower river, catch/transect was not significantly different in years 2008 (27.0 fish/transect) and 2015 (39.9 fish/transect), however, these were significantly lower than catch rates in 2006 (114.4 fish/transect) and 2014 (85.7 fish/transect). In the main river, catch/transect rates were lowest in 2015 (9.1 fish/transect) and 2009 (26.1 fish/transect) compared catch rates in all other years, with the peak catch/transect observed in 2008 (48.2 fish/transect; Table 5). In the upper river, catch/transect was significantly higher in 2014 (37.5 fish/transect) than all other years, which ranged from 7.0 (2015) to 20.1 (2009) fish/transect. Surveys in 2014 coincided with peak yellow perch spawning temperatures, as there was an average yellow perch catch of 31.3 fish/transect collected in 2014, 3 times higher than the previous peak collection rate of 10.7 fish/transect in 2009.

Patterns in mean biomass/transect followed the patterns observed in species richness (Table 4). In the lower and upper St. Marys River and in the Mississagi River, biomass was not significantly different among years (Table 4). In Lake George biomass/transect was highest in 2007 (1112.4 grams/transect) and significantly higher than both 2009 (165.4 grams/transect) 2014 (204.1 grams/transect). In the main river, biomass/transect was lowest in 2015 (458.9 grams/transect) compared to all other years fished (Table 5).

Of the 56 species that were collected during the surveys (Tables 6 and 7), some were ubiquitous throughout the study locations including rainbow smelt (Osmerus mordax), white sucker (Catostomus commersoni), emerald shiner (Notropis atherinoides), spottail shiner (N. hudsonius), mimic shiner (N. volucellus), bluntnose minnow (Pimephales notatus), rock bass (Ambloplites rupestris), yellow perch (Perca flavescens) and Johnny darter (Etheostoma exile). Others such as pink salmon (Oncorhynchus gorbuscha), cisco (Coregonus artedi), central mudminnow (Umbra limi) longnose dace (Rhinichthys cataractae), silver shiner (Notropis photogenis), and white bass (Morone chrysops) were rare and captured only once or within a single sampling year. Some fishes were localized and captured multiple years but only in certain sections of the river; for example, lake whitefish (Coregonus clupeaformis) and round whitefish (Prosopium cylindraceum) were only captured in the upper river, sea lamprey (Petromyzon marinus), Atlantic salmon (Salmo salar), longnose sucker (Catostomus catostomus), blacknose shiner (Notropis heterolepis) and brook stickleback (Culaea inconstans) were only observed in the main river, alewife (Alosa pseudoharengus) was only captured in Lake George, and silver redhorse (Moxostoma anisurum) was only seen in the lower river. Three additional species were collected in the Mississagi River electrofishing surveys; longnose gar (Lepisosteus osseus), bowfin (Amia calva) and common carp (Cyprinus carpio). The invasive fish white bass (Morone chrysops) and alewife were captured in the early sampling years and reported on by Pratt and O'Connor (2011); however, they were not collected in any gear or location sampled in 2014 or 2015. Another invasive fish, the threespine stickleback (Gasterosteus aculeatus), was captured the main river in the early survey years (Pratt and O'Connor 2011). In 2014 and 2015, the threespine stickleback was again collected in the main river, but its range had expanded to include capture in Lake George and the lower river. The threespine stickleback was not observed in the Mississagi River.

The diversity of fishes captured in the trawl transects was low, and with the exception of the age-0 lake sturgeon captured in the Mississagi River, no additional species were added via trawling (Table 7). However, the trawl sampled several species better than boat electrofishing. Brook stickleback, Iowa darter (*Etheostoma exile*), Johnny darter and mottled sculpin (*Cottus bairdi*) were all caught in higher numbers in the trawl.

Historical Comparison

The species catches were ranked and the top ten catches for each fishing area and gear type were contrasted with the rankings of species catch from previous surveys (Table 8). The rankings were compared with the electrofishing surveys of 2006-2009 (Pratt and O'Connor 2011) and the trawl surveys of Liston et al. (1980, 1983) and Liston and McNabb (1986). Similar to the findings of Pratt and O'Connor (2011), trout-perch (*Percopsis omiscomaycus*) catches continued to rank lower than previous surveys, while the ranking of mimic shiner and bluntnose minnow remained higher than in the historic trawling data. Emerald shiner continued to rank strongly in Lake George and the lower river. The addition of the trawl net did improve the catch ranking for Iowa and Johnny darters, bringing the rankings similar to those of the historic trawl surveys. The invasive threespine stickleback was consistently collected in the main river and ranked fifth in abundance in the Lake George trawl survey.

WITHIN SITE COMPARISONS

Index of Biotic Integrity

An IBI was calculated for the Mississagi River, and each of the four sections of the St. Marys River. In the upper river and the Mississagi River, the IBI was not significantly different among years. In the upper river, the mean IBI ranged from 41.8 through 53.2 over the six sampling periods while in the Mississagi River, the mean IBI was 65.9 and 65.6 in 2014 and 2015, respectively. In the lower river, Lake George and the main river, significant differences among years were noted, with the lowest IBI score in all three locations in occurring in 2015 (Tables 9 and 10; Figure 3). The 2015 lower river IBI score (59.4) was significantly lower than in 2006 (78.8) and in 2014 (71.8). The overall mean IBI score for the lower river was 67.5. In Lake George, the 2015 IBI score, 43.9, was significantly lower than in 2007 (54.9) and 2009 (56.7), with a mean IBI score of 52.3 for all years (Table 10, Figure 3). In the main river, the 2015 IBI score, 41.5, was significantly lower than in 2008 (58.4), 2009 (49.3), and 2014 (52.8) with an overall mean IBI score of 50.9 for all years (Table 10, Figure 3).

An assessment of the individual metrics of the IBI was completed for each of the river sections; the upper, main and lower St. Marys River along with Lake George, as well as the Mississagi River (Table 9). Five of the metrics deal with species richness, with four of the metrics, including native species, centrarchid, (turbidity) intolerant, native cyprinid species richness having a positive influence on the IBI score and one, non-native species richness, a negative influence (Table 2). In the Mississagi River, there was no significant difference in any of the species richness factors between the two survey years (Tables 9 and 10, Figures 4 and 5). In the lower river, four of the five metrics were not significantly different over the years, with only the turbidity intolerant species showing significant changes over time (Table 9). 2006 had the

highest number of intolerant species, while the fewest occurred in 2008 (Tables 9 and 10, Figure 4). In Lake George, all five of the species richness metrics were significantly different during the sampling period. Native species richness was lowest in 2015 and highest in 2006 (Tables 9 and 10, Figure 4). Centrarchids were not commonly captured in Lake George; however significantly higher catches occurred in 2007, 2008, 2009, when compared to the other survey years (Tables 9 and 10, Figure 4). Peak captures of turbidity intolerant species occurred in 2006, with the lowest capture rates in 2015 (Tables 9 and 10, Figure 4). Native cyprinid richness was lowest in 2015 compared will all other fishing years and non-native species richness was highest in 2014 (Tables 9 and 10, Figure 5). In the main river section, centrarchid species richness did not change over the sampling period, however, the other four species richness measures differed significantly (Table 9). 2015 had the lowest native species richness and the lowest native cyprinid species richness in the main river and the highest number of non-native species sampled (Tables 9 and 10, Figures 4 and 5). The greatest number of turbidity intolerant species was collected in 2007 (Tables 9 and 10, Figure 4). In the upper river section, there was no difference among years for either the native species or the number of native cyprinids collected during the sampling period (Table 9). Centrarchid species richness was significantly lower in 2014 compared to 2009 (Tables 9 and 10, Figure 4). Intolerant species richness was significantly lower in 2006 and 2014 than in 2009 (Table 9). Non-native species richness was significantly higher in 2006 compared to all other years (Tables 9 and 10, Figure 4).

The biomass metrics, including percent piscivore, percent generalist, and percent specialist fishes, were relatively stable at the individual survey locations among the years with a few exceptions. The percentage of specialists was significantly lower in lower river section in 2015 when compared to 2009, and the percentage of generalists was significantly higher in 2015, compared to 2007 and 2008 (Tables 9 and 10, Figure 5). In the main river, the percentage of generalists was significantly lower in 2015 compared to the survey competed in 2006 (Tables 9 and 10, Figure 5). None of the metrics were significantly different in the Mississagi River over the two sampling periods.

The final four metrics deal with the number of native individual fishes, their biomass, and the percent of non-native species by number and their biomass. The percent of non-natives by number was not significantly different in any of the fishing locations among the years (Table 9, Figure 6). When looking at the percent of non-natives by biomass, only the upper river had a significant difference in capture over the years (Table 9). In 2006 the percent biomass of non-native fishes was higher than any other year (Tables 9 and 10, Figure 6). The highest catches of native fishes the occurred in 2014 in three of the river sections; the upper, main and lower river (Tables 9 and 10, Figure 6); however, a significantly larger biomass of natives was collected in the 2007 surveys in main and Lake George collection sites (Tables 9 and 10, Figure 6). In the Mississagi River, none of the abundance or biomass metrics were significantly different between years.

HABITAT PRODUCTIVITY INDEX

The Habitat Productivity Index (HPI) was calculated for each section of the river over the 6 years of sampling. Across all four study sections of the St. Marys River, the HPI was significantly different among years (Table 9). In the lower river, the HPI in 2006 was not significantly

different than in 2014, but it was significantly higher than all other years surveyed (Tables 9 and 10, Figure 3). In Lake George, the HPI in 2007 was significantly higher than any other year surveyed (Tables 9 and 10, Figure 3). In the main river, the HPIs calculated in 2006-2008 were significantly higher than those in 2009 and 2014-15 (Tables 9 and 10, Figure 3). In the upper river, the HPI for 2014 was higher than all other years, but was only significantly greater (than the 2015 survey (Tables 9 and 10, Figure 3). The HPI was not significantly different between years in the Mississagi River, with a mean HPI of 16.7.

SITE COMPARISON

The fish community sampled at each location in 2014 and 2015 was compared with the results of Pratt and O'Connor (2011) for four metrics, including biomass, species richness, IBI and HPI (Table 9). Like the results from the initial survey years, the upper river and Lake George were significantly lower in biomass and species richness in comparison with the main and lower river areas. The Mississagi River was similar in biomass and species richness to the lower St. Marys River. In 2014, there was large increase in biomass observed in the upper river. However, this was a one year increase as biomass in 2015 was similar to the 2006-2009 catches. When taking into account the 2014 and 2015 data, the lower river had the highest HPI, IBI, and species richness of the four St. Marys River sites sampled, and the Mississagi River values were not significantly different from those of the lower river (Table 9, Figure 3).

ASSESSMENT OF COMMUNITY HEALTH

The St. Marys River fish community was compared to the Mississagi River (control river) in 2014 and 2015 and with the previous St. Marys River surveys (Pratt and O'Connor 2011) as well as with the Penetang Harbour and Hog Bay, two additional Lake Huron locations. These locations were also used as control areas in Pratt and O'Connor (2011; Table 10). Similar to the findings of Pratt and O'Connor (2011), in 2014 and 2015, the St. Marys River still had the highest proportion of native species richness, native cyprinid species richness and the lowest percentage of non-native species when compared with the Lake Huron sites. The Mississagi River had the highest native and native cyprinid species richness and the lowest non-native species of any of the sites fished. Percent piscivore biomass remains low in the St. Marys River, compared with both the Lake Huron and Mississagi River sites while the percent generalist species remain high within the river. The percent specialist species observed was similar at all locations fished. The IBI in the St. Marys River averaged 54 for all years fished, lower than either the Mississagi River (65) and Lake Huron sites (65 and 66), but still within the range of fair (40 – 60) for the fish community.

DISCUSSION

The nearshore electrofishing surveys performed in 2014 and 2015 were completed to complement the surveys accomplished in 2006-2008 and reported on in Pratt and O'Connor (2011), along with an unpublished year of data (2009), as part of the revisiting of the Beneficial Use Impairments (BUIs) for the St. Marys River Area of Concern. These extra sampling years were added to allow an assessment of the temporal stability of the nearshore fish community, and coincided with a change in habitat conditions (increasing flow, lower water temperatures) that

are critical components of fish habitat. Our initial study identified a native fish community that was complex, diverse and healthy, with few apparent impacts of invasive fishes (Pratt and O'Connor 2011). While the additional survey years did not change our overall interpretation, there have been some declines in key measures, including declines in the IBI, species richness and HPI measures.

Habitat conditions in the river might explain some of the apparent decreases observed through time in our surveys. Water temperatures in the last three surveys (2009, 2014 and 2015) were all colder than in the initial 2006-2008 surveys. The winter of 2013-14 had the second highest level of ice coverage in 40 years, and some ice cover was maintained into early June, the longest ever recorded for Lake Superior (Clites et al. 2014). This led to cooler water temperatures in the St. Marys River in 2014, and the cool air temperatures that summer and heavy ice cover again in the winter of 2014-15 helped reduce water temperatures in 2015 to the lowest during any of our surveys. Water temperature impacts electrofishing efficiency by changing the distribution of fishes, fish metabolism and water conductivity (Reynolds and Kolz 2012), resulting in lower electrofishing catches for many fishes under in colder water (e.g., Hall 1986, Borkholder and Parsons 2001). While macrophyte density was not different among the years in either the upper river or main river transects, density was reduced in the lower river transects after 2009 and macrophyte density peaked in Lake George in 2008. In the Mississagi River, macrophyte density was significantly lower in 2015. The reduction in macrophyte density in the last two years of sampling is also likely due to the colder water temperatures as macrophyte production is strongly influenced by water temperature (Barko and Smart 1981). The temperatures in the river in both 2014 and 2015 are reflective of cold water outflow of Lake Superior (Duffy et al. 1987) and the longer ice coverage of both winters.

Another significant change in environmental conditions over the length of the electrofishing surveys were increased water levels increased water levels in the St. Marys River. After 15 years of water level declines, water levels rose rapidly in Lake Superior and Lake Huron and reached their historic average levels in mid-2013 (Gronewold et al. 2016). As a result, the International Joint Commission's International Lake Superior Board of Control adjusted the flow in the St. Marys River to increase to 2980 m³/s through the compensation works, and though reduced to 2650 m³/s for 2014 and 2015, the resulting flows have resulted in higher water levels in later survey years (IJC 2016). This resulted in more wetted bank area, and areas that were unfishable due to shallow depths in earlier survey years were now accessible. The area most affected by the increase in flow was the St. Marys Rapids (which we are unable to sample due to dangerous flows) and the main river section.

The original Degradation of Fish and Wildlife Populations BUI for the St. Marys River fish community stated that the issues were a) trends of lower cisco abundance; b) status of walleye (*Sander vitreum*) spawning stocks within the river; c) potential for exotic, invasive species to invade the fish community, and d) decline northern pike (*Esox lucius*) populations (EC and OME 2011). Our nearshore surveys do not generally sample the larger sport and game fishes due the shallow habitats fished via electrofishing. However, electrofishing can be used to target potential juvenile habitats of these species as well as address the issues of invasive fish species and their effects on the fish community. While our nearshore electrofishing survey is not able to address all of the identified issues alone, additional fisheries surveys, including gill netting and trawling,

have been completed that complement our survey and allow a more comprehensive assessment of the fish community (e.g., Schaeffer et al. 2011, 2017).

The gill netting survey reported on by Schaeffer et al. (2011) addressed the open water fish community trends within the river, and noted that both northern pike and cisco continued to decline between 1975 and 2006, while walleye populations were stable. However, it was noted that the walleye population in the St. Marys River is supported by stocking, as an average ~30% of the fish sampled are of hatchery origin (Schaeffer et al. 2011). Remaining cisco populations in Lake Huron are generally considered to be small, localized, remnant populations and rehabilitation efforts continue (Riley 2010). In Lake Superior, where larger cisco populations support an important commercial fishery, recruitment has been poor and variable over the past two decades and populations are much reduced (Stockwell et al. 2009, Myers et al. 2015). Northern pike populations are declining throughout Lake Huron, and it is hypothesized that populations were negatively influenced by the period of low water levels (Riley 2010). Despite a declining population, the St. Marys River maintains the highest recreational harvest in the Michigan waters of Lake Huron (Riley 2010). It is unlikely that habitat conditions specific to the St. Marys River are responsible for the declines observed in cisco and northern pike, given that the same trends are observed at a broader spatial scale.

It is unlikely that any large bodies of water have been shaped by aquatic invasive species as much as the Great Lakes. There are now over 185 invaders documented in the lakes, and these species have completely reshaped (and continue to reshape) Great Lake's ecosystems (Pagnucco et al. 2015). It is therefore surprising that invasive fishes, while present in the St. Marys River, remain at low levels. Threespine stickleback continued to be caught in the main river, and have moved into both the Lake George and lower river locations. The expansion of this species was also noted in the recent trawl survey (Schaeffer et al. 2017). Rainbow smelt was the most common invasive fish encountered in our surveys, and it continued to be captured in all sections of the river. In our early survey years white bass and alewife were also collected (Pratt and O'Connor 2011), however these species were not collected in the 2009, 2014 and 2015 surveys although a few alewife were captured in the Schaeffer et al. (2017) trawl survey so they remain present in low numbers.

One invasive fish of particular concern to fisheries managers on the St. Marys River is the round goby (*Neogobius melanostomus*), a species that has been sampled in the river and has displaced native fishes in other Great Lakes locations (Kornis et al. 2012). Round goby had just been located at the lowest reaches of the St. Marys River when Pratt and O'Connor (2011) was published, and since then has been captured in low numbers in trawls in the river (Schaeffer et al. 2017). Our inability to sample the round goby is likely due to its continued low abundance in the river, as electrofishing is identified as the most effective tool for sampling this species (Brandner et al. 2013). The migration of Eurasian ruffe (*Gymnocephalus cernua*), another invasive fish, from western areas in Lake Superior appears to have stalled in the Whitefish Bay area (above the entrance to the St. Marys River), as a limited population has been found at the Tahquamenon River. However, annual trawl surveys have not located any within the St. Marys River boundary (Schaeffer et al. 2017) and none were collected in our nearshore surveys.

While some invasive species are located within the river, they do not appear to be displacing the native nearshore fish community from their existing habitats, with the potential exception of the reduction in the number of native sticklebacks compared to the invasive threespine stickleback. While this may change over time, if the offshore shore population of exotic species changes or as potential new invaders move into the nearshore community, at this point the change to the nearshore community from invasive species appears to be very limited. The inability of round goby and other invasive to proliferate in the river as they have in other locations may indicate that the abiotic conditions within the river, as the outflow of Lake Superior, is less prone to successful establishment. It is hypothesized that Lake Superior itself has fewer invasive species than the other Great Lakes due to the inhospitable nature of the lake (Grigorovich et al. 2003). Schaeffer et al. (2017) also hypothesize that the diverse, healthy nature of the St. Marys River fish community has also played a role in resisting the establishment of invasive fishes.

Pratt and O'Connor (2011) noted the differences in the species and abundances collected between the boat electrofishing surveys of 2006-2008 and the previous Liston et al. (1980, 1983) and Liston and McNabb (1986) surveys, and suggested that gear bias could play a role although they argued that there were likely some changes in the existing fish community as well. These suppositions were supported both with the addition of the small Hergog-Armadillo (modified Missouri) trawl net for our 2014 and 2015 surveys, and the trawl survey of Schaeffer et al. (2017). As one would expect, the catches of small bottom-oriented fishes such as Johnny and Iowa darter were higher when using the trawl net; in our trawl survey the ranked catches of Johnny and Iowa darter were again similar to those of the Liston et al. (1980, 1983) and Liston and McNabb (1986) trawl surveys, indicating that multiple gears may be needed to target the full nearshore fish community.

There have been changes within the community between the historic sampling in the 1980s and the more recent sampling, even when taking into account gear differences. The higher abundance of emerald shiner (more tolerant of turbid conditions), bluntnose minnow and mimic shiner, and the reduction in abundance of species such as trout-perch many not be adequately captured in the IBI. While the St. Marys River had fewer non-native species than the other Lake Huron locations, the increase in threespine stickleback in the main river and its movement into Lake George and the lower river may continue to affect the abundance of the native stickleback community. The percent of specialist species collected in the St. Marys River remain high, and Schaeffer et al. (2017) argue that the presence of species such as blacknose shiner is indicative of a healthy community. However, the percent of pisciverous fishes are low and the number of generalist species is high in the river. While it is possible that the number of generalists indicates a lower quality fish community, it is equally possible that the number of generalists indicates species that are more adapted to a riverine environment with its varied habitats such as the St. Marys River, as the Great Lakes IBI was originally developed for non-riverine species (Minns et al. 1994).

The average IBI score calculated for the St. Marys River across all years fished was 54, indicative of a fish community that is in fair condition (Minns et al. 1994). Across all sampling years, the IBI in the lower river was higher, ranging from a low of ~60 to a high of ~80. The lower river IBI was not different from the was average score in the Mississagi River (65) or the other Lake Huron locations, Penetang Harbour (65) and Hog Bay (66). This may be due to its

proximity to the main Lake Huron nearshore fish community, or the more complex and productive habitats available to fishes in that area. The IBI scores in the other three river sections are lower, likely due to low habitat complexity, cool water temperatures and low productivity in those areas. The ability of an IBI to pinpoint individual species and site changes and their effect on the community compared with broad community changes over time can be challenging. For example, Granados (2010) determined that species replacement within the IBI guilds in the Detroit River had hidden changes to the fish community that were not expressed in the IBI. Trebitz et al. (2003) noted that IBIs may detect broad assemblage changes, but may miss the within-site and individual species changes within the IBI area. Gammon and Simon (2000) noted that natural variation and hydrological factors, including temperature and flow, and gear type used all affect the IBI calculation. Similarly, we noted that the water temperatures in 2014 and 2015 were lower than previous years and flow conditions were higher, and that the addition of data from the trawl net returned species rankings for bottom-oriented species (Johnny and Iowa darters) that were missing from our electrofishing surveys. Angermeier and Karr (1986) found that IBI scores increased with the length of stream sampled, due to increased species richness, and that scores also decreased as young-of the year fish were excluded. We did not exclude any fish from our surveys based on age, so this is not an explanation for the lower than expected scores in the St. Marys River. As outlined in Pratt and O'Connor (2011), two factors that are reducing the IBI score in the St. Marys River are low biomass and the low percentage of piscivores. Both of these metrics will be difficult to improve, as low biomass is reflective of low productivity in the river, and the stability of the open water fish community (Schaeffer et al. 2011) means increasing the number of predatory fishes will be difficult. The the index of biotic integrity is sensitive to both of these parameters; in particular, low piscivore abundance is considered evidence of poor conditions (Minns et al. 1994).

Overall, the St. Marys River does support a diverse, species rich community. A total of 52 different species were sampled using boat electrofishing in the river over a period of seven years. The addition of a trawl net in 2014 and 2015 supplemented the abundance of bottom-dwelling species that were otherwise difficult to sample and demonstrated that multiple gear types may be necessary to adequately sample such a diverse nearshore fish community. To obtain an overall picture of the health of the St. Marys River, the nearshore and open water fish community needs to be combined. The continuation of the work completed by the multi-national St. Marys River Task Force, with the large net gillnet survey (Schaeffer et al. 2011), provides an important complement to the work performed herein.

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REFERENCES

Angermeier, P.L., and Karr, J.R. 1986. Applying an Index of Biotic Integrity Based on Sampling and Interpretation. North Am. J. Fish. Manag. 6(3): 418–429. doi:10.1577/1548-

8659(1986)6<418:aaiobi>2.0.co;2.

- Barko, John W., Smart, R.M. 1981. Comparative Influences of Light and Temperature on the Growth and Metabolism of Selected Submersed Freshwater Macrophytes Author (s): John W. Barko and R. Michael Smart Reviewed work (s): Published by : Ecological Society of America Stable URL : htt. Ecol. Monogr. 51(2): 219–235.
- Borkholder, B.D., and Parsons, B.G. 2001. Relationship between Electrofishing Catch Rates of Age-0 Walleyes and Water Temperature in Minnesota Lakes. North Am. J. Fish. Manag. 21(2): 318–325. doi:10.1577/1548-8675(2001)021<0318.</p>
- Brandner, J., Pander, J., Mueller, M., Cerwenka, A.F., and Geist, J. 2013. Effects of sampling techniques on population assessment of invasive round goby Neogobius melanostomus. J. Fish Biol. 82(6): 2063–2079. doi:10.1111/jfb.12137.
- Bray, K.E. 1996. Habitat models as tools for evaluating historic change in the St. Marys River. Can. J. Fish. Aquat. Sci. **53**(S1): 88–98. doi:10.1139/f96-004.
- Brousseau, C.M., Randall, R.G., and Clark, M.G. 2005. Protocol for boat electrofishing in nearshore areas of the lower Great Lakes : transect and point survey methods for collecting fish and habitat data , 1988 to 2002 . Can. Tech. Rep. Fish. Aquat. Sci. **2702**: 85 p.
- Clites, A.H., Wang, J., Campbell, K.B., Gronewold, A.D., Assel, R., Bai, X., and Leshkevich, G.A. 2014. Cold Water and High Ice Cover on Great Lakes in Spring 2014. Earth Obs. Syst. **95**(34): 305–312.
- Duffy, W.G., Batterson, Ted. R., and McNabb, C.D. 1987. The St. Marys River, Michigan: An ecological profile.
- Edsall, T. A. and J. E. Gannon. 1993. A profile of St. Marys River. MICHU-SG-93-700 Michigan Sea Grant College Program. Ann Arbor, Michigan.
- Environment Canada and Ontario Ministry of the Environment. 2011. St. Mary River Area of Concern Canadian Section. Status of beneficial use impairments September 2010. Environment Canada and Ontario Ministry of the Environment. En 164-22/11-2011E
- Gammon, James R., Simon, T.P. 2000. Variation in a Great River Index of Biotic Integrity over a 20-year period. *In* CEUR Workshop Proceedings. *Edited by* S. Jungwirth, M., Muhar, S., Schmutz. Kluwer Academic Publishers. pp. 291–304. doi:10.1023/A.
- Granados, M. 2010. Detecting changes in fish communities in response to habitat rehabilitation: a comparison of multimetric and multivariate approaches. MSc. Thesis. University of Toronto. 98 pp.
- Great Lakes Water Quality Agreement (GLWQA), 1987. Protocolamended by http://www.ijc.org/rel/agree/quality.html 1987.
- Grigorovich, I.A., Korniushin, A. V., Gray, D.K., Duggan, I.C., Colautti, R.I., and MacIsaac, H.J. 2003. Lake Superior: An invasion coldspot? Hydrobiologia **499**: 191–210. doi:10.1023/A:1026335300403.
- Gronewold, A.D., Bruxer, J., Durnford, D., Smith, J.P., Clites, A.H., Seglenieks, F., Qian, S.S., Hunter, T.S., and Fortin, V. 2016. Hydrological drivers of record-setting level rise on Earth's largest lake system. Water Resour. Res. **52**: 1–20. doi:10.1002/2014WR015716.
- Hall, T.J. 1986. Electrofishing Catch per Hour as an Indicator of Largemouth Bass Density in Ohio Impoundments. North Am. J. Fish. Manag. **6**(March): 397–400. doi:10.1577/1548-8659(1986)6<397:ECPHAA>2.0.CO;2.
- Herzog, D.P., Ostendorf, D.E., Hrabik, R.A., and Barko, V.A. 2009. The mini-missouri trawl: A useful methodology for sampling small-bodied fishes in small and large river systems. J. Freshw. Ecol. **24**(1): 103–108. doi:10.1080/02705060.2009.9664270.

- International Joint Commissions International Lake Superior Board of Control. 2016. www.ijc.org/en_/ilsbc/News_Releases.
- Kornis, M.S., Mercado-Silva, N., and vander Zanden, M.J. 2012. Twenty years of invasion: A review of round goby Neogobius melanostomus biology, spread and ecological implications. J. Fish Biol. **80**(2): 235–285. doi:10.1111/j.1095-8649.2011.03157.x.
- Liston, C. R, Duffy, W. G., Ashton, D. E., McNabb, C. D., Koehler, F. E. 1980. Environmental baseline and evaluation of the St. Marys River dredging. U.S. Fish and Wildlife Service. FWS/OBS-80/62.
- Liston, C. R., McNabb, C. D., Duffy, W. G., Bohr, J., Fleisher, G. W., Schutte, and R. Yanusz. 1983. Environmental baseline studies of the St. Marys River near Neebish Island, Michigan, prior to proposed extension of the navigation season. U.S. Fish Wildl. Serv. FWS/OBS-80 (62.3).
- Liston, C. R., McNabb, C. D., 1986. Limnological and fisheries studies of the St. Marys River, Michigan in relation to proposed extension of the navigation season. U. S. Fish Wildl. Srv. FWS/OBS-80(62.3).
- Minns, C.K., Cairns, V.W., Randall, R.G., and Moore, J.E. 1994. an Index of Biotic Integrity (Ibi) for Fish Assemblages in the Littoral-Zone of Great-Lakes Areas of Concern. Can. J. Fish. Aquat. Sci. 51(8): 1804–1822. doi:10.1139/f94-183.
- Moore, J.E., Minns, C.K., Valere, B., Randall, R.G. (n.d.). Productive capacity of Great Lakes nearshore fish habitats: Design and implementation of an electrofishing survey database. Can. Manuscr. Rep. Fish. Aquat. Sci. **2441**: 32.
- Myers, J.T., Yule, D.L., Jones, M.L., Ahrenstorff, T.D., Hrabik, T.R., Claramunt, R.M., Ebener, M.P., and Berglund, E.K. 2015. Spatial synchrony in cisco recruitment. Fish. Res. **165**: 11–21. Elsevier B.V. doi:10.1016/j.fishres.2014.12.014.
- Ontario Ministry of the Environment and Michigan Department of Natural Resources (OMOE and MDNR). 1992. St. Marys Area of Concern: environmental conditions and problem definitions. Remedial Action Plan Stage 1 Report. Toronto, Canada, Ontario Ministry of the Environment.
- Pagnucco, K.S., Maynard, G.A., Fera, S., Yan, N.D., Nalepa, T.F., and Ricciardi, A. 2015. The future of species invsions in the Great Lakes - St. Lawrence River basin. J. Great Lakes Res. 41(Suppl. 1): 96–107.
- Pratt, T.C., and O'Connor, L.M. 2011. An assessment of the health and historical changes of the nearshore fish community of the St. Marys River. J. Great Lakes Res. 37(SUPPL. 2): 61– 69. Elsevier B.V. doi:10.1016/j.jglr.2010.09.009.
- Randall, R.G., and Minns, C.K. 2002. Comparison of a Habitat Productivity Index (HPI) and an Index of Biotic Integrity (IBI) for Measuring the Productive Capacity of Fish Habitat in Nearshore Areas of the Great Lakes. J. Great Lakes Res. 28(2): 240–255. Elsevier. doi:10.1016/S0380-1330(02)70580-7.
- Reynolds, J.B., and Kolz, L.A. 2012. Electrofishing. *In* Fisheries techniques. 3rd ed. *Edited by* A.V. Zale, D.L. Parrish, and T.M. Sutton. American Fisheries Society, Bethesda, Md.
- Riley, S.C. 2010. The State of Lake Huron in 2010. In Great Lakes Fisheries Commission.
- Ripley, M.P., Arbic, B., and Zimmerman, G. 2011. Environmental history of the St. Marys River. J. Great Lakes Res. 37(SUPPL. 2): 5–11. International Association for Great Lakes Research. doi:10.1016/j.jglr.2011.01.008.
- Schaeffer, J.S., Bowen, A.K., and Fielder, D.G. 2017. Community stability within the St. Marys River fish community: Evidence from trawl surveys. J. Great Lakes Res. **43**(2): 399–404.

Elsevier B.V. doi:10.1016/j.jglr.2016.10.014.

- Schaeffer, J.S., Fielder, D.G., Godby, N., Bowen, A., O'Connor, L., Parrish, J., Greenwood, S., Chong, S., and Wright, G. 2011. Long-term trends in the St. Marys River open water fish community. J. Great Lakes Res. 37(SUPPL. 2): 70–79. International Association for Great Lakes Research. doi:10.1016/j.jglr.2011.02.002.
- SPSS 18. SPSS Inc. Released 2009. PASW Statistics for Windows, Version 18.0. Chicago: SPSS Inc.
- Stockwell, J.D., Ebener, M.P., Black, J.A., Gorman, O.T., Hrabik, T.R., Kinnunen, R.E., Mattes, W.P., Oyadomari, J.K., Schram, S.T., Schreiner, D.R., Seider, M.J., Sitar, S.P., and Yule, D.L. 2009. A Synthesis of Cisco Recovery in Lake Superior: Implications for Native Fish Rehabilitation in the Laurentian Great Lakes. North Am. J. Fish. Manag. 29(3): 626–652. doi:10.1577/M08-002.1.
- Trebitz, A.S., Hill, B.H., and McCormick, F.H. 2003. Sensitivity of Indices of Biotic Integrity to Simulated Fish Assemblage Changes. doi:10.1007/s00267-003-0061-y.
- Valere, B.G. 1996. Productive capacity of littoral habitats in the Great Lakes: Field sampling procedures (1988–1995). an Manuscr. Rep. Fish. Aquat. Sci. **2384**: 54. Canadi.

			Number of	Mean	Mean	Mean
Location	Year	Dates	Transects	Temperature	oxygen	conductivity
Upper river	2006	May 30 - June 1	9	11.23 (1.46)	11.97 (0.70)	98.89 (6.01)
	2007	May 28 - 30	17	14.29 (1.43)	12.00 (0.84)	105.88 (6.18)
	2008	June 1-2	15	11.15 (0.73)	11.64 (0.31)	106.67 (4.88)
	2009	May 25 - 26	16	10.78 (1.77)	11.80 (0.41)	108.67 (7.43)
	2014	June 2 - 17	20	11.19 (1.74)	12.70 (0.69)	97.50 (11.18)
	2015	June 4 - 5	19	10.15 (1.38)	11.88 (0.50)	77.39 (24.29)
Main river	2006	June 5-6	12	12.63 (1.42)	12.64 (0.29)	98.33 (7.12)
	2007	June 5-12	19	16.53 (3.23)	10.84 (1.64)	102.78 (6.69)
	2008	June 3-9	15	10.65 (0.99)	12.32 (0.27)	102.00 (4.14)
	2009	May 27 - 31	17	7.39 (0.55)	12.89 (0.40)	103.53 (6.06)
	2014	May 28 - June 10	33	7.07 (1.31)	12.75 (0.25)	102.73 (8.76)
	2015	May 31 - June 1	33	6.41 (1.06)	12.67 (0.70)	75.58 (14.49)
Lake	2006	June 2-3	8	16.11 (0.67)	11.41 (0.23)	103.75 (5.18)
George	2007	June 13-15	22	21.55 (2.59)	9.83 (0.62)	107.14 (5.61)
	2008	June 11	6	15.42 (0.59)	10.14 (0.45)	106.67 (5.16)
	2009	June 12 -13	15	16.01 (0.80)	11.06 (0.39)	102.00 (6.76)
	2014	June 5 - 6	20	13.43 (1.70)	11.80 (0.83)	103.00 (17.2)
	2015	June 2	20	11.93 (2.34)	11.72 (0.34)	90.74 (5.32)
Lower	2006	June 4	5	17.06 (0.37)	11.45 (0.27)	108.00 (4.47)
River	2007	June 17 - 20	13	22.30 (1.19)	9.46 (0.29)	111.54 (5.55)
	2008	June 13 - 18	11	14.75 (0.83)	10.40 (0.40)	108.89 (9.28)
	2009	June 2- 11	25	13.14 (1.95)	11.79 (0.41)	106.80 (5.57)
	2014	June 7 - 9	20	14.38 (1.45)	11.82 (0.30)	104.50 (13.17)
	2015	June 3- 8	20	12.11 (0.90)	11.25 (0.57)	88.31 (14.11)
Mississagi	2014	July 8 - 19	20	16.35 (0.08)	9.26 (0.05)	141.75 (1.12)
River	2015	June 26 - 29	25	19.38 (1.04)	9.10 (0.53)	70.80 (12.83)

Table 1. Sampling dates, effort, and mean water surface temperature, oxygen and conductivity from electrofishing transects in the St. Marys and Mississagi Rivers. Values in brackets represent standard deviations.

Metric Name	Metric Class	Influence on IBI
Native species richness	richness	Positive
Centrarchid species richness	richness	Positive
Intolerant species richness	richness	Positive
Non-native species richness	richness	Negative
Native cyprinid species richness	richness	Positive
Percent piscivore biomass	trophic	Positive
Percent generalist biomass	trophic	Negative
Percent specialist biomass	trophic	Positive
Number of native individuals	number/biomass	Positive
Biomass of natives (kg)	number/biomass	Positive
Percent non-native species by number	number/biomass	Negative
Percent non-native species by biomass	number/biomass	Negative

Table 2. Index of Biotic Integrity (IBI) metrics and their influence on the overall IBI score (positive or negative).

		Number of	Total	Mean Species/	Total	Mean Catch/
Location	Year	Transects	Species	Transect	Catch	Transect
Upper river	2006	9	13	2.8	<u>90</u>	10.0
11	2007	17	16	1.7	279	16.4
	2008	15	14	2.5	135	9.0
	2009	16	16	2.9	321	20.1
	2014	20	12	1.8	749	37.5
	2015	19	14	1.3	133	7.0
Main river	2006	12	27	9.5	573	47.8
	2007	19	30	9.5	725	38.2
	2008	15	25	8.6	723	48.2
	2009	17	24	6.8	443	27.7
	2014	33	25	5.7	978	29.6
	2015	33	20	3.8	300	9.1
Lake	2006	8	14	5.4	171	21.4
George	2007	22	19	4.8	535	24.3
	2008	6	8	4.0	71	11.8
	2009	15	13	4.7	227	15.1
	2014	20	15	4.8	628	31.4
	2015	20	14	2.6	355	17.8
Lower	2006	5	17	9.8	572	114.4
River	2007	13	17	6.2	891	68.5
	2008	11	17	6.6	297	25.1
	2009	25	23	7.7	1473	58.9
	2014	20	24	7.2	1713	85.6
	2015	20	23	7.0	798	39.9
Trawl	2014	35	14		1357	38.8
	2015	36	13		2493	69.3
Mississagi	2014	20	25	6.8	614	30.7
River	2015	25	20	6.1	496	19.8
Trawl	2014	6	9		228	38.0
	2015	9	2		17	1.9

Table 3. Sampling dates, effort, total species, total catch, and mean catch per transect from electrofishing and trawl transects in the St. Marys and Mississagi Rivers.

	St Marys	River Bo	at Electrof	ishing			Mississag Boat Electr		Mississag Trav			ys River awl
	2006	2007	2008	2009	2014	2015	2014	2015	2014	2015	2014	2015
Total species	36	35	31	34	36	29	25	20	9	2	14	13
New species	36	8	1	4	3	0	3	0	1	0	0	0
Cumulative Total	36	44	45	49	52	52	55	55	56	56	56	56

Table 4. Cumulative species catch for the St. Marys and Mississagi rivers, including boat electrofishing and small boat trawl catch.

Table 5. Analysis of variance results for species richness, abundance, and biomass per transect by sites over years from the four locations on the St. Marys River and the Mississagi River. Sites included the upper river (Upper), main river (Main), Lake George (George), lower river (Lower), and Mississagi River (Mississagi).

Parameter	Years	Factor	F	df	р	Tukey HSD
Species/	2006-2015	Upper	1.438	5,90	0.218	
Transect		Main	16.835	5, 123	0.000	2015, 2014<=2009<2006-2008
		George	3.863	5,85	0.003	2015<=2008<=2006-2014
		Lower	1.244	5,88	0.296	
		Mississagi	0.721	1,43	0.401	
Abundance/	2006-2015	Upper	5.191	5,90	0.000	2014>2006-2015
Transect		Main	6.772	5, 123	0.000	2015<=2009<2006-2014
		George	1.768	5,85	0.128	
		Lower	5.497	5,88	0.000	2008, 2015<2006, 2014
		Mississagi	3.552	1,43	0.066	
Biomass/	2006-2015	Upper	1.793	5,90	0.122	
Transect		Main	8.224	5, 123	0.000	2015<2006-2014
		George	4.807	5,85	0.001	2007>2009, 2014
		Lower	1.756	5,88	0.130	
		Mississagi	0.376	1, 43	0.543	
		-				

			Sector and Sector and Sector and Sector and Sector	ientific name Common name	
Sampling location	Year	Actipenser Jutvescens lake sturgeon Petromyzon marinus* sea lamprey Lampetra appendix American brook lamprey	Cono sannon Oncorhynchus gorbuscha pink salmon Alosa pseudoharengus* alewife Amia calva bowfin Lepisosteus osseus longnose gar	Prosopium cylindraceum round whitefish Coregonus clupeaformis lake whitefish Salmo salar Atlantic salmon Oncorhynchus mykiss rainbow trout Oncorhynchus tshawytscha Chinook salmon Oncorhynchus kisutch	Catostomus commersoni white sucker Catostomus catostomus lononose sucker Umbra limi central mudminnow Esox lucius northern pike Osmerus mordax* rainbow smelt Coregonus artedi
St. Marys;	2006			0.22 0.22 0.33 0.11	0.89
Upper river	2007	0.12		0.06 0.47	0.53 1.00
	2008 2009	0.13		0.06 0.06	0.13 1.25 0.75 0.53 3.63
	2014			.10 0.05	0.15 1.55
St. Marys;	2015 2006	$\begin{array}{c} 0.05 \\ 0.42 \ \ 0.42 \end{array}$	($\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{ccc} 0.16 & & 0.68 \\ 4.17 & & 0.5 & 11.2 \end{array}$
Main river	2007	0.42 0.42 0.16		0.11 0.11	0.53 0.05 8.32
	2008	0.07	0.12	0.33 0.07	2.33 0.20 0.5 9.30
	2009 2014		0.12 ($\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$
	2015			.06 0.06 0.45 0.18	1.33 1.70
St. Marys;	2006 2007		$0.13 \\ 0.07$		$\begin{array}{cccccccccccccccccccccccccccccccccccc$
Lake George	2008		0.07		0.83
	2009			1.5 0.05	0.80 0.33
	2014 2015			$\begin{array}{cccccccccccccccccccccccccccccccccccc$	13.8 12.9 0.05
St. Marys;	2006		· · · · · · · · · · · · · · · · · · ·		4.80 0.80 2.60
Lower river	$\begin{array}{c} 2007 \\ 2008 \end{array}$				$\begin{array}{cccc} 0.15 & 0.08 & 0.54 \\ 0.91 & 0.18 & 0.36 \end{array}$
	2009		0.16	0.04	5.04 0.28 0.60
	2014				0.15 0.20 0.70
Mississagi	2015 2014		0.05 0.05	0.15 0.15	$\begin{array}{cccccccccccccccccccccccccccccccccccc$
River	2015				0.56 0.48

Table 6. Scientific and common names of the 56 species collected via electrofishing in the St. Marys and Mississagi rivers 2006-2015. Values are the mean number of fish captured per 100 m transect. Total catch and total species included; (*) marked fishes are considered invasive.

Table 6 continued...

		Scientific name Common name	
Sampling location	Year	Notronis snecies Notropis photogenis silver shiner Semotilus atromaculatus creek chub Rhinichthys atratulus blacknose dace Pimephales notatus blacknose dace Pimephales notatus blacknose shiner Notropis stramineus sand shiner Notropis hudsonius spottail shiner Notropis atherinoides emerald shiner Notemigonus crysoleucas golden shiner Luxilus cornutus common shiner Cyprinus carpio common carp Couesius plumbeus lake chub Moxostoma spp. redhorse sp. Moxostoma anisurum	Ameiurus nebulosus brown bullhead Notropis spp.
St. Marys;	2006	1.67	
Upper river	2007	$0.41 \qquad 0.35 \ 0.06 \ 0.35 \ 3.41$	
	2008	0.13 0.13 0.19 0.47 1.33 0.07	
	2009	0.31 0.38 1.38 1.44	
	2014	0.2 4.9 2.9	
	2015	0.05 0.89 1.68	
St. Marys;	2006	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0.0
Main river	2007 2008	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0.0
	2008	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	
	2009	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	
	2014	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	
St. Marys;	2015	0.25 1.88 2.38 2.5 0.57	0.1
Lake George	2007	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0.0
Luke George	2008	1 2.67 2.17	0.1
	2009	1.8 1.6 1.93 5.8 0.13	-
	2014	7.3 2 0.1 4.6 1.4 0.15	
	2015	0.9 0.5 2.35 0.15	
St. Marys;	2006	0.8 0.2 0.4 8.2 16.4 52.4 1.8	
Lower river	2007	0.08 0.15 0.15 24.6 2 7.46 15.8	0.3
	2008	0.18 0.18 10.9 0.27 3.36 5.18	
	2009	0.12 0.44 0.76 3.12 1.8 1.16 34.3 2.44	
	2014		0.2
	2015	0.05 0.05 6.55 0.65 10.3 13.1	
Mississagi	2014	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	
River	2015	0.12 0.08 0.04 1.08 0.6 0.88 3.88	

Table 6 continued...

								Scienti	fic na	me Co	ommo	n nam	e						To	To
Sampling location	Year	<i>Lota lota</i> burbot	<i>Culaea inconstans</i> brook stickleback	<i>Gasterosteus aculeatus</i> * threespine stickleback	Pungitius pungitius ninespine stickleback	Percopsis omiscomaycus trout-perch	<i>Morone chrysops</i> * white bass	Ambloplites rupestris rock bass	Lepomis gibbosus pumpkinseed	<i>Micropterus dolomieu</i> smallmouth bass	<i>Micropterus salmoides</i> largemouth bass	<i>Perca flavescens</i> yellow perch	<i>Sander vitreum</i> walleye	<i>Etheostoma exile</i> Iowa darter	<i>Etheostoma nigrum</i> Johnny darter	<i>Percina caprodes</i> logperch	<i>Etheostoma</i> spp. darter species	<i>Cottus bairdi</i> mottled sculpin	Total Catch	Total Species
St. Marys;	2006	0.22			0.67	0.33		0.22		0.67		4.33			0.11				90	13
Upper river	2007	0.12				0.18		0.76				8.53		0.06				0.06	279	16
	2008				0.06			0.13		0.00		4.93		0.05	0.33			0.4	135	14
	2009				0.13			1.06		0.06		6.69		0.25	1.38			1.25	321	16
	2014 2015				0.05			0.75 0.58				26.6 1.79			0.1 0.68	0.11		0.1	749 133	12 14
St Momia	2013	0.25	1.08	2.17	0.75	4.25		0.58	0.08	0.83		3.33			0.08	2.92		0.16 2.67	133 573	14 27
St. Marys;	2000	0.23	0.47	2.17	0.75	4.25		1.05	1.37			5.55 9.42	0.32	0.16	0.17	2.92		2.07	725	30
Main river	2007	0.11	0.4	1	0.11	0.2		5.2	0.87	0.52		12.1	0.32		0.93	3.6		1.67	723	25
	2008		0.4	0.82		0.06		3.24		0.12		2.12	0.15	0.15	1.35	1.24		1.76	443	23
	2009		0.00			0.00		2.39	0.00	0.12		5.55	0.06	0.06	0.45	2.09	0.03	0.73	978	25
	2015		0.07	0.03	0.00			0.97	0.12	0.12		1	0.06	0.00	0.73	0.73	0.02	0.64	300	$\frac{1}{20}$
St. Marys;	2006			0.00		0.25		0177		0.12		1.25	0.00		0.13	1		0.0.1	171	14
Lake George	2007					0.27	0.05	0.23	0.05	0.05		5.5	0.05		0.14	1.86			535	19
U	2008					0.5		0.83				2.83							71	8
	2009					0.73		0.2		0.07	0.07	0.93			0.27	0.47			227	14
	2014			0.25		0.2						0.35	0.05		0.4	0.45			628	16
	2015			0.1								0.05			0.25	0.05			355	14
St. Marys;	2006					1		14.8	0.2	2.4		7			0.2	0.4			572	17
Lower river	2007					0		6.92	3.46	1.38	0.05	5.08	0.05		0.08	0.0-		0.15	891	17
	2008			0.04		0.64		0.64	0.27	0.82	0.09	2.73	0.09		0.50	0.09		0.04	297	17
	2009			0.04	0.05	1.84		1.92	0.84	0.48	0.05	2.08	0.04	0.1		0.76		0.04	147	23
	2014			0.15	0.05	1.05		5.75	0.7	1.45	0.05	3.55	0.2	0.1	0.25	0.8		0.1	171	24
Missiagori	2015			0.2		0.35 0.6		2.2	0.1	0.5 0.8	0.05	1.4 6.35	0.2 0.55	0.05	0.45	0.65 1.4		0.1 0.05	798 614	23
Mississagi River	2014 2015	0.16				0.6 1.24		1.75 2.88		0.8	0.03	6.35 3.68	0.55		1.1	1.4	0.04		614 496	25 20
RIVER	2013	0.10				1.24		∠.ðð		0.88		3.08	0.2		1	1.08	0.04	0.16	490	20

Scientific name	Common name	St Marys R	. Trawl	Mississagi	R. Trawl
		2014	2015	2014	2015
Acipenser fulvescens	lake sturgeon			1.5	
Oncorhynchus tshawytscha	Chinook salmon	0.06			
Esox lucius	northern pike	0.09			
Catostomus commersoni	white sucker	0.03		0.33	
Notropis hudsonius	spottail shiner		0.03		
Notropis volucellus	mimic shiner	0.34	0.39		
Pimephales notatus	bluntnose minnow	1.26	0.25		
Culaea inconstans	brook stickleback	2.71	9.31		
Gasterosteus aculeatus*	threespine stickleback	0.2	0.11		
Pungitius pungitius	ninespine stickleback	0.03	0.08		
Percopsis omiscomaycus	trout-perch			3.83	
Ambloplites rupestris	rock bass	0.17	0.06		
Lepomis gibbosus	pumpkinseed	0.94	0.03		
Micropterus dolomieu	smallmouth bass			4.83	
Perca flavescens	yellow perch	0.58	0.22	1.67	
Etheostoma exile	Iowa darter	10.37	15.03	0.83	
Etheostoma nigrum	Johnny darter	18.03	32.36	23.5	1.33
Percina caprodes	logperch	0.23	0.11	0.17	
Cottus bairdi	mottled sculpin	3.71	11.28	1.33	0.56
	Total Catch	1357	2493	228	17
	Total Species	15	13	9	2

Table 7. Scientific and common names of the 19 species collected in the St. Marys andMississagi rivers in 2014-2015 via trawling. Values are the mean number of fish captured per100 m transect. Total catch and total species included.

Table 8. The rank of the ten most abundant species collected in the sampled areas of the St. Marys River with boat electrofishing (this survey: efish 2006-2009 and efish 2014-2015) and trawling (this survey: trawl 2014-2015; past survey; Liston et al. 1980, 1983, and Liston and McNabb 1986) and the Mississagi River (this survey: efish and trawl 2014-2015).

														(Comn	non	nar	ne												
Sampling location	Year	lake sturgeon	coho salmon	rainbow trout	lake whitefish	cisco	rainbow smelt	northern pike	white sucker	common shiner	emerald shiner	spottail shiner	rosyface shiner	sand shiner	mimic shiner	bluntnose minnow	silver shiner	brook stickleback	threespine stickleback	ninespine stickleback	trout-perch	rock bass	pumpkinseed	smallmouth bass	black crappie	yellow perch	Iowa darter	Johnny darter	logperch	mottled sculpin
St. Marys;	efish 2006-2009	-	-	-	-	9	6	_	3	-		10	-	-	4	2	-	-	-	-	-	5	-	-	-	1	-	7		8
Upper river	efish 2014-2015		10				7		4		8				2	3						5				1		6		8
	trawl 2014-2015																	4								5	1	2		3
	trawl 1979-1983				10		7	8	5						4			6		2	10					3	10	1		
St. Marys;	efish 2006-2009						7		1						5	3			10		9	4				2			6	8
Main river	efish 2014-2015			10	į		6		3						1	4						5				2		9	7	8
	trawl 2014-2015															5	7	2	8				6			10	4	1	8	3
	trawl 1979-1983								6			5			3			10		9	2					7		4	8	1
St. Marys;	efish 2006-2009						4		7		1	6		8	3						9	10				2			5	
Lake George	efish 2014-2015		8				1				2	4			3	5										9		6	7	
	trawl 2014-2015																	2	5	6							4	1		3
	trawl 1979-1983						8		6	5		2								9	1	10				3		4	7	
St. Marys;	efish 2006-2009						6				3	7			1	2					9	4	8	10		5				
Lower river	efish 2014-2015								6		3	8			1	2					10	4		7		5			9	
	trawl 2014-2015							8								5		8				7	3			6	2	1		4
	trawl 1979-1983						8		7		6	2			3						1	9			10	4		5		
Mississagi	efish 2014-2015																													
River	trawl 2014-2015																													

Table 9. Analysis of variance results for Index of Biotic Integrity (IBI) metrics: IBI (complete), native, centrarchid, turbidity intolerant, non-native, and native cyprinid species richness; percent piscivore, generalist, and specialist biomass, number of native individuals, biomass of natives (kg); percent non-native species by number and biomass by sites by years and the Habitat Productivity Index (HPI) from the four locations on the St. Marys River and the Mississagi River. Sites included the upper river (Upper), main river (Main), Lake George (George), lower river (Lower), and Mississagi River (Mississagi).

Parameter	Years	Factor	F	df	р	Tukey HSD
IBI	2006-	Upper	1.031	5,90	0.405	
	2015	Main	6.043	5, 123	0.000	2015<2008, 2009, 2014
		George	2.820	5, 85	0.021	2015<2007, 2009
		Lower	2.965	5, 88	0.016	2015< 2006, 2014
		Mississagi	0.004	1, 43	0.950	
Native	2006-	Upper	1.701	5,90	0.142	
Species	2015	Main	16.066	5, 123	0.000	2015<2014<=2009<2006, 2007, 2008
Richness		George	4.876	5, 85	0.001	2015<=2008<2006-2014
		Lower	1.251	5,88	0.292	
		Mississagi	0.780	1,43	0.382	
Centrarchid	2006-	Upper	2.933	5,90	0.017	2014<2009
Species	2015	Main	1.639	5, 123	0.155	
Richness		George	5.143	5, 85	0.000	2006, 2014, 2015<2007, 2008, 2009
		Lower	1.487	5,88	0.202	
		Mississagi	1.626	1, 43	0.209	
Turbidity	2006-	Upper	4.855	5,90	0.001	2015, 2006, 2014<=2007<=2009
Intolerant	2015	Main	3.598	5, 123	0.005	2007>2009, 2015
Species		George	2.503	5, 85	0.037	2006>2015
Richness		Lower	2.651	5,88	0.028	2006>2008
		Mississagi	1.062	1,43	0.309	
Non-native	2006-	Upper	3.741	5,90	0.004	2006>2007-2015
Species	2015	Main	3.809	5, 123	0.003	2014, 2015<2006
Richness		George	3.228	5, 85	0.010	2014>2009, 2007
		Lower	1.692	5, 88	0.145	
		Mississagi	0.154	1,43	0.697	
		0		,		

Parameter	Years	Factor	F	df	р	Tukey HSD
Native	2006-	Upper	1.410	5,90	0.228	
Cyprinid	2015	Main	16.15	5, 123	0.000	2015< 2006, 2008, 2009, 2014< 2007
Species		George	4.527	5, 85	0.0012	2015< 2006, 2009,2014
Richness		Lower	0.420	5, 88	0.834	
		Mississagi	4.200	1, 43	0.047	
Percent		-				
Piscivore	2006-	Upper	1.102	5,90	0.365	
Biomass	2015	Main	0.837	5, 123	0.526	
		George	0.432	5, 85	0.825	
		Lower	1.250	5, 88	0.293	
		Mississagi	0.247	1, 43	0.622	
Percent						
Generalist	2006-	Upper	1.407	5,90	0.229	
Biomass	2015	Main	2.957	5, 123	0.015	2015<2006
		George	0.672	5, 85	0.646	
		Lower	3.146	5, 88	0.012	2015>2007, 2008
		Mississagi	0.004	1, 43	0.947	
Percent						
Specialist	2006-	Upper	0.650	5,90	0.662	
Biomass	2015	Main	2.064	5, 123	0.074	
		George	0.204	5, 85	0.944	
		Lower	2.553	5, 88	0.033	2015<2009
		Mississagi	0.205	1, 43	0.653	
Number of	2006-	Upper	5.201	5,90	0.000	2006-2009, 2015<2014
Native	2015	Main	6.315	5, 123	0.000	2015<=2009<2006, 2007, 2008, 2014
Individuals		George	1.734	5, 85	0.136	
		Lower	5.531	5, 88	0.000	2008, 2015<2006<=2014
		Mississagi	3.562	1, 43	0.066	

Parameter	Years	Factor	F	df	р	Tukey HSD
Biomass of	2006-	Upper	2.257	5,90	0.055	
Natives (kg)	2015	Main	7.142	5, 123	0.000	2009, 2014, 2015<2006, 2007, 2008
		George	4.854	5, 85	0.001	2009, 2014, 2015<2007
		Lower	1.773	5,88	0.127	
		Mississagi	6.33	1, 43	0.431	
Percent of	2006-	Upper	1.205	5,90	0.313	
Non-natives/	2015	Main	1.909	5, 123	0.098	
Number		George	1.984	5, 85	0.089	
		Lower	1.542	5, 88	0.185	
		Mississagi	0.064	1, 43	0.801	
Percent of	2006-	Upper	3.046	5,90	0.014	2006, 2015> 2007-2014
Non-natives/	2015	Main	1.007	5, 123	0.416	
Biomass		George	1.607	5, 85	0.167	
		Lower	0.560	5, 88	0.730	
		Mississagi	0.151	1, 43	0.700	
HPI	2006-	Upper	3.435	5,90	0.007	
	2015	Main	8.532	5, 123	0.000	2015, 2014, 2009<2008, 2007, 2006
		George	8.418	5, 85	0.000	2007>2006-2015
		Lower	5.034	5, 88	0.000	2006>=2006>2007-2015
		Mississagi	0.042	1, 43	0.838	

Table 9 continued...

Table 10. Two-way analysis of variance results for among site and year comparisons from the four locations on the St. Marys River. Data is separated into 2006-2008 (from Pratt and O'Connor 2011), 2009-2015 for the St. Marys River data only and 2006-2015 for the St. Marys River and 2006-2015 all data, which includes the Mississagi River data. Sites included the upper river (Upper), main river (Main), Lake George (George), lower river (Lower), and Mississagi River (Mississagi) for mean transect biomass, species richness, IBI and HPI indices.

Parameter	Years	Factor	F	df	р	Tukey HSD
Biomass	2006-2008	site	23.826	3, 140	< 0.001	Upper, George <main, lower<="" td=""></main,>
		year	5.656	2, 140	0.004	2006, 2007>2008
		site*year	1.409	6, 140	0.215	
	2009 -2015	site	22.358	3,246	< 0.001	George <upper, main<lower<="" td=""></upper,>
	data	year	6.539	2,246	0.002	2015<=2009<=2014
		site*year	1.76	6,246	0.108	
	2006-2015	site	24.952	3, 386	< 0.001	George, Upper< Lower, Main ¹
	data	year	5.154	5, 386	< 0.001	2015<2007, 2006 ¹
		site*year	3.779	15, 386	< 0.001	
						George, Upper< Lower, Main,
	2006-2015 Including	site	17.509	4, 445	< 0.001	Mississagi ¹
	Mississagi	year	4.566	5, 445	< 0.001	2015<2006 ¹
	River	site*year	3.029	16, 455	< 0.001	
Species						
Richness	2006-2008	site	48	3, 140	< 0.001	Upper, George <lower<main< td=""></lower<main<>
		year	3.456	2, 140	0.034	2007<=2006>2008
		site*year	1.427	6, 140	0.208	
	2009 -2015	site	23.807	3, 246	< 0.001	Upper, George <main<lower< td=""></main<lower<>
	data	year	9.959	2,246	< 0.001	2015<2014<=2009
		site*year	1.289	6, 246	0.263	
	2006-2015	site	53.529	3, 386	< 0.001	Upper, George< Main, Lower ¹
	data	year	9.22	5, 386	< 0.001	2015<2006-20141
		site*year	3.907	15, 386	< 0.001	
	2006-2015	site	43.470	4, 445	< 0.001	Upper, George< Main, Lower, Mississagi ¹

	Including					
	Mississagi	year	9.257	5, 445	< 0.001	$2015 < 2006, 2006, 2009^1$
	River	site*year	3.666	16, 455	< 0.001	
IBI	2006-2008	site	21.54	3, 140	< 0.001	Upper <george<main<lower< td=""></george<main<lower<>
		year	1.124	2, 140	0.328	
		site*year	1.581	6, 140	0.157	
	2009 - 2015	site	23.295	3, 246	< 0.001	Upper, Main, George< Lower
	data	year	11.229	2,246	< 0.001	2015<2009,2014
		site*year	0.557	6,246	0.764	
	2006-2015	site	36.925	3, 386	< 0.001	Upper< George, Main < Lower
	data	year	7.855	5, 386	< 0.001	2015<2006-2014
		site*year	1.118	15, 386	0.338	
						Upper, Main,
	2006-2015 Including	site	37.907	4, 445	< 0.001	George <mississagi<=lower 2015<2009, 2006, 2008<= 2014.</mississagi<=lower
	Mississagi	year	7.216	5,445	< 0.001	2007
	River	site*year	1.345	16, 445	0.166	
HPI	2006-2008	site	21.683	3, 140	< 0.001	Upper, George <lower<main< td=""></lower<main<>
		year	1.977	2, 140	0.142	
		site*year	1.8	6, 140	0.12	
	2009 - 2015	site	12.32	3,246	< 0.001	George <upper<=main<lower< td=""></upper<=main<lower<>
	data	year	11.527	2,246	< 0.001	2015<=2009<2014
		site*year	2	6,246	0.066	
	2006-2015	site	28.463	3, 386	< 0.001	George, Upper< Main, Lower ¹
	data	year	8.427	5, 386	< 0.001	2015< 2014, 2007, 2006 ¹
		site*year	4.305	15, 386	< 0.001	
	2006-2015 Including	site	22.447	4, 445	< 0.001	George, Upper< Main, Lower ¹
	Mississagi	year	8.174	5,445	< 0.001	$2015 < 2014, 2007, 2006^1$
	River	site*year	4.454	16, 455	< 0.001	,,

Table 11. Mean values for 10 of the metrics used in the IBI calculation and the HPI for transects sampled in the St. Marys and Mississagi Rivers 2006 - 2015. The values from 2006-2008 are grouped, following Pratt and O'Connor (2011). Penetang Harbour and Hog Bay data from Brousseau et al. (2004) and are provided for comparison.

	Metric name												
Location	Timeframe	Biomass (kg)	Number captured	Species richness	Native species richness	Native cyprinid species richness	Percent piscivore biomass	Percent generalist biomass	Percent specialist biomass	Percent non- native species by number	Percent non- native species by biomass	Index of biotic integrity	Habitat productivity index
	2006-2008	0.6	12.3	3.6	3.5	1	3.0	14.5	75.1	1.1	5.0	48.0	9.5
St. Marys River	2009	0.4	20.1	4.6	4.6	1.3	0.17	22.8	64.6	0	0	50.3	9.3
Upper river	2014	1.2	37.5	3.2	3.1	1.2	1.0	24.6	69.4	0.5	1.0	49.2	28.8
	2015	0.5	7.0	2.9	2.8	0.8	7.8	29.8	57.1	6.6	7.8	43.8	5.7
	2006-2008	2.7	43.9	9.2	8.3	2.2	8.1	64.6	27.4	6.3	2.3	56.9	37.1
St. Marys River	2009	1.1	26.1	6.8	6.0	1.8	0.5	54.1	39.6	5.5	0.8	49.3	16.8
Main river	2014	1.0	29.6	5.7	5.2	1.5	3.0	53.2	43.8	3.8	3.8	52.8	19.3
	2015	0.5	9.1	3.8	3.4	0.5	8.0	36.5	46.4	4.2	7.9	41.5	8.2
	2006-2008	0.8	21.6	4.8	4.8	1.9	3.0	31.8	65.2	0.2	2.2	54.9	11.2
St. Marys River	2009	0.2	15.1	4.7	4.7	2.5	8.1	25.5	66.3	0	0	56.7	3.9
Lake George	2014	0.2	31.4	4.8	4.3	2.7	8.6	17.9	68.5	3.8	9.5	52.5	6.2
	2015	0.1	17.8	2.6	2.25	1.15	6.5	25.2	63.3	4.1	8.3	43.9	2.1
	2006-2008	1.5	60.7	7.0	7.0	3.1	28.6	21.9	49.5	0	0	70.9	27.3
St. Marys River	2009	1.3	60.0	7.3	7.5	3.0	15.6	31.5	52.8	0.2	0.1	66.7	17.7
Lower river	2014	1.5	66.8	7.3	7.1	2.9	26.3	35.2	38.4	0.2	0	71.8	28.0
	2015	1.8	39.9	7.0	6.8	2.9	21.4	55.9	22.7	2.8	0.1	59.4	21.1
St. Marys River	2006-2015	1.1	32.1	5.5	5.2	1.9	9.1	35.4	52.3	2.6	3.2	54.3	17.0
Mississoni Diver	2014	1.7	30.7	6.8	6.7	2.6	39.5	12.0	48.5	0.6	4.7	65.9	17.1
Mississagi River	2015	1.3	19.8	6.1	6.0	1.8	33.9	12.5	53.6	0.8	7.4	65.6	16.3
Penetang Harbour		1.6	30.5	5.4	5.0	1.2	34.4	7.4	54.7	4.5	4.3	64.8	N/A
Hog Bay		4.5	26.1	6.7	6.3	1.1	43.1	20.6	36.3	3.0	15.8	66.0	N/A

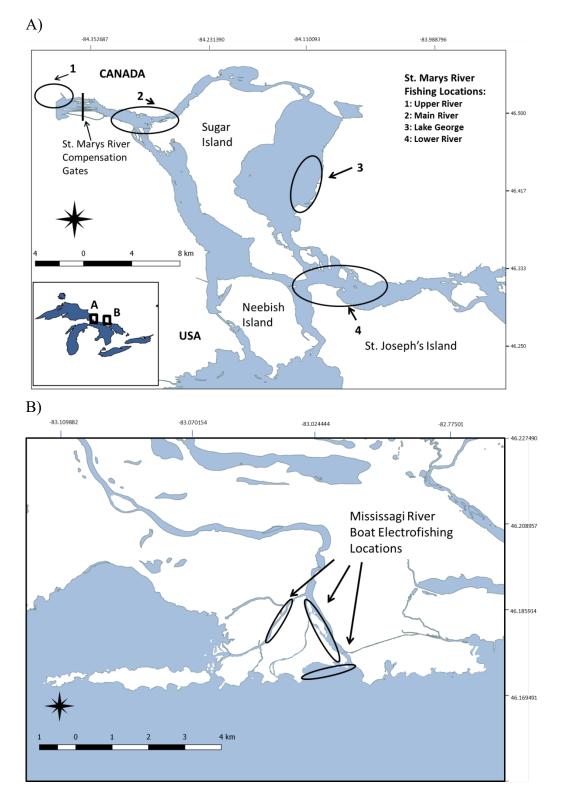


Figure 1: Map of the A) St. Marys River and B) Mississagi River, with survey locations indicated by ovals. The inset shows the location of the rivers in the broader Great Lakes.

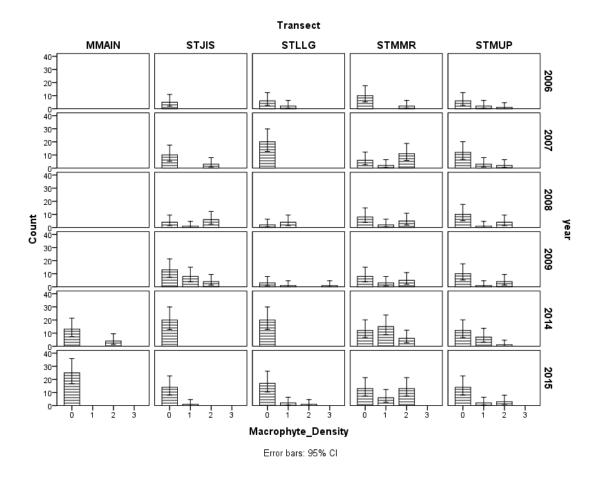


Figure 2: Macrophyte density location electrofished by year. MMain: Mississagi River; STJIS: lower St. Marys River; STLLG: Lake George; STMMR: St. Marys River, Main; STMUP: St. Marys River upper. Macrophyte densities are represented by 0 = none (0%), 1 = sparse (1 to 19% cover), 2 = moderate (20 to 70% cover), and 3 = dense (>70% cover). Error bars represent the 95% confidence limits.

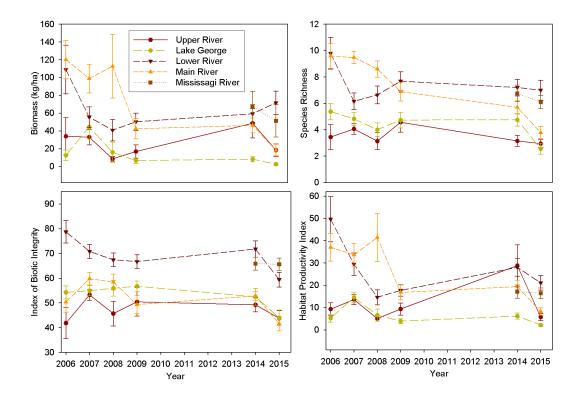


Figure 3: Mean biomass (top left panel), species richness (top right panel), Index of Biotic Integrity (bottom left panel), and Habitat Productivity Index (bottom right panel) by year for each of the four study areas in the St. Marys River and the Mississagi River.

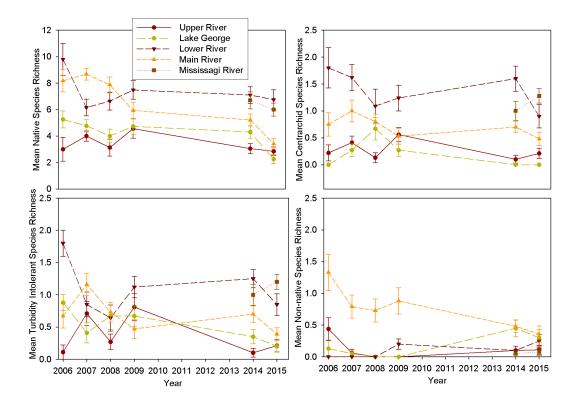


Figure 4: (a) mean native species richness, (b) mean centrarchid species, (c) mean turbidity intolerant species richness, (d) mean non-native species richness by year for each of the four study areas in the St. Marys River and the Mississagi River. Data for 2006-2008 from Pratt and O'Connor (2011).

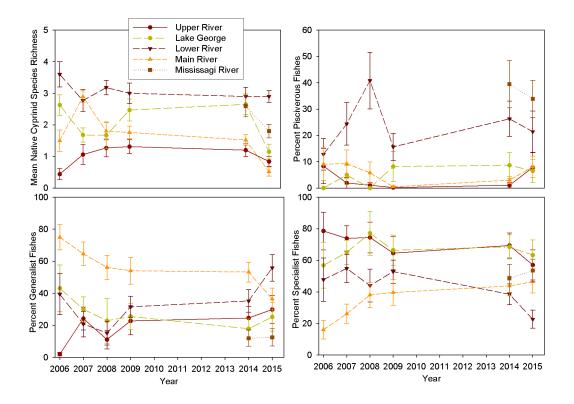


Figure 5: (a) mean native cyprinid species richness, (b) percent pisciverous fishes, (c) percent generalist, (d) percent specialist fishes by year for each of the four study areas in the St. Marys River and the Mississagi River. Data for 2006-2008 from Pratt and O'Connor (2011).

Appendix 1b: An assessment of the health and historical changes of the nearshore fish community of the St. Marys River (Pratt & O'Connor, 2011)

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An assessment of the health and historical changes of the nearshore fish community of the St. Marys River

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ABSTRACT

We examined the nearshore fish community in the St. Marys River using a standardized boat electrofishing protocol to (1) compare the current nearshore fish community of the river to previous surveys, (2) compare the status of fish communities from four distinct areas of the river (the upper river above the compensating gates, the main river, Lake George, and the lower river), and (3) complete an overall assessment of the fish community using an index of biotic integrity approach. The St. Marys River contains a diverse and complex fish community, with dissimilar fish communities located in broad habitat types along the spatial extent of the river. We demonstrated that the nearshore fish community is relatively unaltered over the past 25 years, with many species that were common in the early 1980s remaining important community members today. More invasive fishes now inhabit the river, but unlike many other areas of the Great Lakes, invasives are not common and do not appear to be negatively affecting native species. The overall health of the St. Marys River fish community compared favorably with relatively un-impacted sites from Lake Huron.

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Introduction

The St. Marys River is a 112-km channel that connects Lake Superior with Lake Huron. The river, the only outlet from Lake Superior, is also the largest tributary to Lake Huron and contains a wide diversity of fish habitats (Bray, 1996). This high habitat heterogeneity has resulted in a widely diverse fish community, containing some 75 fishes from 22 families (Duffy et al., 1987). Despite this diversity, the fish community was listed as impaired as part of the Areas of Concern process due to high sea lamprey (*Petromyzon marinus*) abundance, reliance on stocked non-native salmonines to support recreational and concerns about habitat loss and aquatic invasive species (OMOE and MDNR, 1992; Ripley et al., 2011).

Archaeological evidence suggests that lake whitefish (*Coregonus clupeaformis*) was historically the most important fish to First Nations peoples along the St. Marys River (Duffy et al., 1987), but now a mix of warm, cool and coldwater fishes are regularly captured by recreational and subsistence fishermen and assessed by fisheries management agencies active on the river (Schaeffer et al., 2011). While these ongoing efforts to manage harvestable fishes continue, there has been no published assessment of the status of the nearshore fish community in the St. Marys River. Liston et al. (1980, 1983), Liston

component of a comprehensive St. Marys River fisheries assessment plan (Gebhardt et al., 2002; Schaeffer et al., 2011) because the majority of fishes and the juveniles of managed species are too small to be sampled. In this study, we examine the nearshore fish community in the St. Marys River using a standardized boat electrofishing protocol and use the resulting data to meet three main objectives. Four distinct areas of the river were sampled, and we (1) compared the current nearshore fish community from those areas of the St. Marys River to previous

and McNabb (1986) surveyed the nearshore fish community using bottom trawls to inform about potential dredging impacts with proposed improvements to the shipping/navigation channels, but

these voluminous surveys were not synthesized and made available in

the primary literature. An assessment of the nearshore fish commu-

nity in the St. Marys River is recognized as an important but missing

surveys, (2) compared the status of fish communities within each of those areas and (3) completed an overall assessment of the St. Marys River fish community using an index of biotic integrity (Minns et al., 1994) developed for nearshore areas of the Great Lakes.

Methods

Study area

The St. Marys River is an important corridor for aquatic species, and it has also played a significant role in the human develop of the area by providing hydroelectricity, potable water, a commercial shipping channel, and a food supply (Bray, 1996). The ability of

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organisms to move freely along the length of the river has been gradually reduced with increasing human development. The first locks were constructed in 1797, with the final significant modifications occurring in 1921 with the building of compensating gates at the head of the St. Marys River rapids to regulate water levels on the system, and the expansion of the commercial shipping locks in 1943 (Duffy et al., 1987). Cool, oxygenated water discharges from Lake Superior and provides good quality conditions for most fishes throughout the River. Duffy et al. (1987) identified three distinct areas in the St. Marys River; the upper river, the rapids area, and the lower river, and four primary fish habitats; the open water main stem and embayment reaches, emergent wetlands bordering protected reaches, sand beaches, and the rapids area. We sampled four nearshore areas and habitats in Canadian waters; the upper river, the main river (representing the open water main stem and embayment habitat), Lake George (representing the sand beach habitat), and the lower river (representing emergent wetlands and protected reaches) (Fig. 1). The rapids habitat was not sampled as it is too shallow and swift to sample with an electrofishing boat. Similar sites were fished in the last broad examination of the St. Marys River fish community completed by Liston et al. (1980, 1983), Liston and McNabb (1986). Each site is described in more detail below, and sampling dates and habitat descriptions for each of the areas when fishing occurred are available in Table 1.

Site descriptions

Upper river

Transects in the upper site were located in Marks and Leigh Bays, well above the compensating gates (Fig. 1). The substrate was predominately sand, and there was little structural cover with the exception of sparse aquatic macrophytes (primarily quillwort (*Isoetes riparia*)) and sunken logs leftover from historic logging operations located on the western side of Marks Bay. Water temperature was consistently colder in this part of the river, reflecting its closer proximity to and influence of Lake Superior (Table 1).

Main river

Transects were fished in this location from just below the rapids to above Bells Point (Fig. 1). The main stem of the river had the greatest habitat diversity and highest flows of any of the sampling locations. The substrate was largely cobble and boulders around Topsail Island, with silt and aquatic macrophytes (mostly *Potomogeton* spp. and quillwort) providing physical structure above Bells Point. Abiotic conditions were generally similar to those of the upper river (Table 1).

Lake George

Lake George transects were located off of the mouth of the Bar River. They were dominated by sand substrates and contained almost no

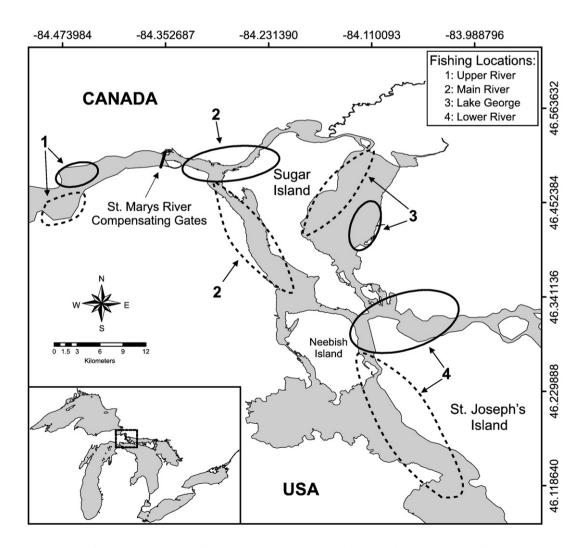


Fig. 1. Map of study area and general fishing locations. The solid ellipses represent the sampling areas from this study, while the hatched ellipses represent the trawling areas from the Liston et al. (1980, 1983) and Liston and McNabb (1986) reports.

Table 1

Sampling dates, effort, and mean water surface temperature, oxygen and conductivity from electrofishing transect sites in the St. Marys River. Values in brackets represent standard deviations.

Location	Year	Sampling dates	# transects	Mean temperature	Mean oxygen	Mean conductivity
Upper river	2006	May 30– June 1	9	11.2 (1.5)	12.0 (0.7)	98.9 (6.0)
	2007	May 28-30	17	14.3 (1.4)	12.0 (0.8)	105.9 (6.2)
	2008	June 1–2	16	11.2 (0.7)	11.6 (0.3)	106.7 (4.9)
Main	2006	June 5–6	12	12.6 (1.4)	12.6 (0.3)	98.3 (7.2)
river	2007	June 5–12	19	16.5 (3.2)	10.8 (1.6)	102.8 (6.7)
	2008	June 3–9	15	10.7 (1.0)	12.3 (0.3)	102.0 (4.1)
Lake	2006	June 2–3	8	16.1 (0.7)	11.4 (0.2)	103.8 (5.2)
George	2007	June 13–15	22	21.6 (2.6)	9.8 (0.6)	107.1 (5.6)
	2008	June 11	6	15.4 (0.6)	10.1 (0.5)	106.6 (5.2)
Lower	2006	June 4	5	17.1 (0.4)	11.5 (0.3)	108.0 (4.5)
river	2007	June 17–20	13	22.3 (1.2)	9.5 (0.3)	111.5 (5.6)
	2008	June 13–18	11	14.8 (0.8)	10.4 (0.4)	108.9 (9.3)

structural cover (Fig. 1). Water temperatures were warmer, the conductivity higher, and dissolved oxygen levels lower for the Lake George sites in comparison to the upper and main river sites (Table 1).

Lower river

Sites in the lower river were fished primarily in the area of Richard's Landing. Habitat diversity was high at the lower river sites; transects in this location were fished over sand, silt and cobble substrates and a variety of aquatic macrophytes were detected growing on the silt substrates (Fig. 1). The lower river sites had abiotic condition similar to Lake George, with higher temperatures and conductivity, but lower dissolved oxygen concentrations than the upper and main river sites (Table 1).

Electrofishing methods

The electrofishing technique used in this survey was developed to assess nearshore fish communities in the Laurentian Great Lakes by Fisheries and Oceans Canada (Valere, 1996; Brousseau et al., 2005). A total of 153 transects were fished in late spring 2006 (34 transects), 2007 (71 transects) and 2008 (48 transects). The earliest and latest dates fished were in 2007, when sampling was initiated on May 28 and ceased on June 20 (Table 1).

Complete details on the sampling procedure are available in Valere (1996) and Brousseau et al. (2005), but a summary is presented below. Prior to the onset of dusk, floats adorned with reflective tape were placed and marked via GPS 100 m apart along a 1.5 m depth contour. Habitat data, including dissolved oxygen (mg/L), conductivity (uS/cm), and water temperature (°C) were taken at the middle of each transect. An Ekman dredge was used to sample substrate at each transect marker, except when the marker was over cobble or boulder substrates which were identified visually. Finer substrates were texturally classified as silt, sand, or gravel. Organic and woody debris were identified in the substrate sample, and dominant (>50%), subdominant (10 to 50%) and trace (<10%) components of the substrate were also recorded. Macrophytes were identified to genus, and macrophyte abundance was determined visually and assigned to one of four categories: none (0% cover), sparse (1% to 19% cover), moderate (20% to 70% cover) or dense (>70% cover).

Surveys were conducted using a Smith-Root SR-14H electrofishing boat (length = 4.4 m, width = 2.0 m). The electric current was regulated through a 5.0 gas-powered pulsator, and the output was kept to 4 A. Transects were 100 m in length, parallel to shore along the 1.5 m depth contour. All fishing occurred after dark, and our target electrofishing time transect was 300 sec per transect. Fishes captured over each transect were held onboard in an aerated live well. Collected fishes were identified to species by the field crew, individually measured (length in mm, weight in grams) and then returned to the water. When fish could not be identified onboard, they were fixed in 10% formalin and returned to the laboratory for later identification and processing. Fish were individually processed up to 20 per species for a given transect, at which point any remaining individuals of that species were counted and batch weighed.

Habitat and fish data from each transect were entered into a database developed specifically for Great Lakes nearshore fish communities (Moore et al., 1998). The database was developed to facilitate the comparison of fish communities, species richness and biomass in areas with different habitat attributes. In addition, the database calculates a nearshore Great Lakes index of biotic integrity (Minns et al., 1994) and a habitat-productivity index (Randall and Minns, 2002). Minns et al. (1994) developed the index of biotic integrity specifically for Great Lakes littoral habitats to quantitatively measure the level of impairment and degree of restoration of fishrelated intrinsic uses of the ecosystem. It uses four classes of factors that determine the condition of Great Lakes littoral fish assemblages which are incorporated into the index of biotic integrity: non-native species, water quality, physical habitat, and piscivore abundance (Minns et al., 1994). Randall and Minns (2002) developed the habitatproductivity index as a measure of habitat productive capacity for fish assemblages (Randall and Minns, 2002). The habitat-productivity index, the product of fish biomass and the production to biomass ratio (P/B) summed for all species captured from a transect accounts for the size structure of the fish community and was used an index of fish production. Together, the habitat-productivity index and the index of biotic integrity provide indices of fish production and diversity (Randall and Minns, 2002). For our survey, total fish biomass from each transect was converted to kg/ha by multiplying the biomass by 40, assuming that the electrofishing boat was effective over a 2.5-m strip along the length of the 100 m transect and that our capture efficiency was 100%.

Data analysis

Historical comparison

While no previous studies allow for a direct comparison of current and past fish community conditions on the St. Marys River, we compared our abundance rankings with those of Liston et al. (1980, 1983), Liston and McNabb (1986) who sampled the nearshore fish community using a 4.9-m bottom trawl with a 3-mm cod end liner deployed for five minute intervals at night. Trawls were performed monthly at two depth intervals, 1.5 m and 3.0 m, from May-November, and the total catches from those trawls were combined in the Liston et al. (1983) and Liston and McNabb (1986) reports. The comprehensive data available from the Liston et al. (1980, 1983), Liston and McNabb (1986) surveys are the only data that have collected small fishes from nearshore areas, so despite the differences in gear (electrofishing versus trawling) we believe that the comparisons may still illustrate potential fish community changes over the past 25 years. Rank abundance data were compared between the current (catches from all three years combined) and Liston et al. (1983) and Liston and McNabb (1986) surveys using the nonparametric gamma correlation coefficient, which is preferable to Spearman R statistic when there are many tied observations (Siegel and Castellan, 1988). For the Liston and McNabb (1986) data, site I was compared with our upper site, sites II and III were combined and assessed against our main site, while sites IV through VII were combined and compared against our lower site (Fig. 1). The data from Liston et al. (1983) were used for the Lake George comparison. The electrofishing and Liston et al. (1983) and Liston and McNabb (1986) trawling sites were in similar areas and habitats but not in the same locations, as we were restricted to Canadian waters and Liston et al. (1983) and Liston and McNabb (1986) were restricted to American waters.

Site comparison

Total fish biomass and productivity, species richness and fish assemblage measures were compared among the four sites using two-factor (site by year) analysis of variance. Biomass values were $\ln (x+1)$ transformed prior to analysis. When significant differences were detected, Tukey honest significant difference tests were used to separate factors (Zar, 1999).

Species and site relationships were also summarized using correspondence analysis, and presented graphically to demonstrate separation (Jackson and Harvey, 1989). Correspondence analysis was selected because it was developed for data where objects of interest are measured by abundance, as abundance and site values are based on an average of each other, giving an ordination of both abundance and sites at the same time (Manly, 1994). We used the type 2 eigenvector standardization recommended by Legendre and Legendre (1998) to maximize distances among species, plotting the mean ordination of each site for visual simplicity.

Assessment of community health

An index of biotic integrity used was developed to evaluate Great Lakes nearshore fish assemblages as an indicator of ecosystem health and habitat quality (Minns et al., 1994). The index was developed using fish data collected under the protocols described in Valere (1996). These standardized fish sampling protocols were followed in this study, making the data collected herein comparable with other published studies that have used the same protocols. Fish community health was assessed with individual index of biotic integrity metrics, and by comparing those metrics and the overall index of biotic integrity with other Great Lakes Area of Concern (Hamilton Harbour and Bay of Quinte, Lake Ontario) reference (West Lake, Lake Ontario) and delisted Area of Concern (Penetang Harbour and Hog Bay, Lake Huron) locations. West Lake, a coastal wetland connected to Lake Ontario near the Bay of Quinte, was selected as a reference area for the Bay of Quinte Area of Concern. Penetang Harbour and Hog Bay are both located in the former Severn Sound Area of Concern. Fish communities were last assessed in Penetang Harbour and Hog Bay in 2002, Hamilton Harbour in 2006, and the Bay of Quinte and West Lake in 2007 (Brousseau et al., 2004; Brousseau and Randall, 2008), and the data from these assessments were used in comparison with our data from the St. Marys River.

Results

Historical comparison

A total of 46 species were captured during our St. Marys River surveys (Table 2). These represented a mix of primarily demersal cold, cool and warmwater fishes. A number of species, including rainbow smelt (Osmerus mordax), white sucker (Catostomus commersoni), emerald (Notropis atherinoides), spottail (N. hudsonius) and mimic (N. volucellus) shiner, bluntnose minnow (Pimephales notatus), troutperch (Percopsis omiscomaycus), rock bass (Ambloplites rupestris), and yellow perch (Perca flavescens), were ubiquitous to all areas and abundant (Table 2). Many other species were rarely captured (e.g. central mudminnow (Umbra limi), rosyface shiner (N. rubellus), blacknose (Rhinichthys atratulus) and longnose dace (R. cataractae)), or only appeared in certain sections of the river (e.g. lake whitefish, lake chub (Couesius plumbeus), burbot (Lota lota); Table 2). Thirtyseven species were captured in the main river, more than the lower (25) and upper (23) river or Lake George (21).

There were significant differences in the species composition in the upper river and Lake George between the trawl data from Liston et al. (1983) and Liston and McNabb (1986) and the contemporary electrofishing survey (upper river $\gamma = 0.16$, P = 0.29, n = 26; Lake George $\gamma = 0.18$, P = 0.21, n = 27), but we found no significant changes in the main and lower river areas (main river $\gamma = 0.44$,

P < 0.001, n = 41; lower river $\gamma = 0.36$, P = 0.004, n = 35). The few differences observed may be due to changes in fish community composition over the past three decades, or they may be attributable to the two gear types used in the studies. Species such as trout-perch and Johnny darter (Etheostoma nigrum) were among the most numerous fish in the early 1980s, but were less common in our electrofishing survey (Table 3). In contrast, bluntnose minnow was more readily observed. Emerald shiner was the most common species in Lake George in our survey, yet the species was not reported in Lake George by Liston et al. (1983). Many species including yellow perch, white sucker, rainbow smelt and spottail and mimic shiner were common in the Liston et al. (1983) and Liston and McNabb (1986) surveys and they remain important community members today. It is also apparent that more invasive fishes have appeared in the St. Marys River. Native sticklebacks (brook stickleback (Culaea inconstans) and ninespine stickleback (Pungitius pungitius)) that were abundant in the historic surveys are no longer as common, and the invasive threespine stickleback (Gasterosteus aculeatus) is becoming more abundant in the main river. Other invasive fishes such as white bass (Morone chrysops) and alewife (Alosa pseudoharengus) were captured in our sampling, but not in the early 1980s work.

Site comparison

There were differences in fish community composition along the downstream gradient of the St. Marys River, with significant differences detected among sites in all four parameters examined in this study (Table 4). The upper river and Lake George had significantly lower total biomass, species richness and indices of biotic integrity and productivity than the main and lower river sites (Fig. 2a–d). The main river had significantly higher species richness and greater productivity than any other site (Fig. 2b, d), while the lower river had the highest index of biotic integrity (Fig. 2c). 2008 collections had the lowest total biomass, richness and productivity of any year. Amongyear patterns in total biomass, species richness and indices of biotic integrity and productivity were relatively consistent in three of the four areas (Fig. 2a-d). The exception was the lower river, which showed a sharp decrease in all four parameters after our initial assessment in 2006, and these declines accounted for the significant year affects in total biomass and species richness.

The among-site fish assemblage differences are reflected in the correspondence analysis, though the amount of variation in species and site relationships was low (Fig. 3). The two locations in the lowest part of the watershed, Lake George and lower river, are in close proximity along with the warm-water species that are mostly restricted to the warmer parts of the river including the main river, with its unique species assemblage composed of cool water species adapted to flowing waters, separates out on the right side of the Figure. Ubiquitous species, such as bluntnose minnow, spottail shiner and rock bass, are located in the middle of the plot, while species adapted to warmer or more turbid conditions (white bass, emerald shiner, largemouth bass (*Micropterus salmoides*)) are associated with the Lake George and lower river sites.

Assessment of community health

The fish community in the St. Marys River appears most similar to the Lake Huron locations, Penetang Harbour and Hog Bay, but there are unique qualities to the St. Marys River community that make it quite different than any of the other Great Lakes locations that were assessed by the same methods. The biomass and number of fish captured and species and native species richness from a given transect, and the overall index of biotic integrity, were all similar to the Lake Huron sites (Table 5). When compared across all sites, the St. Marys River had the highest proportion of native species observed, the highest cyprinid richness, the lowest percentage of non-indigenous

Table 2

The scientific and common names of the 46 species captured via boat electrofishing transects in four areas of the St. Marys River in 2006–2008. Values represent the mean number of fish captured per 100 m transect in a given year. The two letter codes in brackets after the common name represent the species abbreviations found in Fig. 3. Fishes marked with an asterisk (*) are considered invasive.

Family	Scientific and common name	Upper	river		Main river			Lake George			Lower river		
		2006	2007	2008	2006	2007	2008	2006	2007	2008	2006	2007	2008
Petromyzontidae	Lampetra appendix American brook lamprey (AL)			0.13	0.42								
	*Petromyzon marinus sea lamprey (SL)				0.42	0.16	0.07						
Clupeidae	*Alosa pseudoharengus alewife (AW)							0.13	0.07				
Cyprinidae	Couesius plumbeus lake chub (LC)				0.58	0.42	0.13						
	Luxilus cornutus common shiner (CS)			0.13		1.05	0.13			1.00		0.15	0.18
	Notemigonus crysoleucas golden shiner (GS)										0.20	0.15	
	Notropis atherinoides emerald shiner (ES)		0.41	0.13	0.08	0.42	1.33	1.88	12.14	2.67	0.40	24.62	10.91
	Notropis heterolepis blacknose shiner (BN)					0.05							
	Notropis hudsonius spottail shiner (ST)		0.35	0.19	0.25	0.74		2.38	0.27		8.20	2.00	0.27
	Notropis rubellus rosyface shiner (RF)							0.13					
	Notropis stramineus sand shiner (SS)		0.06			0.11							
	Notropis volucellus mimic shiner (MS)		0.35	0.19	5.00	1.47	1.60	2.50	1.73	2.17	16.40	7.46	3.36
	Pimephales notatus bluntnose minnow (BM)	1.67	3.41	1.25	1.92	5.63	5.67	0.50	0.32		52.40	15.85	5.09
	Rhinichthys atratulus blacknose dace (BD)				0.08								
	Rhinichthys cataractae longnose dace (LD)				0.08								
	Semotilus atromaculatus creek chub (CC)			0.06			0.07				1.80		
Catostomidae	Catostomus catostomus longnose sucker (LS)				0.50		0.47						
	Catostomus commersoni white sucker (WS)		1.00	0.50	11.25	8.32	9.40	0.25	0.82	0.83	2.60	0.54	0.36
	Moxostoma macrolepidotum shorthead redhorse (SR)							0.25	0.41		0.80	0.08	0.09
	Moxostoma anisurm silver redhorse (SV)					0.26							0.09
Ictaluridae	Ameiurus nebulosus brown bullhead (BB)					0.05		0.13	0.09	0.17		0.38	
Esocidae	Esox lucius northern pike (NP)					0.05	0.20		0.06		0.80	0.15	0.18
Umbridae	Umbra limi central mudminnow (CM)											0.08	
Osmeridae	*Osmerus mordax rainbow smelt (RS)	0.89	0.53	0.19	4.17	0.53	2.33	10.63	0.27		4.80		0.91
Salmonidae	Coregonus clupeaformis lake whitefish (LW)	0.33	0.47										
	Oncorhynchus tshawytscha Chinook salmon (CH)	0.22			2.08	0.11	0.33						
	Oncorhynchus mykiss rainbow trout (RT)	0.22	0.06		0.08	0.11	0.07						
	Prosopium cylindraceum round whitefish (RW)	0.11											
D 11	Salmo salar Atlantic salmon (AS)	0.00	0.40		0.25	1.10	0.07	0.05	0.07	0.5	1.00		0.04
Percopsidae	Percopsis omiscomaycus trout-perch (TP)	0.33	0.18		4.25	1.16	0.27	0.25	0.27	0.5	1.00		0.64
Gadidae	Lota lota burbot (BU)	0.22	0.12		0.25	0.11	0.40						
Gasterosteidae	Culaea inconstans brook stickleback (BS)				1.08	0.47	0.40						
	* <i>Gasterosteus aculeatus</i> threespine stickleback (TS)	0.67	0.00	0.00	2.17	1.00	1.00						
C	Pungitius pungitius ninespine stickleback (NS)	0.67	0.06	0.06	0.75	0.11	0.20					0.00	
Cottidae	Cottus bairdi mottled sculpin (MO)		0.06	0.38	1.25	1.00	1.67					0.08	
Develop the states	Cottus cognatus slimy sculpin (SC)					1.42			0.05			0.08	
Percichthyidae	*Morone chrysops white bass (WB)	0.00	0.70	0.40	2.00	1.05	5.05		0.05	0.00	4400	6.00	0.04
Centrarchidae	Ambloplites rupestris rock bass (RB)	0.22	0.76	0.13	2.08	1.05	5.07		0.23	0.83	14.80	6.92	0.64
	Lepomis gibbosus pumpkinseed (PU)	0.67			0.08	1.37	0.87		0.05		0.20	3.46	0.27
	Micropterus dolomieu smallmouth bass (SB)	0.67			0.83	0.32			0.05		2.40	1.38	0.82
Deside a	Micropterus salmoides largemouth bass (LB)		0.00			0.10	0.12						0.09
Percidae	Etheostoma exile Iowa darter (ID)	0.11	0.06	0.04	0.45	0.16	0.13	0.45	0.1.		0.00	0.00	
	Etheostoma nigrum Johnny darter (JD)	0.11	0.50	0.31	0.17	0.11	0.93	0.13	0.14	2.02	0.20	0.08	264
	Perca flavescens yellow perch (YP)	4.33	8.53	4.63	3.33	9.42	12.00	1.25	5.50	2.83	7.00	5.08	2.64
	Percina caprodes logperch (LP)				2.92	2.11	3.13	1.00	1.86		0.40		0.09
	Sander vitreum walleye (WE)					0.32	0.13		0.05				0.09

species observed, and the highest percent specialist biomass (Table 5). However, the high specialist biomass meant that there was a low piscivore and generalist biomass in St. Marys River. Only the Hamilton Harbour Area of Concern had a lower piscivore biomass, and the St. Marys River had the lowest percentage of generalists (Table 5). Three of the four locations fished in the St. Marys River had similar individual index of biotic integrity metrics, with the exception of the lower river sites which had greater piscivore biomass (Table 5).

Discussion

Our examination of the nearshore fish community in the St. Marys River fulfills a long-standing desire of the agencies charged with managing the important fishery on this river to concurrently assess juvenile and forage fish status along with the status of the managed game fishes (Gebhardt et al., 2002; for results of managed fish assessment see Schaeffer et al., 2011). Our assessment highlighted a number of interesting findings about the nearshore fish community in the St. Marys River. First, we demonstrated that the nearshore fish community is relatively unaltered over the past 25 years, with many species that were common in the Liston et al. (1980, 1983), Liston and McNabb (1986) surveys remaining important community members today. Secondly, over the past 25 years, more invasive fishes, including threespine stickleback, white bass and alewife have moved into the St. Marys River, but unlike many other areas of the Great Lakes invasives are not common and do not appear to be negatively affecting resident native species. Thirdly, our assessment demonstrated that the high habitat heterogeneity available to fishes in, and large spatial area covered by, the St. Marys River have led to a diverse and complex fish community, with dissimilar fish communities located in broad habitat types along the spatial extent of the river. Lastly, the overall health of the St. Marys River fish community compared favorably with relatively unimpacted sites from Lake Huron.

We observed only limited changes in the St. Marys River fish community from the last time the community was surveyed in the early 1980s. Riverine biodiversity depends upon the interaction of natural and anthropomorphic disturbance patterns and environmental gradients, which produces the spatio-temporal habitat heterogeneity necessary for

Table 3

The rank of the ten most abundant fishes from nearshore boat electrofishing (this survey; Current) and trawling (Liston et al., 1983; Liston and McNabb (1986); Past) in four areas (upper river, main river, Lake George, lower river) of the St. Marys River.

Species	Location							
	Upper ri	ver	Main riv	er	Lake Geo	orge	Lower ri	ver
	Current	Past	Current	Past	Current	Past	Current	Past
Lake whitefish	7	10						
Rainbow smelt	4	7	7		3	8	9	8
Northern pike		8						
White sucker	3	5	1	6	7	6	10	7
Shorthead redhorse					9			
Common shiner						5		
Emerald shiner	7				1	5	2	6
Spottail shiner	7			5	6	2	6	2
Mimic shiner	9	4	6	3	4	2	3	3
Bluntnose	2		3	5	9		1	5
minnow Brook stickleback		6		10				
		0	9	10				
Threespine stickleback			9					
Ninespine	10	2		9		9		
stickleback								
Trout-perch		10	8	2	9	1		1
Rock bass	5		5			10	4	9
Pumpkinseed							7	
Smallmouth bass							8	
Black crappie								10
Yellow perch	1	3	2	7	2	3	5	4
Iowa darter		10						
Johnny darter		1		4		4		5
Logperch			4	8	5	7		
Mottled sculpin			10	1				

highly diverse fish communities (Ward, 1998; Allan, 2004). Fish communities can be transformed by physical or physio-chemical habitat alteration (e.g. Bain et al., 1988; Winston and Taylor, 1991; Wolter et al., 2000; Gehrke et al., 2002; Quinn and Kwak, 2003) or the introduction of invasive species (e.g. Crowl et al., 1992; Mills et al., 1994; Crivelli, 1995). As the majority of landscape-level habitat alterations in the St. Marys River area occurred over 100 years ago (Duffy et al., 1987; Bray, 1996), it may not be surprising large fish community alterations were not observed, even in a disturbed system such as the St. Marys River. Our results are consistent with the findings of Schaeffer et al. (2011) for managed fishes in the St. Marys River, as the community index netting program indicates relative stability in the managed fish community since that program's inception in 1975, and other studies have demonstrated long-term fish assemblage persistence in riverine communities (e.g. Ross et al., 1985;

Table 4

Two-way analysis of variance results for among-site and year comparisons from four locations on the St. Marys River. Mean transect biomass, species richness, Index of Biotic Integrity (IBI), and Habitat-Productivity Index (HPI) were assessed at the upper river (Upper), main river (Main), Lake George (George) and lower river (Lower) sites.

Parameter	Factor	F [*]	Р	Tukey HSD
Biomass	Site	23.8 _{3, 140}	< 0.001	Upper, George <main, 2006,<="" lower="" td=""></main,>
	Year	5.7 _{2, 140}	0.004	2007>2008
	Site * Year	$1.4_{6, 140}$	0.21	
Richness	Site	48.3 _{3, 140}	< 0.001	Upper, George <lower<main< td=""></lower<main<>
	Year	3.62, 140	0.03	$2007 \le 2006 > 2008$
	Site * Year	1.4 _{6,140}	0.21	
IBI	Site	40.13, 137	< 0.001	Upper <george<main<lower< td=""></george<main<lower<>
	Year	0.32, 137	0.76	
	Site * Year	2.0 _{6, 137}	0.07	
HPI	Site	21.7 _{3, 140}	< 0.001	Upper, George <lower<main< td=""></lower<main<>
	Year	2.0 _{2, 140}	0.14	
	Site * Year	1.86, 140	0.12	

It is important to note that our fish community survey was based on boat electrofishing data, while the Liston et al. (1980,1983), Liston and McNabb (1986) surveys relied on 5 min bottom trawls. These two techniques are both capable of successfully characterizing nearshore fish communities (e.g. Yoder and Smith, 1999; Lyons et al., 2001; Bronte et al., 2003), and we believe that both techniques provided a reasonable representation of the relative abundance rankings used in our analysis. Obviously trying to compare more quantitative measures, such as catchper-unit effort among gears, would not be understandable, but others have used relative rankings from different sampling techniques to assess temporal changes in fish communities (Quinn and Kwak, 2003). It is possible that some of the limited differences detected between the Liston et al. (1980, 1983), Liston and McNabb (1986) and our surveys are due to the differences in survey techniques; species that live on the bottom such as Johnny and Iowa darter (Etheostoma exile) may be more likely to be sampled by a trawl than boat electrofishing. However, gear differences are not able to explain the changes in other species, such as the decreases in trout-perch or increases in bluntnose minnow, which occupy a similar position in the water column.

Despite the presence of an increasing number of invasive fishes in the St. Marys River, there is little evidence that the invasive fishes are negatively impacting the fish community. There is concern that the recent invasion of threespine stickleback may result in the displacement of native brook stickleback, as appears to be occurring at other Lake Superior locations (Pratt et al., 2009), and brook stickleback was less commonly observed in comparison with the Liston et al. (1980, 1983), Liston and McNabb (1986) St. Marys River surveys. However, overall less than 1% of the fish captured, and just over 2% of total biomass in our surveys, were non-native despite most invasives being in the community for many years. This seems to indicate that the collapse of native fishes that occurred in the main basins of the Great Lakes (e.g. Vanderploeg et al., 2002; Mills et al., 2003; Dobiesz et al., 2005) has not transpired in the St. Marys River.

There are other examples of communities where invasive fishes have not resulted in large-scale changes in the fish community (Jowett et al., 1998; Eby et al., 2003), and there is a growing body of literature which contends that certain biotic and abiotic conditions make it less likely for invasive species to successfully invade a given community. These include unsuitable or harsh abiotic conditions (Eby et al., 2003), and the presence of a highly heterogeneous receiving community (García-Ramos and Rodríguez, 2002; Olden et al., 2004). Both factors may play a role in protecting the St. Marys River from invasive fishes. Abiotic conditions in the St. Marys River reflect the harsh conditions of Lake Superior, with primarily cold, clear, low productivity waters (Duffy et al., 1987). Grigorovich et al. (2003) observed that Lake Superior had few invasive species despite receiving a high proportion of ballast water discharges, and they attributed this to abiotic conditions that were unfavourable for the establishment and spread of most aquatic invasive species. Additionally, our fish community assessment identified that the St. Marys River fish community is healthy and diverse, containing cold, cool, and warmwater fishes. Together, the physical-chemical and biological conditions may have helped limit the establishment and spread of invasive fishes in the St. Marys River. The resilience of the St. Marys River fish community to invasive fishes will be severely tested in the near future, however, as first reports of the round goby, a fish that has radically altered ecosystems elsewhere in the Great Lakes (Vanderploeg et al., 2002), were just received from the lower river (Anjanette Bowen, USFWS, personal communication).

Our assessment supported the contention that the St. Marys River contains a diverse fish community, representing fishes from a number of thermal niches (Liston and McNabb, 1986; Duffy et al., 1987; Kauss, 1991). Others have characterized the St. Marys River as being a percid-dominated fish community due to the presence of walleye, yellow perch, northern pike, and white sucker (Duffy et al., 1987; Kauss, 1991), but we feel that this depiction is too simplistic and does

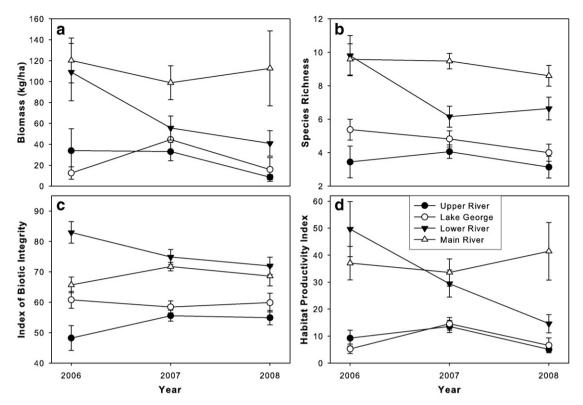


Fig. 2. (a) Mean biomass, (b) species richness, (c) Index of Biotic Integrity and (d) Habitat Productivity Index by year for each of the four study areas in the St. Marys River.

not capture the full complexity of the community. Two areas in the river, the upper river and Lake George sites, were relatively depauperate, had lower biomass and productivity, and were not percid-dominated. The biota of the upper river, closest to Lake Superior's outflow, is heavily influenced by the cold, oligotrophic water arising in Lake Superior (Liston and McNabb, 1986), which was reflected in the consistently lower water temperatures we observed. Consequently, the upper river contained some cold tolerant fishes that were not observed in other locations. In contrast, Lake George's depauperate fish community is comprised of fishes that are primarily large-lake oriented, tolerant of a wide-range of abiotic conditions or fluctuating turbidity (Lane et al., 1996; Scott and Crossman, 1998).

Both the main and lower river sections contained a more diverse fish fauna, as indicated by the high index of biotic integrity scores for those sections of the river. The main river fish community had cold, cool, and warm-water fish represented, likely indicative of the high habitat heterogeneity available in that part of the river (Copp, 1989; Bray, 1996). The lower river was more dominated by warm-water fishes such as cyprinids and centrarchids. This pattern reflects the lotic-to-lentic succession of fish communities that is commonly observed in large rivers, where geomorphic and hydrologic changes occur along the spatial extent of the system (e.g. Amoros et al., 1987; Copp, 1989; Ward, 1998). We observed a trend of decreasing fish community health measures in the lower river over the three years of our study, but the short study duration makes it difficult to know whether managers should be concerned about these decreases. High inter-annual variability in riviverine fish community composition is not uncommon (e.g. Godinho et al., 2000; Bernardo et al., 2003), and to date there is no apparent response from the managed fishes on the river (Schaeffer et al., 2011).

Our assessment indicates that the St. Marys River fish community is reasonably healthy. The index of biotic integrity used in our assessment was developed for nearshore fish assemblages in the Great Lakes, and has been used an indicator of ecosystem health (Minns et al., 1994) and habitat quality (Randall and Minns, 2002). The health of the St. Marys River fish community, based on the mean overall index of biotic integrity score, would be classified by Minns et al. (1994) as good. The overall score and many individual index of biotic integrity metrics were similar to those observed in two Lake Huron locations (Brousseau and Randall, 2008), though the St. Marys River had a number of unique community attributes that indicated a robust and healthy fish community. These included a high proportion of specialists in the community, a high number of cyprinids, and a small contribution of invasive fishes. It is possible that the highly heterogeneous habitats available in the river are providing opportunities for many of these specialized nearshore forage fishes to thrive (Bray, 1996). A score of 80 is required for a fish community to be assessed as excellent, and it is likely that the low overall biomass and low percentage of piscivores captured during our surveys prevented the St. Marys River fish community from scoring higher, as the index of biotic integrity is sensitive to these parameters (Minns et al., 1994). In particular, low piscivore abundance is considered evidence of poor conditions (Minns et al., 1994), so a better balance of specialists and piscivores would have improved the overall assessment.

It is important to recognize that the index of biotic integrity was developed for Lake Ontario (Bay of Quinte and Hamilton Harbour) and Lake Huron (Georgian Bay) locations (Minns et al., 1994), and there may be issues with extending the index outside of those areas (e.g. Schulz et al., 1999). In particular, the low productivity, depauperate nature of the upper river and Lake George sites may not be well-suited to the index as it was developed using more productive areas. That said, as (1) Minns et al. (1994) demonstrated that the index was robust across broad spatial areas and diverse fish assemblages, (2) our sampling locations, with the exception of the upper river location, are considered part of Lake Huron, which includes two sites from which the index was developed, and (3) the overall fish communities are similar among the Lake Huron locations (Brousseau et al., 2004), we are confident that the index provides a fair assessment of the St. Marys River fish community.

In summary, our evaluation of the St. Marys River nearshore fish community is the first spatially extensive assessment of the river in over

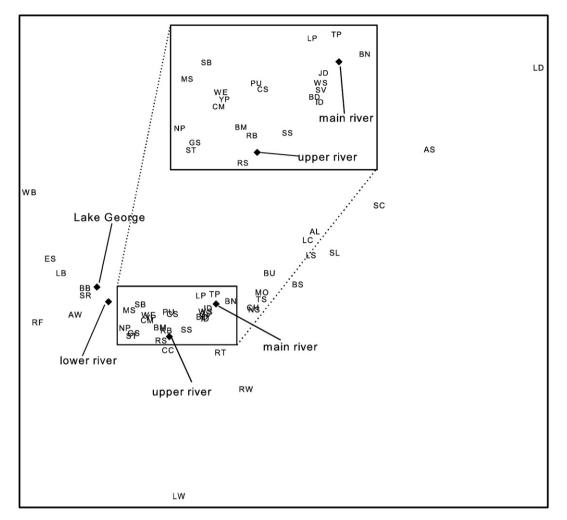


Fig. 3. Dimensions 1 and 2 from a correspondence analysis of fish captures by site (n = 152) from night boat electrofishing transects. Mean ordinations for sites (n = 4) are represented by black diamonds, while species (n = 46) are represented by 2 letter codes. The upper box is an enlarged insert to better display the ordination of the many species clustered near the center of the figure. Dimensions 1 and 2 explain 8.9% and 6.3%, of the variance in the species by St. Marys River location analysis, respectively. Species codes can be found in Table 2.

25 years. We demonstrated that the community has not altered appreciatively over that timeframe, and that it is diverse, robust, and relatively un-impacted by invasive fishes. The concerns that resulted in the fish community being listed as impaired as part of the Great Lakes

Areas of Concern process appear to be mostly ameliorated. While sea lampreys continue to infest the river, recent efforts to reduce sea lamprey abundance with increasing trapping effort, targeted Bayluscide treatments, and a sterile male release program have been successful in

Table 5

The average biomass, catch in numbers, species richness per 100 m transect, and the average index of biotic integrity metrics, from the four St. Marys River sampling locations and five Great Lakes sites. Hamilton Harbour and the Bay of Quinte are both Areas of Concern in Lake Ontario, while West Lake (Lake Ontario), Penetang Harbour (Lake Huron) and Hog Bay (Lake Huron) are reference areas.

St. Marys River	Marys River									Hog
Metric name	Upper river Main river		Lake George	Lake George Lower river		Harbour ^a	Quinte ^b	Lake ^c	Harbour ^c	Bay ^c
Biomass (kg)	0.6	2.7	0.8	1.5	1.5	7.5	5.9	2.4	1.6	4.5
Number captured	12.3	44.0	21.6	60.7	33.3	20.5	72.9	53.7	30.5	26.1
Species richness	3.6	9.2	4.8	7.0	6.2	5.6	8.8	11.1	5.4	6.7
Native species richness	3.5	8.7	4.8	7.0	6.0	4.0	8.1	10.4	5.0	6.3
Native cyprinid species richness	1.0	2.2	1.8	3.1	2.0	0.5	0.4	1.9	1.2	1.1
Percent piscivore biomass	2.6	8.0	3.0	28.6	9.3	5.0	35.7	44.8	34.4	43.1
Percent generalist biomass	2.2	2.2	7.7	4.3	3.9	55.6	11.6	21.7	7.4	20.6
Percent specialist biomass	87.9	89.7	89.3	67.1	84.8	39.4	52.6	33.6	54.7	36.3
Percent non-indigenous species by number	1.1	1.4	0.2	0.0	0.8	36.8	4.5	1.7	4.5	3.0
Percent non-indigenous species by biomass	5.0	0.8	2.2	0.0	2.1	21.2	5.0	0.6	4.3	15.8
Index of biotic integrity	50.2	69.8	59.2	75.2	63.0	40.1	73.5	79.9	64.8	66.0

^a Brousseau and Randall 2008.

^b Christine Brousseau, personal communication.

^c Brousseau et al. 2004.

reducing larval sea lamprey abundance (Schleen et al., 2003), though the number of parasitic sea lampreys remains above target in Lake Huron. Additionally, while non-native salmonines continue to provide extensive angling opportunities in the river, and other invasive fishes have appeared in the community, there appears to be few impacts on native fishes in the St. Marys River from these invasives.

Acknowledgments

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References

- Allan, J.D., 2004. Landscapes and riverscapes: the influence of land use on stream ecosystems. Annu. Rev. Ecol. Evol. Syst. 35, 257–284.
- Amoros, C., Rostan, J.C., Pautou, G., Bravard, J.P., 1987. The reversible process concept applied to the environmental management of large river systems. Environ. Manage. 11, 607–617.
- Bain, M.B., Finn, J.T., Booke, H.E., 1988. Streamflow regulation and fish community structure. Ecology 69, 382–392.
- Bernardo, J.M., Ilhéu, M., Matono, P., Costa, A.M., 2003. Interannual variation of fish assemblage structure in a Mediterranean River: implications of streamflow on the dominance of native or exotic species. River Res. Appl. 19, 521–532.
- Bray, K., 1996. Habitat models as tools for evaluating historic change in the St. Marys River. Can. J. Fish. Aquat. Sci. 53 (Suppl. 1), 88–98.
- Bronte, C.R., Ebener, M.P., Schreiner, D.R., DeVault, D.S., Petzold, M.M., Jensen, D.A., Richards, C., Lozano, S.J., 2003. Fish community change in Lake Superior, 1970– 2000. Can. J. Fish. Aquat. Sci. 60, 1552–1574.
- Brousseau, C.M., Randall, R.G., Clark, M.G., 2004. Comparison of Severn Sound fish assemblages in Hog Bay and Penetang Bay, 1990 and 2002, using two indices of productive capacity. Can. Man. Rep. Fish. Aquat. Sci. No. 2686.
- Brousseau, C.M., Randall, R.G., Clark, M.G., 2005. Protocol for boat electrofishing in nearshore area of the lower Great Lakes: transect and point survey methods for collecting fish and habitat data, 1988 to 2002. Can. Man. Rep. Fish. Aquat. Sci. No. 2702.
- Brousseau, C.M., Randall, R.G., 2008. Assessment of long-term trends in littoral fish community of Hamilton Harbour using an Index of Biotic Integrity. Can. Tech. Rep. Fish. Aquat. Sci. No. xxxx.
- Copp, G.H., 1989. The habitat diversity and fish reproductive function of floodplain ecosystems. Environ. Biol. Fish. 26, 1–26.
- Crivelli, A.J., 1995. Are fish introductions a threat to endemic freshwater fish in the northern Mediterranean region? *Biol.* Cons. 72, 311–320.
- Crowl, T.A., Townsend, C.R., Mcintosh, A.R., 1992. The impact of introduced brown and rainbow trout on native fish: the case of Australasia. Rev. Fish. Bio. Fisheries 2, 217–241.
- Dobiesz, N.E., McLeish, D.A., Eshenroder, R.L., Bence, J.R., Mohr, L.C., Ebener, M.P., Nalepa, T.F., Woldt, A.P., Johnson, J.E., Argyle, R.L., Makarewicz, J.C., 2005. Ecology of the Lake Huron fish community, 1970–1999. Can. J. Fish. Aquat. Sci. 62, 1432–1451.
- Duffy, W.G., Batterson, T.R., McNabb, C.D., 1987. The St. Marys River, Michigan: an ecological profile. U.S. Fish Wildl. Serv. Biol. Rep. 85.
- Eby, L.A., Fagan, W.F., Minckley, W.L., 2003. Variability and dynamics of a desert stream community. Ecol. Appl. 13, 1566–1579.
- García-Ramos, G., Rodríguez, D., 2002. Evolutionary speed of species invasions. Evolution 56, 661–668.
- Gebhardt, K., Fielder, D., Greenwood, S., Robbins, H., Sutton, T., 2002. St. Marys River Fisheries Assessment Plan. Great Lakes Fisheries Commission, Ann Arbor.
- Gehrke, P.C., Gilligan, D.M., Barwick, M., 2002. Changes in fish communities of the Shoalhaven River 20 years after contraction of Tallowa Dam. Australia. River Res. Appl. 18, 265–286.
- Godinho, F.N., Ferreira, M.T., Santos, J.M., 2000. Variation in fish community composition along an Iberian river basin from low to high discharge: relative contributions of environmental and temporal variables. Ecol. Freshw. Fish 9, 22–29.
- Grigorovich, I.A., Korniushin, A.V., Gray, D.K., Duggan, I.C., Colautti, R.I., MacIsaac, H.J., 2003. Lake Superior: an invasion coldspot? Hydrobiologia 499, 191–210.
- Grossman, G.D., Dowd, J.F., Crawford, M., 1990. Assemblage stability in stream fishes: a review. Environ. Manage. 14, 661–671.
- Jackson, D.A., Harvey, H.H., 1989. Biogeographic associations in fish assemblages: local vs. regional processes. Ecology 70, 1472–1485.
- Jowett, I.G., Hayes, J.W., Deans, N., Eldon, G.A., 1998. Comparison of fish communities and abundance in unmodified streams of Kahurangi National Park with other areas of New Zealand. N.Z. J. Mar. Freshw. Res. 32, 307–322.
- Kauss, P.B., 1991. Biota of the St. Marys River: habitat evaluation and environmental assessment. Hydrobiologia 219, 1–35.

- Lane, J.A., Portt, C.B., Minns, C.K., 1996. Adult habitat characteristics of Great Lakes fishes. Can. Man. Rep. Fish. Aquat. Sci, No. 2358.
- Legendre, P., Legendre, L., 1998. Numerical ecology, second edition. Elsevier, Amsterdam.
- Liston, C.R., Duffy, W.G., Ashton, D.E., McNabb, C.D., Koehler, F.E., Liston, C.R., Duffy, W. G., Ashton, D.E., McNabb, C.D., Koehler, F.E., Liston, C.R., Duffy, W.G., Ashton, D.E., McNabb, C.D., Koehler, F.E., 1980. Environmental baseline and evaluation of the St. Marys River dredging. U.S. Fish and Wildlife Service. FWS/OBS-80/62.
- Liston, C.R., McNabb, C.D., Duffy, W.G., Bohr, J., Fleischer, G.W., Schutte, J., Yanusz, R., 1983. Environmental baseline studies of the St. Marys River near Neebish Island, Michigan, prior to proposed extension of the navigation season, 1981. U.S. Fish Wildl. Serv. FWS/OBS-80-62.2.
- Liston, C.R., McNabb, C.D., 1986. Limnological and fisheries studies of the St. Marys River, Michigan in relation to proposed extension of the navigation season. U.S. Fish Wildl. Serv. FWS/OBS-80(62.3).
- Lyons, J., Piette, R.R., Niermeyer, K.W., 2001. Development, validation, and application of a fishbased index of biotic integrity for Wisconsin's large warmwater rivers. Trans. Am. Fish. Soc. 130, 1077–1094.
- Manly, B.F.J., 1994. Multivariate statistical methods: a primer, second edition. Chapman and Hall, London.
- Meffe, G.K., Minckley, W.L., 1987. Persistence and stability of fish and invertebrate assemblages in a repeatedly disturbed Sonoran Desert stream. Am. Midl. Nat. 117, 177–191.
- Mills, E.L., Leach, J.H., Carlton, J.T., Secor, C.L., 1994. Exotic species and the integrity of the Great Lakes. Bioscience 44, 666–676.
- Mills, E.L., Casselman, J.M., Dermott, R., Fitzsimons, J.D., Gal, G., Holeck, K.T., Hoyle, J.A., Johannsson, O.E., Lantry, B.F., Makarewicz, J.C., Millard, E.S., Munawar, I.F., Munawar, M., O'Gorman, R., Owens, R.W., Rudstam, L.G., Schaner, T., Stewart, T.J., 2003. Lake Ontario: food web dynamics in a changing ecosystem (1970–2000). Can. J. Fish. Aquat. Sci. 60, 471–490.
- Minns, C.K., Cairns, V.W., Randall, R.G., Moore, J.E., 1994. An index of biotic integrity (IBI) for fish assemblages in the littoral zone of Great Lakes areas of concern. Can. J. Fish. Aquat. Sci. 51, 1804–1822.
- Moore, J.E., Minns, C.K., Valere, B., Randall, R.G., 1998. Productive capacity of Great Lakes nearshore fish habitats: design and implementation of an electrofishing survey database. Can. Man. Rep. Fish. Aquat. Sci, No. 2441.
- Olden, J.D., Poff, N.L., Douglas, M.R., Douglas, M.E., Fausch, K.D., 2004. Ecological and evolutionary consequences of biotic homogenization. Trends Ecol. Evol. 19, 18–24.
- Ontario Ministry of the Environment and Michigan Department of Natural Resources (OMOE and MDNR, 1992. St. Marys Area of Concern: environmental conditions and problem definitions. Remedial Action Plan—Stage 1 Report. Toronto, Canada, Ontario Ministry of the Environment.
- Pratt, T.C., Quinlan, H.R., Czypinski, G.D., Schram, S.T., Gorman, O.T., 2009. Inshore fish community: ecological interactions. In: Gorman, O.T., Ebener, M.P. (Eds.), The state of Lake Superior in 2005: Great Lakes Fish. Comm. Spec. Pub, vol. 09-01. pp. xx-xx.
- Quinn, J.W., Kwak, T.J., 2003. Fish assemblage changes in an Ozark River after impoundment: a long-term perspective. Trans. Am. Fish. Soc. 132, 110–119.
- Randall, R.G., Minns, C.K., 2002. Comparison of a Habitat Productivity Index (HPI) and an Index of Biotic Integrity (IBI) for measuring the productive capacity of fish habitat in nearshore areas of the Great Lakes. J. Great Lakes Res. 28, 240–255.
- Ripley, M.P., Arbic, B., Zimmerman, G., 2011. Environmental history of the St. Marys River. J. Great Lakes Res. 37 (Supplement 2), 5–11.
- Ross, S.T., Matthews, W.J., Echelle, A.A., 1985. Persistence of stream fish assemblages: effects of environmental change. Am. Nat. 126, 24–40.
- Schaeffer, J.S., Fielder, D.G., Godby, N., Bowen, A., O'Connor, L., Parrish, J., Greenwood, S., Chong, S., Wright, G., 2011. Long-term trends in the St. Marys River open water fish community. J. Great Lakes Res. 37 (Supplement 2), 70–79.
- Schleen, L.P., Christie, G.C., Heinrich, J.W., Bergstedt, R.A., Young, R.J., Morse, T.J., Lavis, D. S., Bills, T.D., Johnson, J.E., Ebener, M.P., 2003. Development and implementation of an integrated program for control of sea lampreys in the St. Mary's River. J. Great Lakes Res. 29 (Suppl. 1), 677–693.
- Schulz, E.J., Hoyer, M.V., Canfield, D.E., 1999. An index of biotic integrity: a test with limnological and fish data from sixty Florida Lakes. Trans. Am. Fish. Soc. 128, 564–577.
- Scott, W.B., Crossman, E.J., 1998. Freshwater Fishes of Canada. Galt Publishing Ltd., Oakville, Ontario.
- Siegel, S., Castellan, N.J., 1988. Nonparametric Statistics for the Behavioral Sciences, 2nd edition. McGraw-Hill, New York.
- Valere, B.G., 1996. Productive capacity of littoral habitats in the Great Lakes: field sampling procedures (1988–1995). Can. Mans. Rep. Fish. Aquat. Sci, No. 2384.
- Vanderploeg, H.A., Nalepa, T.F., Jude, D.J., Mills, E.L., Holeck, K., Liebig, J.R., Grigorovich, I. A., Ojaveer, H., 2002. Dispersal and emerging ecological impacts of Ponto-Caspian species in the Laurentian Great Lakes. Can. J. Fish. Aquat. Sci. 59, 1209–1228.
- Ward, J.V., 1998. Riverine landscapes: biodiversity patterns, disturbance regimes and aquatic conservation. Biol. Cons. 83, 269–278.
- Winston, M.R., Taylor, C.M., 1991. Upstream extirpation of four minnow species due to damming of a prairie stream. Trans. Am. Fish. Soc. 120, 98–105.
- Wolter, C., Minow, J., Vilcinskas, A., Grosch, U.A., 2000. Long-term effects of human influence on fish community structure and fisheries in Berlin waters: an urban water system. Fish. Manage. Ecol. 7, 97–104.
- Yoder, C.O., Smith, M.A., 1999. Using fish assemblages in a state biological assessment and criteria program: essential concepts and considerations. In: Simon, T.P. (Ed.), Assessing the sustainability and biological integrity of water resources using fish assemblages. CRC Press, Boca Raton, Florida, pp. 17–56.
- Zar, J.H., 1999. Biostatistical analysis, fourth ed. Prentice-Hall, Upper Saddle River, New Jersey.

Appendix 2a: A synthesis of sport fishing activity in the St. Marys River (Godby et al. 2019)

A synthesis of sport fishing activity In the St. Marys River

May through October 2017

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Introduction

The St. Marys River provides world-class fishing opportunities for a variety of species. The river is unique in the diversity of recreational sport fisheries and the magnitude of the fishery. In order to quantify the sport fisheries, the Michigan Department of Natural Resources (MDNR) and the Ontario Ministry of Natural Resources and Forestry (OMNRF) undertook an open water creel survey of sport anglers in 2017. The survey covered both Ontario and Michigan waters from the compensating works downstream to Lake Huron at DeTour. This is only the second whole-river creel study of the St. Marys River, with the first conducted in 1999-2000 (Fielder et al. 2002). Partial river creel surveys were conducted during a number of years, but extrapolation to whole-river estimates proved difficult (Greenwood et al. 2011). The 2017 creel survey was also accompanied by a fish community gillnet survey of the river that same year by the St. Marys River Fisheries Task Group (SMRFTG) and its member agencies (O'Connor et al. 2019).

This survey is intended to quantify angler harvest and effort in the St. Marys River and identify any trends in the river recreational sport fishery. This information will supplement other individual management agency data, and the joint river-wide fish community gillnet survey, and it will be used to help inform management of the river. Fish Community Objectives for the St. Marys River are being developed, and common sport fishing regulations for this binational waterbody remain a goal of the St. Marys River Fisheries Task Group. These surveys equip managers with data necessary to make informed decisions about the world-class fisheries the river supports.

The St. Marys River is the connecting channel between Lakes Superior and Huron (Figure 1). The river flows southeasterly from Lake Superior's Whitefish Bay for 112 km and empties into northern Lake Huron at DeTour, Michigan, and into the North Channel of Lake Huron in Ontario at St. Joseph Island. The river holds the international boundary line between Ontario, Canada and Michigan, United States of America. Although the fish community has been described as percid dominated (Duffy and Batterson 1987, Ryder and Kerr 1978), the river supports a diverse mix of migratory and resident warm, cool, and coldwater species (St. Marys River RAP Team 1992). Resident species of interest to anglers include Northern Pike (Esox Lucius), Smallmouth Bass (Micropterus dolomieu), Walleye (Sander vitreus), and Yellow Perch (Perca flavescens). These species are well distributed throughout the river and its numerous bays. These four species, along with Cisco (Coregonus artedii) will be highlighted throughout this report. The river, main rapids in Sault Ste. Marie, the Little Rapids adjacent to Sugar Island, and several St. Marys tributaries are also seasonally used by salmonid species, providing popular fisheries. Aquatic habitats vary throughout the river's length, often changing abruptly from one habitat type to another. Habitats are generally characterized as open water, embayments, sand and gravel beaches, rapids and emergent wetlands (Duffy and Batterson 1987). The lower reaches of the river, Potagannissing Bay, Raber Bay, and Lake Munuscong, are more lacustrine in form and at least seasonally contain feeding aggregations of Cisco or migrating Pacific salmons and Rainbow Trout (Oncorhynchus mykiss) resulting in short term, seasonal fisheries. Considerable shoreline and channel alteration and hardening, dredging, and flow control and flow redirection have occurred over the past two centuries. Both the distribution of habitat types and anthropogenic stresses influence the species angler target in the various river reaches and the intensity and seasonality of fishing effort.

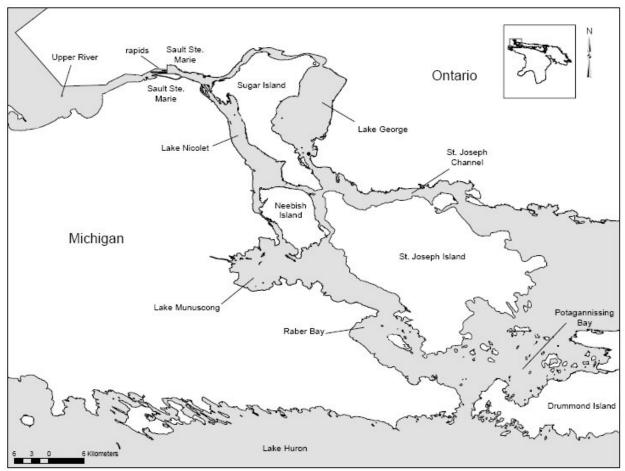


Figure 1. Map of the St. Marys River Methods

Creel Methodology

In 2017 the MDNR, in conjunction with Ontario Ministry of Natural Resources and Forestry (OMNRF), the Environmental Protection Agency (EPA), and the United States Fish and Wildlife Service (USFWS) performed a river-wide survey of the St. Mary's River employing MDNR creel census methodology. The results in this report are derived from the survey performed during the open water season (May 1st - October 31st). Due to hiring delays creel census did not begin in Michigan until May10th.

The creel survey was based on a stratified design using three-stage sampling within the strata: i) days; ii) shifts; and iii) count times, and followed the methods detailed by Lockwood et al. (1999) and Su and Clapp (2013). Strata included 'sites' (Figure 2) fished by month, by day-type (weekday-weekend/holiday), and by mode of fishing (e.g, boat, pier/dock, shore). Catch and effort estimates were made for each stratum and then combined to give monthly and seasonal figures. The estimation sites were:

- Site 403; the northernmost sampling location. The area includes the St. Mary's River rapids on the Canadian side of the river, north of the Locks in Sault Ste. Marie, USA.
- Site 209; the river from the Locks in Sault Ste. Marie moving south to include Lake Nicolet and the middle Neebish Channel ending at Oak Point on Neebish Island and east to Harwood Point.
- Site 208; the river branch north of Sugar Island and all of Lake George, including Echo Bay and ending on a line north from Harwood Point to the Canadian mainland.
- Site 405; the river south of Lake George to the North Channel of Lake Huron, including the St. Joseph Channel.

- Site 207; the river south of Neebish Island and west of St. Joseph Island to the village of Detour, including Munuscong Lake and Raber Bay.
- Site 210; the river east of the village of Detour including Potagannissing Bay extending from Milford Haven on St. Joseph Island to Chippewa Point on Drummond Island.

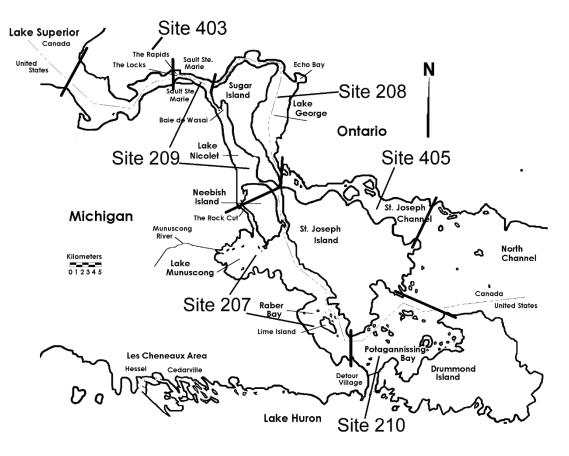


Figure 2. Creel Survey sites sampled in 2017.

Both weekend days and three randomly selected weekdays were sampled each week. The entire angling day from dawn to dusk was covered in each month. This was accomplished by breaking each day into two 8-hour work shifts, then randomly selecting one shift to be worked. The first shift began at daylight and ended in the afternoon; the second shift began in the afternoon and ended at sunset. Shift hours varied by month due to varying length of daylight among months. Each individual clerk was responsible for sampling more than one area, thus the interview site for each clerk was also randomly selected for each shift. Two types of data were collected for each area sampled: angler party interviews for catch rates and angler (boat, pier and shore) counts for effort. An angler party was defined as one or more anglers who fished together.

Two seasonal clerks were employed by the MDNR and two seasonal clerks were employed by the OMNR to perform the survey. All four clerks were trained by MDNR Fisheries personnel at the beginning of the field season to ensure the consistency of data collected. Data was submitted monthly by all clerks for review by the MDNR.

Clerks interviewed as many anglers as possible by encountering boats that returned to access sites or shore and pier anglers that fished during a scheduled shift. If the boater did not fish, that was recorded on the form as a non-fishing party and the interview was ended. If fishing did take place, anglers were queried as to their mode of fishing (i.e., boat, pier, shore), where they fished (site and grid number), how long they fished, what they fished for, the number of lines fished, how often they fished the area, the numbers (by species) of fish they caught and kept, the numbers (by species) of fish released, and the number of fishing trips they made or intended to make that day. Additional data were collected for one member of each party such as age and sex, zip code, and the types of angling method used (casting, still fishing, trolling, etc.).

Fishing effort was determined through instantaneous counts of boats and fishing piers made from airplanes. Five flights were made each week at randomly selected starting times; one each weekend day, and one on each of three randomly selected week days. Flights were also randomly started at the northern and southern end of the route. The proportion of boaters interviewed by creel clerks who indicated they were not fishing was used to adjust the aerial counts for non-fishing effort. Creel clerks also recorded a ratio of anglers to nonanglers that were present on fishing piers at randomly selected times of day. This angler ratio was applied to the airplane count of total people present on any piers. Shore anglers present at Site 403 (the Canadian rapids) were counted by the clerk twice daily at randomly assigned times on scheduled shifts.

Effort estimates were made for each site and mode by month. Harvest estimates were made for each site and mode by month for all fish species observed in the harvest by creel clerks. Catch estimates were made for each site and mode by month and included numbers harvested and numbers of legal-sized fish released for those selected species. Standard mathematical formulas for creel survey (Lockwood et al. 1999) were used to calculate all estimates. Three measures of fishing effort were calculated: angler hours, angler trips, and angler days. An angler trip is one completed fishing excursion and an angler day is composed of one or more fishing excursions during a 24-hour period. Uncertainty estimates for all catch and effort estimates in this report are defined as two standard errors of their mean estimates (2 times the square root of the variance for an estimate).

Creel clerks also collected biological data from harvested fish (total length and weight and fin clip information) encountered during on-site angler interviews. Dorsal fin spines or rays were also collected for certain species for age estimation. Monthly target sample sizes for age analysis were based on a minimum number needed to provide a reasonable representation of the age structure of the harvest each month, balanced with the logistical feasibility of the creel clerk to collect biological data samples without negatively impacting angler interview numbers.

Results

Interview and Angling Effort

A total of 3,558 anglers were interviewed on the St. Marys River by the creel clerks from May 1-October 31, 2017.

Total annual effort for the entire river was 232,921 angler hours in 2017 (Table 1). This does not include site 403 (the Rapids), since it was not surveyed in all years. This is the lowest value of the seven years for which we have estimated effort, down from a high of 565,095 angler hours in 2001. In 2017, site 210 (Potagannissing Bay) had the most effort, followed by site 207 (Munuscong and Raber Bays) and site 208 (Lake George). Anglers targeting walleye contributed a total of 118,484 hours of angling effort (Table 2), or approximately 51% of the effort on the river.

Table 1. Estimated recreational effort (in hours) for all species from open-water sport fisheries in the St. Marys River 1999-2001, 2005-2007, and 2017. Italics denotes data obtained through extrapolation methods described in the Methods section (Greenwood et al. 2011). Note that site 403 (Rapids) is not included, as it was not surveyed every year. Two standard errors are in parentheses.

Year	207	208	209	210	404	405	Total
1999	112,283	96,732	68,441	140,743	58,561	65,307	556,399
	(19,570)	(16,256)	(11,010)	(27,674)	(11,454)	(12,611)	(42,820)
2000	93, 301	60,816	60,564	131,107	55,616	61,572	462,976
	(15,420)	(12,794)	(11,511)	(20,871)	(46,434)	(76,874)	(183,904)
2001	124,823	97,111	76,694	123,878	67,671	74,918	565,095
	(28,135)	(17,919)	(14,401)	(17,646)	(64,802)	(106,689)	(249,592)
2005	68,289	51,245	54,378	131,887	47,410	74,105	427,314
	(12,840)	(10,260)	(232,480)	(35,124)	(60,886)	(14,371)	(365,960)
2006	93,025	70,944	84,845	152,254	58,378	*52,984	512,430
	(24,502)	(14,685)	(15,437)	(36,035)	(72,178)	*(9,645)	(172,483)
2007	139,310	35,273	71,430	183,668	62,276	45,112	537,069
	(34,103)	(8,859)	(313,367)	(60,215)	(107,502)	(11,057)	(535,103)
2017	45,491	33,122	24,332	98,966	Not	31,010	232,921
	(26,153)	(17,306)	(9,559)	(47,311)	Surveyed	(12,455)	(112,784)

* In 2006, no interviews were conducted for Site 405, see Greenwood et al. (2011) for methods.

**In 2017, site 404 was not surveyed.

Table 2. Estimated targeted effort (in hours) for selected species for the open-water sport fisheries in the St. Marys River 1999-2001, 2005-2007, and 2017. Two standard errors are in parentheses.

	Year											
	1999	2000	2001	2005	2006	2007	2017					
Walleye	126,988	119,122	161,526	168,031	168,333	200,006	118,484					
	(25,083)	(24,654)	(28,699)	(53,463)	(37,817)	(63,957)	(16,713)					
Yellow Perch	89,238	60,607	78,869	32,414	58,191	65,326	39,885					
	(18,255)	(16,698)	(20,924)	(14,026)	(23,031)	(30,758)	(6,648)					
Salmonids	122,280	56,988	79,529	76,419	49,241	66,488	29,635					
	(20,238)	(14,007)	(21,306)	(43,360)	(14,674)	(44,801)	(3,985)					

Harvest

Twenty-four species were reported as caught during the 2017 St. Marys River creel survey. Harvest estimates for 16 of these species are presented in Table 3, with some rarely encountered species grouped together in the "other" category. White Bass (*Morone chrysops*) was reported as harvested in 2017 as it had been previously, but the invasive White Perch (*Morone Americana*) was not reported during this survey. Estimated harvest numbers and harvest rates by species for the survey series are reported in Appendix 1, Table 1.

			All S	Sites				
				М	onth			
Species	Harvest per hour	May	June	July	August	September	October	Season
Atlantic Salmon	0.0042	28	124	492	275	99	7	1,025
	(0.0064)	(50)	(133)	(275)	(197)	(90)	(12)	(756)
Brown Trout	0.0000	0	0	0	2	0	0	2
	(0.0000)	(0)	(0)	(0)	(5)	(0)	(0)	(5)
Channel Catfish	0.0003	0	38	0	0	31	0	69
	(0.0012)	(0)	(76)	(0)	(0)	(60)	(0)	(136)
Chinook Salmon	0.0006	0	16	9	18	73	22	138
	(0.0018)	(0)	(32)	(18)	(36)	(87)	(36)	(209)
Coho Salmon	0.0003	63	0	0	0	1	0	64
	(0.0009)	(103)	(0)	(0)	(0)	(2)	(0)	(105)
Freshwater Drum	0.0004	0	75	0	0	0	20	95
	(0.0016)	(0)	(149)	(0)	(0)	(0)	(38)	(187)
Cisco	0.1313	0	0	32,267	0	0	0	32,267
	(0.1545)	(0)	(0)	(18,255)	(0)	(0)	(0)	(18,255)
Lake Trout	0.0000	0	0	0	0	9	0	9
	(0.0002)	(0)	(0)	(0)	(0)	(18)	(0)	(18)
Lake Whitefish	0.0245	457	457	4,962	25	73	37	6,011
	(0.0409)	(600)	(515)	(3,506)	(26)	(117)	(71)	(4,836)
Northern Pike	0.0161	899	1,543	755	302	430	39	3,968
	(0.0329)	(1,384)	(635)	(938)	(310)	(539)	(80)	(3,885)
Other	0.0001	0	16	0	0	16	0	32
	(0.0005)	(0)	(32)	(0)	(0)	(31)	(0)	(62)
Pink Salmon	0.0005	0	0	0	30	93	0	123
	(0.0013)	(0)	(0)	(0)	(44)	(114)	(0)	(158)
Pumpkinseed	0.0009	0	161	0	59	0	0	220
	(0.0027)	(0)	(244)	(0)	(76)	(0)	(0)	(320)
Rainbow Trout	0.0033	241	190	218	49	80	29	807
	(0.0074)	(267)	(193)	(210)	(67)	(74)	(58)	(870)
Smallmouth Bass	0.0125	174	1,262	1,228	88	321	0	3,073
	(0.0286)	(294)	(1,318)	(1,121)	(101)	(545)	(0)	(3,379)
Walleye	0.0568	3,076	4,629	2,718	1,167	1,495	878	13,963
	(0.1174)	(3,998)	(3,865)	(3,019)	(921)	(1,168)	(893)	(13,865)
Yellow Perch	0.1597	942	2,220	2,963	2,043	18,145	12,928	39,241
	(0.2510)	(1,388)	(2,252)	(2,515)	(2,518)	(10,492)	(10,492)	(29,657)
Angler hours		23,718	55,467	82,113	29,882	40,604	13,991	245,775
		(17,400)	(30,830)	(34,796)	(11,805)	(14,857)	(8,445)	(118,134)
Angler trips		5,692	13,326	19,791	6,428	10,018	3,115	58,370
		(4,029)	(5,731)	(8,901)	(3,275)	(3,822)	(1,931)	(27,688)
Angler Days		5,293	12,838	18,353	6,146	9,714	3,060	55,404
		(3,740)	(5,546)	(8,368)	(3,163)	(3,738)	(1,902)	(26,456)

Table 3. Estimated harvest per hour and total number harvested by species for each month. Angler effort (angler hours, trips, and days) for the St. Marys River system (Michigan and Ontario), for all sites including Site 403 (Rapids) and all non-charter modes of sportfishing, for each month in 2017. Two standard errors are in parentheses.

Harvest numbers for the survey series are presented in Table 4 below for Cisco, Northern Pike, Smallmouth Bass, Walleye, and Yellow Perch. Site-specific harvest numbers for each species are detailed in Appendix 2. Total catch by species, including released fish, is listed in Appendix 3.

Table 4. Estimated harvest (numbers of fish) for Cisco, Northern Pike, Smallmouth Bass, Walleye, and Yellow Perch from open-water sport fisheries in the St. Marys River 1999-2001 and 2005-2007. Two standard errors are in parentheses.

		Northern	Smallmouth		Yellow
Year	Cisco	Pike	Bass	Walleye	Perch
1999	31,258	5,408	1,188	9,890	62,646
	(40,040)	(5,170)	(1,797)	(8,255)	(32,274)
2000	113,621	12,402	3,235	17,064	86,098
	(182,114)	(17,744)	(9,001)	(17,768)	(100,284)
2001	131,662	14,336	3,653	39,568	91,120
	(199,643)	(22,768)	(10,371)	(30,643)	(96,696)
2005	48,163	1,547	4,216	32,134	84,097
	(92,339)	(3,516)	(10,329)	(24,882)	(96,889)
2006	168,988	14,894	5,322	38,743	118,214
	(211,690)	(18,288)	(10,567)	(46,952)	(150,617)
2007	158,141	4,231	4,030	60,733	125,391
	(372,281)	(5,322)	(7,691)	(56,668)	(180,500)
2017	32,267	3,968	3,073	13,963	39,241
	(18,255)	(3,885)	(3,379)	(13,865)	(29,657)

Species

Harvest Rates

River-wide, species-specific harvest rates (harvest per hour) based on total effort for all species, Cisco, Northern Pike, Smallmouth Bass, Walleye, and Yellow Perch - for the survey series are presented in Table 5. These rates are presented for all species by site specific locations in Appendix 2 (Tables 1-7).

Table 5. Mean annual harvest per hour for Cisco, Northern Pike, Smallmouth Bass, Walleye, and Yellow Perch (based on total effort) from open-water sport fisheries in the St. Marys River (from all sites including Potagannissing Bay), 1999-2001, 2005-2007, and 2017. Two standard errors are in parentheses.

Year	Cisco	Northern Pike	Smallmouth Bass	Walleye	Yellow Perch
1999	0.0562	0.0097	0.0021	0.0178	0.1126
	(0.0721)	(0.0093)	(0.0032)	(0.0149)	(0.0586)
2000	0.1631	0.0284	0.0069	0.0376	0.1314
	(0.1688)	(0.1626)	(0.1167)	(0.1606)	(0.3123)
2001	0.1790	0.0269	0.0055	0.0687	0.1462
	(0.1766)	(0.1685)	(0.1568)	(0.1872)	(0.3438)
2005	0.0708	0.0037	0.0072	0.0747	0.1297
	(0.2116)	(0.1457)	(0.1302)	(0.2265)	(0.3742)
2006	0.2303	0.0305	0.0108	0.0830	0.1705
	(0.2176)	(0.1775)	(0.1794)	(0.2294)	(0.4210)
2007	0.1587	0.0093	0.0080	0.0997	0.1686
	(0.5180)	(0.1829)	(0.2191)	(0.3410)	(0.3787)
2017	0.1313	0.0161	0.0125	0.0568	0.1597
	(0.1545)	(0.0329)	(0.0286)	(0.1174)	(0.2510)

Biological Summary of Angler Harvest

In 2017, a total of 745 fish were sampled by creel clerks. Biological data collected from those samples are summarized in Table 6. Aging structures (e.g., scales or fin spines) were not collected for all samples because a much larger sample size of fish were aged during the 2017 fish community gillnet survey. That companion survey was fishery-independent, and provides a much better view of the age and size structure of the population. See O'Connor et al. (2019) for the additional biological data.

Table 6. Summary of biological data collected from the St. Marys River during the open water sport fishery of 2017, by capture sites. N = sample size and appears in parentheses if different than reported. Data for species with a sample size smaller than 5 were not included in this table. A complete list of biological data for 2017 and previous creel surveys can be found in Appendix 4.

				Mean	Mean Length	Mean Wt
Species	Year	Capture sites	Ν	Age	(cm)	(g)
Atlantic Salmon	2017	208, 209, 403	117	2.2 (41)	56.6	2174 (116)
Chinook Salmon	2017	208, 403	6	2.8 (5)	78.1	5934
Cisco	2017	209, 210	92		38.5	605 (91)
Northern Pike	2017	207, 208, 209, 210	74	4.6	67.6	1876
Pink Salmon	2017	208, 209, 403	31	2 (7)	50.5	1283
Rainbow Trout	2017	208, 209, 403	32	2.5	48.5	1549
Smallmouth Bass	2017	207, 208, 209, 210	57	4 (2)	39.6	1012 (32)
Walleye		207, 208, 209, 210,				
-	2017	403	157	5.3 (21)	46.3	1019 (116)
Yellow Perch	2017	207, 208, 209, 210	170	5.3 (3)	22.9	165 (155)

Angler Interview Summary

Angler Origin – Although most anglers were from Michigan and Ontario, the St. Marys River draws anglers from throughout the continent (Figure 3). Approximately 26% of the anglers were considered "local", defined as those whose reported home zip code is within 40 or 80 kilometers of the river. In addition to Michigan and Ontario, anglers were from 17 other states and 3 other provinces. American anglers comprised 68% of the anglers interviewed. Anglers hailed from as far away as Hawaii, Washington, British Columbia, and the maritime provinces.

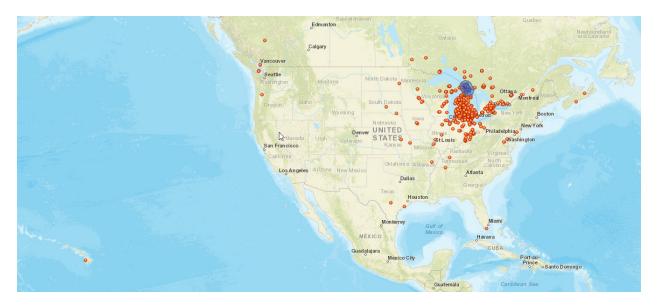


Figure 3. Map of angler origin for those fishing the St. Marys River in 2017, based on U.S. Zip/Canadian postal codes provided during the interviews.

Angling target species, angling method, mode, and party size

The St. Marys River presents diverse angling opportunities in terms of the species that can be targeted and the angling methods that can be used. Anglers targeted a variety of species throughout the season (Table 7). Walleye were the most frequently targeted species (28.2%), followed by general salmon and salmon or trout (totaling 11.5%), Rainbow Trout (11.0%) and Atlantic Salmon (10.9%). A fair percentage (14.2%) indicated they were fishing for anything.

Table 7. Percent of angling parties in the St. Marys River open water (May-Oct.) sport fishery reporting target species, by location as reported during angler interviews. N denotes number of respondents.

TARGET	207	208	209	210	403	404	405	Total
ANY	15.9%	12.3%	11.3%	17.0%	9.2%	0.0%	64.6%	14.2%
Atlantic Salmon	0.0%	4.6%	32.9%	0.0%	10.2%	0.0%	0.0%	10.9%
Chinook Salmon	0.0%	1.5%	0.7%	0.0%	2.3%	0.0%	0.0%	0.9%
Coho Salmon	0.0%	0.3%	0.0%	0.0%	0.5%	0.0%	0.0%	0.2%
Cisco	1.4%	0.0%	0.0%	3.7%	0.0%	0.0%	0.0%	0.9%
Largemouth Bass	0.0%	0.0%	0.0%	0.0%	0.3%	0.0%	0.0%	0.1%
Muskellunge	0.3%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.1%
Northern Pike	6.1%	13.8%	5.3%	8.0%	0.0%	50.0%	12.5%	6.5%
Other	2.4%	0.0%	1.8%	6.2%	4.1%	0.0%	0.0%	2.8%
Pink Salmon	0.3%	5.2%	1.8%	0.0%	1.8%	0.0%	0.0%	1.8%
Rainbow Trout	0.0%	4.3%	0.2%	0.0%	47.3%	0.0%	0.0%	11.0%
Salmon	0.0%	11.4%	0.0%	0.0%	10.7%	0.0%	0.0%	4.3%
Salmon or Trout	0.0%	1.5%	17.3%	0.0%	13.5%	50.0%	0.0%	7.4%
Smallmouth Bass	2.4%	0.9%	0.5%	6.5%	0.0%	0.0%	4.2%	1.9%
Walleye	65.1%	33.8%	19.8%	35.9%	0.3%	0.0%	18.8%	28.2%
Walleye and Perch	0.3%	0.3%	0.0%	0.6%	0.0%	0.0%	0.0%	0.2%
Yellow Perch	5.8%	9.8%	8.3%	22.0%	0.0%	0.0%	0.0%	8.6%
Ν	295	325	434	323	393	2	48	1820

The St. Marys River Rapids (site 403) continues to be a popular fishing location, where the target species changes based on seasonal migration patterns (Table 8). Rainbow Trout (steelhead) are the most sought after species in May, June, and October, but Atlantic Salmon are the preferred target in July and August.

	May	June	July	August	September	October
ANY	2%	6%	14%	9%	18%	12%
Atlantic Salmon			38%	34%	3%	
Chinook Salmon				5%	8%	
Coho Salmon					1%	2%
Largemouth Bass					1%	
Other	2%	11%	6%	2%		5%
Pink Salmon				2%	8%	
Rainbow Trout	95%	73%	14%	11%	10%	41%
Salmon	1%	3%	2%	16%	34%	10%
Salmon and Trout		7%	26%	21%	15%	29%
Walleye					1%	
N	103	70	50	56	73	41

Table 8. Percent of angler parties reporting species targeted in the St. Marys River rapids sport fishery (Site 403) by month in 2017. N denotes sample size.

Additional, customized queries of the dataset may be run from the open data portal at: <u>http://gis-midnr.opendata.arcgis.com/search?q=creel</u>.

Discussion

Fishing Effort

Overall fishing effort in the St. Marys River was 232,921 angler hours in 2017. While this is approximately 50% less than it has been in previous creel surveys, the overall magnitude of the fishery remains substantial. Based on number of angler hours, the amount of fishing effort in the St. Marys River in 2017 was 34.5% as much as all the fishing effort in the Michigan waters of Lake Huron (Table 9). Although the amount of effort is down, that level of effort tracks consistent with the Michigan waters of Lake Huron.

The amount of fishing effort in the St. Marys River is even more impressive when examined based on surface acreage compared to other popular fisheries in the region. In 2017, the St. Marys River had more fishing effort per unit surface area than Big Bay de Noc and Saginaw Bay, and compared favorably with the fishing pressure per surface area in Little Bay de Noc (Figure 4).

Table 9. Comparison of fishing effort expressed as hours in the St. Marys River (excluding site 403 - the Rapids) to the lake wide fishery in the Michigan waters of Lake Huron and for Saginaw Bay. Site 403, the St. Marys rapids is excluded.

Year	St. Marys River effort (Hours)	% as much of Lake Huron effort (Hours)	% as much of Saginaw Bay effort (hours)
1999	542,067	27.7	60.0
2000	462,976	26.7	61.2
2001	565,095	31.4	70.0
2005	427,314	32.3	57.3
2006	512,430	44.8	79.7
2007	537,069	38.7	62.7
2017	232,921	34.5	53.2

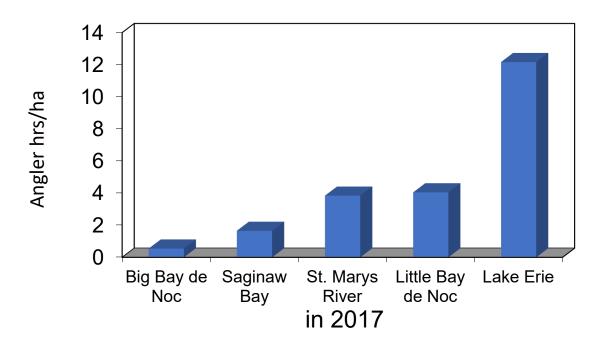


Figure 4. Angler hours per hectare in 2017 for a number of popular fishing locations in Michigan. Note that the Lake Erie estimate is for Michigan waters only.

Harvest and Harvest Rates

Cisco

Cisco harvest and harvest rates were down by approximately 80% in 2017 compared with previous years (2006, 2007), with harvest the lowest since 1999. Harvest rates were approximately the same as they were in 2005.

The Cisco fishery in the St. Marys River is often driven by an early summer feeding congregation of these fish in sites 207, 209, and 210. The feeding congregation of Cisco is keying in on emerging mayflies and can create substantial fishing effort in these areas of the river.

Cisco harvest is highly variable and is dependent upon factors such as weather conditions during the feeding congregation, as well as the timing and duration of that feeding congregation. Both harvest and harvest rate are within their historical range. The fish community gillnet survey also conducted in 2017 indicated cisco were at a low abundance for that time series (O'Connor et al. 2019). This suggests that cisco may genuinely be at a lower abundance that year and affecting the fishery.

The only notable change in fishing regulations for the river since the last creel survey were for Cisco. Ontario regulations for Cisco changed in 2008, when it went from no daily possession limit to a daily possession limit of 25 Cisco for a Sportfishing licnese or 12 Cisco for a Conservation license. Although Cisco harvest was down in 2017, it is likely not entirely attributable to the Ontario change in daily possession limit since harvest rate was also down. The SMRFTG is working towards developing common regulations between Ontario and Michigan for the remaining sport fisheries.

Northern Pike and Muskellunge

Northern Pike harvest and harvest rate are still much lower than they were historically, but are trending upwards. Coastal wetlands provide critical spawning and nursery habitat for Northern Pike. Great Lakes water levels, and therefore St. Marys River water levels, have increased since 2013. Lake Superior outflows, (St. Marys River flows) are partially controlled through the compensating gates at the head of the rapids. River flows in recent years have been higher than they've been in decades and these higher water levels have led to more spawning and nursery habitat resulting in improved pike numbers. Good numbers of Age-5 and younger pike captured in the companion survey (O'Connor et al. 2019) support the idea that the higher water levels since 2013 have helped the Northern Pike population. Higher water levels likely also improved survival of Northern Pike, as evidenced by the lowest total annual mortality rates seen in the survey series.

The mean age of Northern Pike captured in the sport fishery was 4.6, while the mean age in the fish community gillnet survey was 4.0. Mean length in the sport fishery was 676mm and was 552mm in the 2017 gillnet survey (O'Connor et al. 2019). These differences could point to gear selectivity; i.e., gill nets aren't effectively sampling older/larger Northern Pike, however it is also likely that the sport fishery selects for larger (older) pike. Northern Pike regulations differ between Michigan and Ontario. Michigan has a 610 mm (24 in) minimum size limit, while Ontario has no minimum size limit. In 2017, approximately 12% of the Northern Pike sampled during the creel survey were less than 610 mm. It is important to continue both the fishery-independent (gill net) survey and the creel survey of the sport fishery in order to get a complete picture of the fish community.

Muskellunge remain an important part of the fish community and provide a popular fishery. An estimated 214 muskies were caught in the St. Marys River during the open-water season in 2017.

Smallmouth Bass

Overall harvest of Smallmouth Bass is down slightly from the 2007 estimate, but is higher than the estimate from our last full-river creel in 1999. While harvest rate has increased by almost 67%, Smallmouth Bass are plentiful in the river and can provide a good fishery, though it remains an untapped resource. Only 1.9% of angling parties river-wide reported targeting Smallmouth Bass in 2017, with Potagannissing Bay, Site 210, having the highest percentage (6.5%) of angling parties reporting they were targeting Smallmouth Bass. Angler harvest and harvest rate remain substantially higher than in 1999, and harvest rate is above the mean 1999-2017harvest rate of 0.008.

Walleye

Walleye continue to be an important sport fish in the St. Marys River, and were the species most targeted by anglers. Overall harvest and harvest rate of Walleye in the river declined since 2007, but both are higher than they were in 1999 during our last full-river creel. The 2017 harvest rate (0.0568), however, is slightly lower than the mean harvest rate of 0.064 (1999-2017). Current estimates of harvest and harvest rate are within the historical range of this survey series.

Walleye were harvested from every creel site in 2017, but by far the most harvest (50%) came from site 207-Lake Munuscong and Raber Bay. That was followed by site 210-Potagannissing Bay (20.9%), site 208-Lake George (12.6%), and site 405-St. Joseph Channel (11.6%).

Walleye have been stocked in the St. Marys River since about 1985 at varying levels and by different entities. It is unknown, however, to what extent that stocking contributes to the fishery. To answer that question and others, a 10-year Walleye Stocking and Evaluation Plan was developed by the St. Marys River Fisheries Task Group and was implemented 2009-2018. The plan called for the annual stocking of 290,000 to 400,000 spring fingerling OTC-marked Walleye in different reaches and follow-up evaluations each fall. 2018 is the last year of evaluation under that plan, and results are expected soon to help inform the Walleye stocking strategy for the river.

Yellow Perch

Yellow Perch support a popular fishery in the St. Marys River, with 8.6% of angling parties targeting Yellow Perch river-wide throughout the open water season. They were the target species of 22% of the angling parties in site 210-Potagannissing Bay, second only to Walleye in popularity at that site. Although overall harvest of Yellow Perch is down in 2017 compared to previous years, harvest rate has remained relatively stable over the survey series.

Economic Value

The 2016 National Survey of Fishing, Hunting, and Wildlife-Associated Recreation, conducted by the U.S. Fish and Wildlife Service, estimated daily expenditures for hunters and anglers (USDI 2016). For Great Lakes anglers in 2016, USDI 2016 reports that Great Lakes anglers spend \$153 per day on fishing trip-related expenses. The 2017 survey of the St. Marys River estimates 55,404 angler days of fishing effort were spent on the river, with a value of approximately \$8.5 million US Dollars.

Supplemental Questions

MDNR was considering a change to the statewide regulations for Yellow Perch at the time of this survey. The daily possession limit for Yellow Perch at the time of the survey was 50; and the MDNR was considering changing the daily possession limit to 25 perch. In order to gage public opinion about the potential change, a supplement question was included in the creel survey. The following questions was asked: Would you support a decreased bag limit of 25 Yellow Perch?

Angler responses indicated support for the regulation change. In the Upper St. Marys River, 72% of those respondents with a definite opinion supported the change (28% did not support). In the Lower St. Marys River, 93% of those with a definite opinion supported the reduced bag limit. Overall, anglers in the St. Marys River supported going to a 25 fish bag limit for Yellow Perch. The reduced bag limit was subsequently approved and will be implemented on a statewide basis in 2019.

Recommendations

The river-wide creel survey should be conducted every five years in conjunction with the fish community gillnet survey on the same frequency. Recreational harvest regulations should be aligned between Ontario and Michigan waters of the river.

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The St. Marys River Fisheries Task Group is organized under the Great Lakes Fishery Commission's Lake Huron Committee, and coordinates fisheries management among the various agencies with jurisdiction over the St. Marys River (Fielder 2002).

References

- Duffy, W.E., and T. R. Batterson. 1987. The St. Marys River, Michigan: an ecological profile. USFWS Biological Report 85 (7.10).
- Fielder, D.G., A.K. Bowen, K.J. Gebhardt, and S.J. Greenwood. 2002. Harvest of fishes in the St. Marys River, May, 1999 through March 2000. Great Lakes Fishery Commission, Miscellaneous publication.

- Greenwood, S.J., D.G. Fielder, N. Godby, and T. Kolb. 2011. A synthesis of sport fishing activity in the St. Marys River, 1999-2001 and 2005-2009. Great Lake Fishery Commission, Miscellaneous Publication.
- Lockwood, R.N., D. Benjamin, and J.R. Bence. 1999. Estimating angling effort and catch from Michigan roving and access site angler survey data. Michigan Department of Natural Resources, Fisheries Research Report 2044, Ann Arbor, Michigan.
- O'Connor, L.M., A. Bowen, S. Chong, D.G. Fielder, N. Godby, R. Aikens. 2019. Population dynamics of the St. Marys River fish community 1975-2017. In prep.
- Ryder, R.A., and K.R. Keerr. 1978. The adult walleye in the percid community a niche definition based on feeding behavior and food specificity. Am. Fish. Soc. Spec. Publi. 11:39-51.
- Su, Z., and D. Clapp. 2013. Evaluation of sample design and estimation methods for Great Lakes angler surveys, Transactions of the American Fisheries Society, 142:234-246.
- USDI. 2016. 2016 National survey of fishing, hunting, and wildlife-associated recreation. U.S. Department of the Interior, Fish and Wildlife Service, and U.S. Department of Commerce, U.S. Census Bureau. Available from: <u>https://wsfrprograms.fws.gov/subpages/nationalsurvey/nat_survey2016.pdf</u>. [Accessed 2/2019].

Appendixes

Appendix 1, Table 1. Estimated species harvest numbers and harvest rate (in italics) from open-water sport fisheries in the St. Marys River, 1938 to 2017.

Species	1938	1987	1991	1999	2000	2001	2005	2006	2007	2017
Atlantic Salmon	0	6	64	509	95	787	0	716	2,039	1,025
	0	<0.0001	0.0001	0.0009	0.0002	0.0014	0	0.0014	0.0038	0.0042
Bluegill				107	0	0	0	0	0	
				0.0002	0	0	0	0	0	
Brown Trout										2
										0
Channel Catfish				109	5	12	131	0	13	69
				0.0002	0	0	0.0003	0	0	0.0003
Chinook Salmon	0	4,662	469	6 249	5,707	6,785	1,619	3,632	4,042	138
	0	0.0062	0.0008	0.0112	0.0123	0.012	0.0038	0.0071	0.0075	0.0006
Cisco	289	141,386	14,528	31,258	113,620	131,662	48,163	168,988	158,141	32,267
	0.15	0.1880	0.0244	0.0562	0.2454	0.233	0.1127	0.3298	0.2945	0.1313
Coho Salmon				381	65	42	129	104	321	64
				0.0007	0.0001	0.0001	0.0003	0.0002	0.0006	0.0003
Freshwater Drum				0	19	0	1,180	1,729	1,168	95
				0	0	0	0.0028	0.0034	0.0022	0.0004
Lake Trout				1	0	0	162	0	454	9
				0	0	0	0.0004	0	0.0008	0
Lake whitefish	16	25,187	204	19,769	13,154	16,594	17,877	37,880	50,973	6,011
	0.008	0.0335	0.0003	0.0355	0.0284	0.0294	0.0418	0.0739	0.0949	0.0245
Largemouth Bass				114	202	51	0	0	0	0
				0.0012	0.0004	0.0001	0	0	0	0

Muskellunge				34	8	56	110	0	0	0
				0.0001	0	0.0016	0.0003	0	0	0
Northern Pike	184	20,965	26,116	5,408	12,402	14,336	1,547	14,894	4,231	3,968
	0.09	0.0279	0.0438	0.0097	0.0268	0.0254	0.0036	0.0291	0.0079	0.0161
Other				1,124	995	2,427	138	338	4,832	32
				0.002	0.0021	0.0043	0.0003	0.0007	0.009	0.0001
Pink Salmon	0	5.699	17,573	2,073	1,899	5,042	1,437	3,719	2,743	123
	0	0.0076	0.0295	0.0037	0.0041	0.0089	0.0034	0.0073	0.0051	0.0005
Pumpkinseed				161	0	0	175	0	1	220
				0.0003	0	0	0.0004	0	0	0.0009
Rainbow Trout	13	1,990	192	380	133	89	220	449	359	807
	0.007	0.0026	0.0003	0.0007	0.0003	0.0002	0.0005	0.0009	0.0007	0.0033
Rock Bass	166	13,708	19,718	70	105	0	720	428	448	0
	0.08	0.0182	0.0311	0.0003	0.0002	0	0.0017	0.0008	0.0008	0
Round whitefish				516	1,651	0	1,348	1,416	1,603	
				0.0009	0.0036	0	0.0032	0.0028	0.003	
Smallmouth Bass	3	2,779	9,497	1,188	3,235	3,653	4,216	5,322	4,030	3,073
	0.002	0.0036	0.0159	0.0032	0.007	0.0065	0.0099	0.0104	0.0075	0.0125
Walleye	102	25,602	26,435	9,898	17,064	39,568	32,134	38,743	60,733	13,963
	0.05	0.0340	0.0443	0.0178	0.0369	0.07	0.0752	0.0756	0.1131	0.0568
White bass					0	127	280	1,396	70	
					0	0.0002	0.0007	0.0027	0.0001	
White perch					0	229	0	0	578	
					0	0.0004	0	0	0.0011	
Yellow Perch	2,465	316,436	91,019	62,646	86,098	91,120	84,097	118,214	125,391	39,241
	1.25	0.4207	0.1526	0.1126	0.186	0.1612	0.1968	0.2307	0.2335	0.1597

Appendix 2, Table 1. Estimated harvest per hour, number harvested, and effort (angler hours, trips, and days) of sport fishing for all sites combined on the St. Marys River, by all modes (non-charter) in 2017. Area covered is from the compensating works at the head of the rapids to DeTour (including Potagannissing Bay). Two standard errors in parentheses.

				All Sites	6			
				М	onth			
Species	Harvest per hour	May	June	July	August	September	October	Season
Atlantic Salmon	0.0042	28	124	492	275	99	7	1,025
	(0.0064)	(50)	(133)	(275)	(197)	(90)	(12)	(756)
Brown Trout	0.0000	0	0	0	2	0	0	2
	(0.0000)	(0)	(0)	(0)	(5)	(0)	(0)	(5)
Channel Catfish	0.0003	0	38	0	0	31	0	69
	(0.0012)	(0)	(76)	(0)	(0)	(60)	(0)	(136)
Chinook Salmon	0.0006	0	16	9	18	73	22	138
	(0.0018)	(0)	(32)	(18)	(36)	(87)	(36)	(209)
Coho Salmon	0.0003	63	0	0	0	1	0	64
	(0.0009)	(103)	(0)	(0)	(0)	(2)	(0)	(105)
Freshwater Drum	0.0004	0	75	0	0	0	20	95
	(0.0016)	(0)	(149)	(0)	(0)	(0)	(38)	(187)
Cisco	0.1313	0	0	32,267	0	0	0	32,267
	(0.1545)	(0)	(0)	(18,255)	(0)	(0)	(0)	(18,255)
Lake Trout	0.0000	0	0	0	0	9	0	9
	(0.0002)	(0)	(0)	(0)	(0)	(18)	(0)	(18)
Lake Whitefish	0.0245	457	457	4,962	25	73	37	6,011
	(0.0409)	(600)	(515)	(3,506)	(26)	(117)	(71)	(4,836)
Northern Pike	0.0161	899	1,543	755	302	430	39	3,968
	(0.0329)	(1,384)	(635)	(938)	(310)	(539)	(80)	(3,885)
Other	0.0001	0	16	0	0	16	0	32
	(0.0005)	(0)	(32)	(0)	(0)	(31)	(0)	(62)
Pink Salmon	0.0005	0	0	0	30	93	0	123
	(0.0013)	(0)	(0)	(0)	(44)	(114)	(0)	(158)
Pumpkinseed	0.0009	0	161	0	59	0	0	220
	(0.0027)	(0)	(244)	(0)	(76)	(0)	(0)	(320)
Rainbow Trout	0.0033	241	190	218	49	80	29	807
	(0.0074)	(267)	(193)	(210)	(67)	(74)	(58)	(870)
Smallmouth Bass	0.0125	174	1,262	1,228	88	321	0	3,073
	(0.0286)	(294)	(1,318)	(1,121)	(101)	(545)	(0)	(3,379)
Walleye	0.0568	3,076	4,629	2,718	1,167	1,495	878	13,963
	(0.1174)	(3,998)	(3,865)	(3,019)	(921)	(1,168)	(893)	(13,865)
Yellow Perch	0.1597	942	2,220	2,963	2,043	18,145	12,928	39,241
	(0.2510)	(1,388)	(2,252)	(2,515)	(2,518)	(10,492)	(10,492)	(29,657)
Angler hours		23,718	55,467	82,113	29,882	40,604	13,991	245,775
		(17,400)	(30,830)	(34,796)	(11,805)	(14,857)	(8,445)	(118,134)
Angler trips		5,692	13,326	19,791	6,428	10,018	3,115	58,370
		(4,029)	(5,731)	(8,901)	(3,275)	(3,822)	(1,931)	(27,688)
Angler Days		5,293	12,838	18,353	6,146	9,714	3,060	55,404
		(3,740)	(5,546)	(8,368)	(3,163)	(3,738)	(1,902)	(26,456)

Appendix 2, Table 2. Estimated harvest per hour, number harvested, and effort (angler hours, trips, and days) of sport fishing for the area from Sweets Point to the Neebish Island Ferry (includes the Michigan and Ontario waters of Raber Bay, Munuscong Lake, and Neebish Channel), by all modes (non-charter) in 2017. Two standard errors in parentheses.

				Site 207	7			
				N	lonth			
Species	Harvest per hour	May	June	July	August	September	October	Season
Channel Catfish	0.0013	0	38	0	0	23	0	61
	(0.0046)	(0)	(76)	(0)	(0)	(44)	(0)	(121)
Freshwater Drum	0.0021	0	75	0	0	0	20	95
	(0.0071)	(0)	(149)	(0)	(0)	(0)	(38)	(187)
Northern Pike	0.0100	144	95	76	8	118	12	453
	(0.0271)	(298)	(142)	(109)	(16)	(120)	(24)	(709)
Smallmouth Bass	0.0015	0	0	68	0	0	0	68
	(0.0041)	(0)	(0)	(107)	(0)	(0)	(0)	(107)
Walleye	0.1543	2,482	2,402	1,091	348	572	123	7,018
	(0.2493)	(3,261)	(1,905)	(628)	(204)	(373)	(150)	(6,520)
Yellow Perch	0.1121	431	1,301	1,888	160	570	749	5,099
	(0.1753)	(857)	(1,354)	(1,152)	(141)	(528)	(552)	(4,584)
Angler hours		7,409	15,682	11,545	3,369	5,560	1,926	45,491
		(6,997)	(11,312)	(4,062)	(1,280)	(1,545)	(957)	(26,153)
Angler trips		1,852	3,634	2,860	664	1,532	461	11,003
		(1,599)	(2,538)	(1,110)	(266)	(460)	(258)	(6,230)
Angler Days		1,634	3,444	2,730	650	1,532	458	10,448
		(1,454)	(2,414)	(1,087)	(259)	(460)	(253)	(5,928)

Appendix 2, Table 3. Estimated harvest per hour, number harvested, and effort (angler hours, trips, and days) of sport fishing for Lake George to Green Point (includes Little Lake George, and the area from Bellview Marina to Stribling Point in Ontario, Canada), by all modes (non-charter) in 2017. Two standard errors in parentheses.

				Site 20	8			
					Month			
Species	Harvest per hour	May	June	July	August	September	October	Season
Atlantic Salmon	0.0010	10	0	11	11	0	0	32
	(0.0038)	(22)	(0)	(23)	(21)	(0)	(0)	(66)
Chinook Salmon	0.0009	0	0	0	8	21	0	29
	(0.0027)	(0)	(0)	(0)	(17)	(30)	(0)	(47)
Coho Salmon	0.0009	31	0	0	0	0	0	31
	(0.0025)	(43)	(0)	(0)	(0)	(0)	(0)	(43)
Lake Whitefish	0.0053	166	0	0	0	11	0	177
	(0.0167)	(267)	(0)	(0)	(0)	(21)	(0)	(288)
Northern Pike	0.0491	88	1,184	42	243	70	0	1,627
	(0.0457)	(102)	(226)	(88)	(210)	(165)	(0)	(791)
Pink Salmon	0.0002	0	0	0	8	0	0	8
	(0.0010)	(0)	(0)	(0)	(17)	(0)	(0)	(17)
Rainbow Trout	0.0040	36	0	65	8	0	23	132
	(0.0110)	(46)	(0)	(76)	(17)	(0)	(51)	(190)
Smallmouth Bass	0.0027	0	14	76	0	0	0	90
	(0.0080)	(0)	(30)	(109)	(0)	(0)	(0)	(139)
Walleye	0.0532	144	992	225	85	64	252	1,762
	(0.0963)	(207)	(683)	(254)	(108)	(129)	(286)	(1,667)
Yellow Perch	0.1504	511	499	610	913	230	2,218	4,981
	(0.3252)	(532)	(485)	(628)	(1,362)	(447)	(2,175)	(5,629)
Angler hours		2,321	9,441	6,898	5,789	5,567	3,106	33,122
		(1,752)	(5,699)	(2,755)	(2,813)	(2,415)	(1,870)	(17,306)
Angler trips		489	2,293	1,533	1,267	1,183	554	7,319
		(376)	(738)	(755)	(686)	(575)	(332)	(3,463)
Angler Days		458	2,293	1,511	1,228	1,183	554	7,227
		(356)	(738)	(752)	(670)	(575)	(332)	(3,423)

Appendix 2, Table 4. Estimated harvest per hour, number harvested, and effort (angler hours, trips, and days) of sport fishing for the area from the Neebish Island Ferry to the rapids in Sault Ste. Marie (includes Lake Nicolet, the St. Marys River below the rapids and the area from the rapids to Bellview Marina in Ontario, Canada), by all modes (non-charter) in 2017. Two standard errors in parentheses.

				Site 20	9			
				ſ	Month			
Species	Harvest per hour	May	June	July	August	September	October	Season
Atlantic Salmon	0.0402	18	124	481	255	94	7	979
	(0.0700)	(29)	(133)	(252)	(163)	(81)	(12)	(669)
Channel Catfish	0.0003	0	0	0	0	8	0	8
	(0.0016)	(0)	(0)	(0)	(0)	(15)	(0)	(15)
Chinook Salmon	0.0028	0	16	9	0	43	0	68
	(0.0099)	(0)	(32)	(18)	(0)	(45)	(0)	(95)
Coho Salmon	0.0005	13	0	0	0	0	0	13
	(0.0025)	(24)	(0)	(0)	(0)	(0)	(0)	(24)
Cisco	0.0025	0	0	62	0	0	0	62
	(0.0087)	(0)	(0)	(83)	(0)	(0)	(0)	(83)
Lake Trout	0.0004	0	0	0	0	9	0	9
	(0.0019)	(0)	(0)	(0)	(0)	(18)	(0)	(18)
Lake Whitefish	0.0634	291	457	708	25	62	0	1,543
	(0.1401)	(333)	(515)	(369)	(26)	(96)	(0)	(1,339)
Northern Pike	0.0086	0	50	100	7	52	0	209
	(0.0303)	(0)	(48)	(153)	(14)	(75)	(0)	(289)
Other	0.0013	0	16	0	0	16	0	32
	(0.0065)	(0)	(32)	(0)	(0)	(31)	(0)	(62)
Pink Salmon	0.0028	0	0	0	22	47	0	69
	(0.0083)	(0)	(0)	(0)	(27)	(52)	(0)	(79)
Rainbow Trout	0.0149	98	117	113	0	35	0	363
	(0.0433)	(144)	(141)	(93)	(0)	(35)	(0)	(414)
Smallmouth Bass	0.0011	0	16	10	0	0	0	26
	(0.0054)	(0)	(32)	(19)	(0)	(0)	(0)	(51)
Walleye	0.0263	0	185	27	286	141	0	639
	(0.0622)	(0)	(154)	(54)	(259)	(127)	(0)	(595)
Yellow Perch	0.1032	0	278	135	334	1,418	347	2,512
	(0.2841)	(0)	(245)	(273)	(361)	(1,242)	(595)	(2,716)
Angler hours		1,357	4,718	8,166	4,009	5,729	353	24,332
		(1,071)	(1,931)	(2,828)	(1,352)	(2,104)	(274)	(9,559)
Angler trips		373	1,006	1,854	976	1,276	77	5,562
		(309)	(418)	(729)	(392)	(493)	(60)	(2,400)
Angler Days		356	984	1,565	850	1,230	77	5,062
		(295)	(411)	(628)	(361)	(475)	(60)	(2,229)

				Site 210				
				M	onth			
Species	Harvest per hour	May	June	July	August	September	October	Season
Cisco	0.3254	0	0	32,205	0	0	0	32,205
	(0.3841)	(0)	(0)	(18,172)	(0)	(0)	(0)	(18,172)
Lake Whitefish	0.0430	0	0	4,254	0	0	0	4,254
	(0.0663)	(0)	(0)	(3,138)	(0)	(0)	(0)	(3,138)
Northern Pike	0.0140	372	214	537	44	190	27	1,384
	(0.0356)	(570)	(220)	(588)	(70)	(179)	(56)	(1,682)
Pumpkinseed	0.0022	0	161	0	59	0	0	220
	(0.0068)	(0)	(244)	(0)	(76)	(0)	(0)	(320)
Smallmouth Bass	0.0268	174	1,232	1,074	88	86	0	2,654
	(0.0562)	(294)	(1,257)	(885)	(101)	(121)	(0)	(2,658)
Walleye	0.0295	450	569	232	448	718	499	2,916
	(0.0570)	(531)	(507)	(320)	(349)	(539)	(450)	(2,697)
Yellow Perch	0.2636	0	142	58	636	15,637	9,614	26,087
	(0.3301)	(0)	(169)	(118)	(654)	(7,506)	(7,170)	(15,616)
Angler hours		6,754	17,346	43,507	8,756	16,342	6,261	98,966
-		(6,172)	(10,342)	(18,686)	(2,556)	(5,801)	(3,755)	(47,311)
Angler trips		1,520	3,978	10,682	1,473	4,040	1,395	23,088
·		(1,382)	(1,647)	(4,741)	(642)	(1,455)	(866)	(10,734)
Angler Days		1,409	3,805	9,727	1,378	3,791	1,350	21,460
		(1,278)	(1,593)	(4,358)	(587)	(1,392)	(845)	(10,054)

Appendix 2, Table 5. Estimated harvest per hour, number harvested, and effort (angler hours, trips, and days) of sport fishing for Potagannissing Bay (Michigan and Ontario), by allmodes (non-charter), in 2017. Two standard errors in parentheses.

Appendix 2, Table 6. Estimated harvest per hour, number harvested, and effort (angler hours, trips, and days) of sport fishing for the Rapids in Sault Ste. Marie, Ontario, by all modes (non-charter) in 2017. Two standard errors in parentheses.

				Site 40)3			
					Month			
Species	Harvest per hour	Мау	June	July	August	September	October	Season
Atlantic Salmon	0.0011	0	0	0	9	5	0	14
	(0.0040)	(0)	(0)	(0)	(12)	(9)	(0)	(21)
Brown Trout	0.0002	0	0	0	2	0	0	2
	(0.0009)	(0)	(0)	(0)	(5)	(0)	(0)	(5)
Chinook Salmon	0.0032	0	0	0	10	9	22	41
	(0.0125)	(0)	(0)	(0)	(19)	(12)	(36)	(67)
Coho Salmon	0.0016	19	0	0	0	1	0	20
	(0.0071)	(36)	(0)	(0)	(0)	(2)	(0)	(38)
Lake Whitefish	0.0029	0	0	0	0	0	37	37
	(0.0132)	(0)	(0)	(0)	(0)	(0)	(71)	(71)
Pink Salmon	0.0036	0	0	0	0	46	0	46
	(0.0116)	(0)	(0)	(0)	(0)	(62)	(0)	(62)
Rainbow Trout	0.0243	107	73	40	41	45	0 (0) 0 (0) 22 (36) 0 (0) 37 (71) 0	312
	(0.0498)	(77)	(52)	(41)	(50)	(39)	(7)	(266)
Walleye	0.0003	0	0	0	0	0	4	4
	(0.0012)	(0)	(0)	(0)	(0)	(0)	(6)	(6)
Angler hours		4,604	2,469	1,796	869	1,641	1,475	12,854
		(1,338)	(1,342)	(752)	(460)	(732)	(727)	(5,350)
Angler trips		1,164	700	573	302	544	434	3,717
		(363)	(390)	(258)	(164)	(251)	(226)	(1,653)
Angler Days		1,142	694	531	294	535	427	3,623
		(356)	(389)	(237)	(161)	(247)	(224)	(1,614)

Appendix 2, Table 7. Estimated harvest per hour, number harvested, and effort (angler hours, trips, and days) of sport fishing for the St. Joseph Channel, Ontario, Canada, by all modes (non-charter) in 2017. Two standard errors in parentheses.

				Site 4	05			
	Month							
Species	Harvest per hour	May	June	July	August	September	October	Season
Northern Pike	0.0095	295	0	0	0	0	r October 0 (0) 0 (0) 0 (0) 0 (0) 0 (0) 0 0 (0)	295
	(0.0332)	(414)	(0)	(0)	(0)	(0)	(0)	(414)
Smallmouth Bass	0.0076	0	0	0	0	235	0	235
	(0.0340)	(0)	(0)	(0)	(0)	(424)	(0)	(424)
Walleye	0.0524	0	481	1,143	0	0	0	1,624
	(0.1910)	(0)	(616)	(1,763)	(0)	(0)	(0)	(2,379)
Yellow Perch	0.0181	0	0	272	0	290	0	562
	(0.0893)	(0)	(0)	(344)	(0)	(769)	(0)	(1,112)
Angler hours		1,273	5,811	10,201	7,090	5,765	870	31,010
		(70)	(204)	(5,714)	(3,344)	(2,261)	(862)	(12,455)
Angler trips		294	1,715	2,289	1,746	1,443	194	7,681
		(0)	(0)	(1,307)	(1,124)	(589)	(189)	(3,209)
Angler Days		294	1,618	2,289	1,746	1,443	194	7,584
		(0)	(0)	(1,307)	(1,124)	(589)	(189)	(3,209)

			Site	e All Sites				
				Μ	lonth		1	
Species	Catch per hour	May	June	July	August	September	October	Seasor
Atlantic Salmon	0.0071	185	219	761	454	116	18	1,753
	(0.0021)	(38)	(30)	(82)	(57)	(26)	(12)	(244)
Brown Trout	0.0000	0	0	0	2	4	0	6
	(0.0001)	(0)	(0)	(0)	(3)	(4)	(0)	(7)
Channel Catfish	0.0031	91	353	201	13	89	23	770
	(0.0014)	(19)	(51)	(48)	(10)	(24)	(10)	(162)
Chinook Salmon	0.0013	0	16	28	21	94	165	324
	(0.0008)	(0)	(8)	(11)	(13)	(32)	(26)	(89)
Coho Salmon	0.0004	63	0	0	0	1	46	110
	(0.0004)	(27)	(0)	(0)	(0)	(2)	(14)	(43)
Freshwater Drum	0.0007	0	75	23	37	0	43	178
	(0.0005)	(0)	(17)	(10)	(16)	(0)	(19)	(62)
Cisco	0.1369	0	0	33,658	0	0	0	33,658
	(0.0034)	(0)	(0)	(404)	(0)	(0)	(0)	(404)
Lake Trout	0.0001	0	0	0	0	9	9	18
	(0.0001)	(0)	(0)	(0)	(0)	(6)	(6)	(12)
Largemouth Bass	0.0001	0	0	0	21	0	4	25
Largemodin Dass	(0.0002)	(0)	(0)	(0)	(14)	(0)	(4)	(18)
Lake Whitefish	0.0262	506	536	5,232	25	111	37	6,447
Lake Willensh	(0.0030)	(63)	(46)		(10)		(12)	(349)
Musky	0.0009	48	(40) 116	(192) 0	0	(27) 15	35	(349)
IVIUSKY				-	-			
Northern Pike	(0.0006)	(14)	(32)	(0)	(0)	(8)	(12)	(66)
	0.0963	3,395	7,832	6,088	2,809	2,794	754	23,672
Other	(0.0133)	(238)	(414)	(354)	(225)	(225)	(114)	(1,569)
Other	0.0110	175	874	1,102	185	343	27	2,706
Diale Calassa	(0.0029)	(37)	(110)	(87)	(41)	(51)	(10)	(338)
Pink Salmon	0.0047	0	0	0	30	512	624	1,166
D	(0.0011)	(0)	(0)	(0)	(15)	(60)	(50)	(125)
Pumpkinseed	0.0033	12	224	215	342	0	27	820
	(0.0011)	(7)	(41)	(29)	(40)	(0)	(10)	(128)
Rainbow Trout	0.0181	2,327	1,131	437	131	130	285	4,441
	(0.0035)	(127)	(100)	(71)	(35)	(32)	(50)	(415)
Rock Bass	0.0077	0	1,256	495	52	51	41	1,895
	(0.0019)	(0)	(121)	(65)	(14)	(14)	(13)	(227)
Smallmouth Bass	0.0691	1,048	8,037	5,889	498	1,414	104	16,990
	(0.0083)	(93)	(350)	(283)	(79)	(147)	(32)	(985)
Walleye	0.1583	10,563	16,121	5,063	3,712	2,220	1,218	38,897
	(0.0134)	(272)	(472)	(286)	(269)	(171)	(120)	(1,589)
White Bass	0.0001	0	0	0	31	0	0	31
	(0.0001)	(0)	(0)	(0)	(11)	(0)	(0)	(11)
Yellow Perch	0.4034	2,069	5,624	11,041	5,454	44,131	30,827	99,146
	(0.0202)	(146)	(297)	(419)	(286)	(664)	(579)	(2,391)

Appendix 3, Table 1. Estimated catch including legal and nonlegal release plus harvest for all sites combined on the St. Marys River in 2017. Two standard errors are in parentheses.

Appendix 3, Table 1. Cont.

Angler hours	23,718	55,467	82,113	29,882	40,604	13,991	245,775
	(17,400)	(30,830)	(34,796)	(11,805)	(14,857)	(8,445)	(118,134)
Angler trips	5,692	13,326	19,791	6,428	10,018	3,115	58,370
	(4,029)	(5,731)	(8,901)	(3,275)	(3,822)	(1,931)	(27,688)
Angler Days	5,293	12,838	18,353	6,146	9,714	3,060	55,404
	(3,740)	(5,546)	(8,368)	(3,163)	(3,738)	(1,902)	(26,456)

			S	ite 207-2	017			
				Ν	Ionth			
Species	Catch per hour	Мау	June	July	August	September	October	Season
Channel Catfish	0.0077	0	264	0	5	81	0	350
	(0.0021)	(0)	(32)	(0)	(4)	(18)	(0)	(55)
Freshwater Drum	0.0023	0	75	0	8	0	20	103
	(0.0012)	(0)	(17)	(0)	(6)	(0)	(9)	(32)
Cisco	0.0027	0	0	122	0	0	0	122
	(0.0008)	(0)	(0)	(22)	(0)	(0)	(0)	(22)
Largemouth Bass	0.0001	0	0	0	5	0	0	5
	(0.0002)	(0)	(0)	(0)	(4)	(0)	(0)	(4)
Musky	0.0042	48	95	0	0	15	35	193
	(0.0020)	(14)	(19)	(0)	(0)	(8)	(12)	(53)
Northern Pike	0.1493	1,046	2,377	1,756	401	841	371	6,792
	(0.0146)	(65)	(98)	(84)	(40)	(58)	(39)	(383)
Other	0.0046	0	152	43	13	0	0	208
	(0.0017)	(0)	(25)	(13)	(7)	(0)	(0)	(45)
Rock Bass	0.0155	0	623	43	0	0	41	707
	(0.0029)	(0)	(50)	(13)	(0)	(0)	(13)	(76)
Smallmouth Bass	0.0295	95	436	437	51	295	29	1,343
	(0.0064)	(19)	(42)	(46)	(14)	(34)	(11)	(167)
Walleye	0.5692	9,842	11,547	2,792	653	862	196	25,892
	(0.0251)	(198)	(215)	(106)	(51)	(59)	(28)	(657)
Yellow Perch	0.2256	526	3,171	3,518	470	1,012	1,565	10,262
	(0.0182)	(46)	(113)	(123)	(43)	(64)	(86)	(475)
Angler hours		7,409	15,682	11,545	3,369	5,560	1,926	45,491
		(6,997)	(11,312)	(4,062)	(1,280)	(1,545)	(957)	(26,153)
Angler trips		1,852	3,634	2,860	664	1,532	461	11,003
		(1,599)	(2,538)	(1,110)	(266)	(460)	(258)	(6,230)
Angler Days		1,634	3,444	2,730	650	1,532	458	10,448
		(1,454)	(2,414)	(1,087)	(259)	(460)	(253)	(5,928)

Appendix 3, Table 2. Estimated catch including legal and nonlegal release plus harvest for Site 207. Two standard errors are in parentheses.

			S	ite 208 -				
			1	1	Month			1
Species	Catch per hour	May	June	July	August	September	October	Season
Atlantic Salmon	0.0032	73	0	23	11	0	0	107
	(0.0019)	(17)	(0)	(10)	(7)	(0)	(0)	(33)
Channel Catfish	0.0055	91	0	67	0	0	23	181
	(0.0026)	(19)	(0)	(16)	(0)	(0)	(10)	(45)
Chinook Salmon	0.0009	0	0	0	8	21	0	29
	(0.0009)	(0)	(0)	(0)	(6)	(9)	(0)	(15)
Coho Salmon	0.0009	31	0	0	0	0	0	31
	(0.0006)	(11)	(0)	(0)	(0)	(0)	(0)	(11)
Freshwater Drum	0.0014	0	0	23	0	0	23	46
	(0.0011)	(0)	(0)	(10)	(0)	(0)	(10)	(19)
Largemouth Bass	0.0001	0	0	0	0	0	4	4
-	(0.0002)	(0)	(0)	(0)	(0)	(0)	(4)	(4)
Lake Whitefish	0.0053	166	0	0	0	11	0	177
	(0.0019)	(26)	(0)	(0)	(0)	(7)	(0)	(32)
Musky	0.0004	0	14	0	0	0	0	14
•	(0.0004)	(0)	(7)	(0)	(0)	(0)	(0)	(7)
Northern Pike	0.2095	461	3,396	1,589	942	449	101	6,938
	(0.0241)	(43)	(160)	(90)	(61)	(42)	(20)	(417)
Other	0.0071	71	143	0	11	11	0	236
	(0.0031)	(17)	(24)	(0)	(7)	(7)	(0)	(54)
Pink Salmon	0.0002	0	0	0	8	0	0	8
	(0.0003)	(0)	(0)	(0)	(6)	(0)	(0)	(6)
Rainbow Trout	0.0056	43	28	65	25	0	23	184
	(0.0034)	(13)	(11)	(16)	(10)	(0)	(10)	(59)
Smallmouth Bass	0.0441	46	360	931	78	43	4	1,462
	(0.0085)	(14)	(38)	(61)	(18)	(13)	(4)	(147)
Walleye	0.1251	189	2,519	735	349	85	268	4,145
-	(0.0161)	(27)	(100)	(61)	(37)	(18)	(33)	(278)
Yellow Perch	0.3309	1,377	885	1,239	1,876	925	4,659	10,961
	(0.0282)	(74)	(59)	(70)	(87)	(61)	(137)	(488)
Angler hours		2,321	9,441	6,898	5,789	5,567	3,106	33,122
		(1,752)	(5,699)	(2,755)	(2,813)	(2,415)	(1,870)	(17,306)
Angler trips		489	2,293	1,533	1,267	1,183	554	7,319
		(376)	(738)	(755)	(686)	(575)	(332)	(3,463)
Angler Days		458	2,293	1,511	1,228	1,183	554	7,227
		(356)	(738)	(752)	(670)	(575)	(332)	(3,423)

Appendix 3, Table 3. Estimated catch including legal and nonlegal release plus harvest for Site 208. Two standard errors are in parentheses.

			S	ite 209 -	2017			
					Month			
Species	Catch per hour	May	June	July	August	September	October	Season
Atlantic Salmon	0.0648	112	219	708	419	111	7	1,576
	(0.0187)	(21)	(30)	(61)	(41)	(21)	(5)	(179)
Channel Catfish	0.0007	0	0	0	8	8	0	16
	(0.0012)	(0)	(0)	(0)	(6)	(6)	(0)	(11)
Chinook Salmon	0.0042	0	16	28	0	59	0	103
	(0.0036)	(0)	(8)	(11)	(0)	(15)	(0)	(34)
Coho Salmon	0.0005	13	0	0	0	0	0	13
	(0.0008)	(7)	(0)	(0)	(0)	(0)	(0)	(7)
Cisco	0.0025	0	0	62	0	0	0	62
	(0.0016)	(0)	(0)	(16)	(0)	(0)	(0)	(16)
Lake Trout	0.0004	0	0	0	0	9	0	9
	(0.0006)	(0)	(0)	(0)	(0)	(6)	(0)	(6)
Lake Whitefish	0.0779	340	536	895	25	100	0	1,896
	(0.0181)	(37)	(46)	(60)	(10)	(20)	(0)	(173)
Musky	0.0003	0	7	0	0	0	0	7
	(0.0006)	(0)	(5)	(0)	(0)	(0)	(0)	(5)
Northern Pike	0.0434	27	521	191	70	195	53	1,057
	(0.0161)	(10)	(46)	(39)	(17)	(28)	(15)	(154)
Other	0.0020	0	16	9	0	23	0	48
	(0.0025)	(0)	(8)	(6)	(0)	(10)	(0)	(24)
Pink Salmon	0.0028	0	0	0	22	47	0	69
	(0.0028)	(0)	(0)	(0)	(9)	(17)	(0)	(26)
Rainbow Trout	0.0238	107	231	167	7	43	24	579
	(0.0110)	(21)	(30)	(26)	(5)	(13)	(10)	(105)
Smallmouth Bass	0.0159	0	336	10	26	16	0	388
	(0.0064)	(0)	(37)	(6)	(10)	(8)	(0)	(61)
Walleye	0.0502	0	532	46	443	201	0	1,222
	(0.0142)	(0)	(46)	(14)	(48)	(28)	(0)	(136)
Yellow Perch	0.2065	0	574	480	1,076	2,378	517	5,025
	(0.0314)	(0)	(48)	(44)	(66)	(98)	(45)	(300)
Angler hours		1,357	4,718	8,166	4,009	5,729	353	24,332
		(1,071)	(1,931)	(2,828)	(1,352)	(2,104)	(274)	(9,559)
Angler trips		373	1,006	1,854	976	1,276	77	5,562
		(309)	(418)	(729)	(392)	(493)	(60)	(2,400)
Angler Days		356	984	1,565	850	1,230	77	5,062
		(295)	(411)	(628)	(361)	(475)	(60)	(2,229)

Appendix 3, Table 4. Estimated catch including legal and nonlegal release plus harvest for Site 209. Two standard errors are in parentheses.

			S	ite 210 - 2	017			
				Μ	onth			
Species	Catch per hour	Мау	June	July	August	September	October	Season
Channel Catfish	0.0004	0	0	41	0	0	0	41
	(0.0003)	(0)	(0)	(13)	(0)	(0)	(0)	(13)
Freshwater Drum	0.0003	0	0	0	29	0	0	29
	(0.0002)	(0)	(0)	(0)	(11)	(0)	(0)	(11)
Cisco	0.3382	0	0	33,474	0	0	0	33,474
	(0.0077)	(0)	(0)	(366)	(0)	(0)	(0)	(366)
Largemouth Bass	0.0002	0	0	0	16	0	0	16
	(0.0002)	(0)	(0)	(0)	(10)	(0)	(0)	(10)
Lake Whitefish	0.0438	0	0	4,337	0	0	0	4,337
	(0.0028)	(0)	(0)	(132)	(0)	(0)	(0)	(132)
Northern Pike	0.0559	1,271	851	1,635	523	1,074	180	5,534
	(0.0074)	(71)	(58)	(81)	(48)	(66)	(27)	(351)
Other	0.0222	104	552	1,047	161	309	27	2,200
	(0.0043)	(20)	(47)	(65)	(27)	(35)	(10)	(205)
Pumpkinseed	0.0076	12	161	215	342	0	27	757
	(0.0024)	(7)	(25)	(29)	(40)	(0)	(10)	(112)
Rock Bass	0.0120	0	633	452	52	51	0	1,188
	(0.0032)	(0)	(71)	(52)	(14)	(14)	(0)	(152)
Smallmouth Bass	0.1010	907	4,166	4,069	343	438	71	9,994
	(0.0087)	(60)	(129)	(128)	(37)	(42)	(17)	(413)
Walleye	0.0510	532	836	347	1,506	1,072	750	5,043
	(0.0072)	(46)	(58)	(37)	(78)	(65)	(55)	(339)
White Bass	0.0003	0	0	0	31	0	0	31
	(0.0002)	(0)	(0)	(0)	(11)	(0)	(0)	(11)
Yellow Perch	0.7278	166	925	5,532	2,032	39,291	24,086	72,032
	(0.0218)	(26)	(61)	(149)	(90)	(396)	(310)	(1,032)
Angler hours		6,754	17,346	43,507	8,756	16,342	6,261	98,966
		(6,172)	(10,342)	(18,686)	(2,556)	(5,801)	(3,755)	(47,311)
Angler trips		1,520	3,978	10,682	1,473	4,040	1,395	23,088
		(1,382)	(1,647)	(4,741)	(642)	(1,455)	(866)	(10,734)
Angler Days		1,409	3,805	9,727	1,378	3,791	1,350	21,460
		(1,278)	(1,593)	(4,358)	(587)	(1,392)	(845)	(10,054)

Appendix 3, Table 5. Estimated catch including legal and nonlegal release plus harvest for Site 210. Two standard errors are in parentheses.

			S	ite 403-	2017						
	Month										
Species	Catch per hour	May	June	July	August	September	October	Season			
Atlantic Salmon	0.0054	0	0	30	24	5	11	70			
	(0.0060)	(0)	(0)	(11)	(10)	(4)	(7)	(32)			
Brown Trout	0.0005	0	0	0	2	4	0	6			
	(0.0013)	(0)	(0)	(0)	(3)	(4)	(0)	(7)			
Channel Catfish	0.0142	0	89	93	0	0	0	182			
	(0.0071)	(0)	(19)	(19)	(0)	(0)	(0)	(38)			
Chinook Salmon	0.0149	0	0	0	13	14	165	192			
	(0.0075)	(0)	(0)	(0)	(7)	(7)	(26)	(40)			
Coho Salmon	0.0051	19	0	0	0	1	46	66			
	(0.0045)	(9)	(0)	(0)	(0)	(2)	(14)	(24)			
Lake Trout	0.0007	0	0	0	0	0	9	9			
	(0.0011)	(0)	(0)	(0)	(0)	(0)	(6)	(6)			
Lake Whitefish	0.0029	0	0	0	0	0	37	37			
	(0.0023)	(0)	(0)	(0)	(0)	(0)	(12)	(12)			
Other	0.0011	0	11	3	0	0	0	14			
	(0.0019)	(0)	(7)	(3)	(0)	(0)	(0)	(10)			
Pink Salmon	0.0847	0	0	0	0	465	624	1,089			
	(0.0174)	(0)	(0)	(0)	(0)	(43)	(50)	(93)			
Pumpkinseed	0.0049	0	63	0	0	0	0	63			
	(0.0030)	(0)	(16)	(0)	(0)	(0)	(0)	(16)			
Rainbow Trout	0.2861	2,177	872	205	99	87	238	3,678			
	(0.0468)	(93)	(59)	(29)	(20)	(19)	(31)	(250)			
Walleye	0.0003	0	0	0	0	0	4	4			
	(0.0007)	(0)	(0)	(0)	(0)	(0)	(4)	(4)			
Angler hours		4,604	2,469	1,796	869	1,641	1,475	12,854			
		(1,338)	(1,342)	(752)	(460)	(732)	(727)	(5,350)			
Angler trips		1,164	700	573	302	544	434	3,717			
		(363)	(390)	(258)	(164)	(251)	(226)	(1,653)			
Angler Days		1,142	694	531	294	535	427	3,623			
		(356)	(389)	(237)	(161)	(247)	(224)	(1,614)			

Appendix 3, Table 6. Estimated catch including legal and nonlegal release plus harvest for Site 403. Two standard errors are in parentheses.

				Site 405	- 2017						
Species	Month										
	Catch per hour	May	June	July	August	September	October	Season			
Northern Pike	0.1081	590	687	917	873	235	49	3,351			
	(0.0213)	(49)	(52)	(61)	(59)	(31)	(14)	(265)			
Smallmouth Bass	0.1226	0	2,739	442	0	622	0	3,803			
	(0.0158)	(0)	(105)	(42)	(0)	(50)	(0)	(197)			
Walleye	0.0836	0	687	1,143	761	0	0	2,591			
	(0.0141)	(0)	(52)	(68)	(55)	(0)	(0)	(175)			
Yellow Perch	0.0279	0	69	272	0	525	0	866			
	(0.0077)	(0)	(17)	(33)	(0)	(46)	(0)	(95)			
Angler hours		1,273	5,811	10,201	7,090	5,765	870	31,010			
		(70)	(204)	(5,714)	(3,344)	(2,261)	(862)	(12,455)			
Angler trips		294	1,715	2,289	1,746	1,443	194	7,681			
		(0)	(0)	(1,307)	(1,124)	(589)	(189)	(3,209)			
Angler Days		294	1,618	2,289	1,746	1,443	194	7,584			
		(0)	(0)	(1,307)	(1,124)	(589)	(189)	(3,209)			

Appendix 3, Table 7. Estimated catch including legal and nonlegal release plus harvest for Site 405. Two standard errors are in parentheses.

Appendix 4. Summary of biological data collected from the St. Marys River during the open water sport fishery for the years 1999-2000, 2005-2009 and 2017, by capture sites. N = sample size and appears in parentheses if different than reported.

Species	Year	Capture sites	N	Mean Age	Mean Length (cm)	Mean Wt (g)
Atlantic Salmon	1999	209, 210	15	3.2 (14)	73.0	4 810
	2001	209	13	2.5 (13)	65.7	3 148
	2006	209	11	3.5 (11)	65.3	3 336
	2007	210	1	3.0(1)	59.7	2 041
	2008	208, 209	109	2.6 (95)	65.8	3 247
	2009	403	6	2.8 (6)	67.1	2 895
	2017	208, 209, 403	117	2.2 (41)	56.6	2174 (116)
Brown Trout	2017	403	1	3	46.0	1202
Channel Catfish	2017	207	2		49.9	1429
Chinook Salmon	1999	208, 209	214	3.0 (205)	84.0	6 698
	2000	208	14	3.1	86.0	7 355
	2001	208, 209, 210	14	2.9	81.0	5 621
	2006	208, 209, 210	56	3.5 (41)	77.0	4 248
	2007	208, 210, 405	62	2.7	75.0	4 090
	2008	208, 209, 403	47	2.7 (36)	75.4	4 611
	2009	207, 403	7	2.9	82.4	5 242
	2017	208, 403	6	2.8 (5)	78.1	5934
Coho Salmon	1999	209	18	2.7 (16)	60.2	2 694
	2006	208, 209	5	2.0 (4)	52.1	1 760
	2007	208, 405	6	2.7	69.6	3 145
	2008	208, 209, 403	36	2	57.4	1 971
	2009	403	18	2.1	64.9	2 389
	2017	403	1	2	43.4	1202
Cisco	1999	207, 209, 210	138	4.2	33.8	540 (110)
	2000	207, 209, 210	88	3.8 (85)	31.6	469
	2001	207, 209, 210	58	4	33.8	
	2005	207, 210	53	3	32.4	358
	2006	210	70	5.3	36.8	526
	2007	207, 210	65	4.1	34.9	447
	2008	209	23	4.2	36.4	531
	2009	207, 210	95	4.9 (93)	38.0	606
	2017	209, 210	92	, , , , , , , , , , , , , , , , , , , ,	38.5	605 (91)
Freshwater Drum	2017	207	1		50.8	2540
Lake Whitefish	1999	209, 404	157	4.6 (154)	41.5	614 (156)
	2000	210	2	5	49.5	1 270
	2006	209	7	3.3	41.2	719
	2007	207, 210	12	4.9	42.9	794
	2008	209	60	3.7	38.3	584
	2009	210	6		41.6	726
	2009	208, 405	11	3.1	36.3	920
	2008	210	6	5.7	41.6	726
	2007	208, 210, 403	4	4 (3)	46.4	1008
Largemouth Bass	2017	208, 210, 405	11	3.2	36.3	920
Largemouth Dass	2008	210	1	5.2	38.1	771

Appendix 4, Cont.

			ŊŢ	Mean	Mean Length	Mean Wt
Species	Year	Capture sites	N	Age	(cm)	(g)
Muskellunge	2000	207	8		108.4	8 541
	2007	207	1		94.5	
	2008	209	1	10	108.0	8 618
	2009	207	3	4	76.7	33 817
		207, 208, 209,				
Northern Pike	1999	210	88	4.6 (86)	66.8 (87)	1 852 (87)
		207, 208, 209,				
	2000	210	42	4.3	66.8	1 702
	2001	207, 208, 210	22	5.6	69.6	2 208
	2005	207, 210	15	5.1	73.4	2 734
	2006	208, 207, 209	83	3.8	65.1	1 855
		207, 208, 210,				
	2007	405	70	4.4	67.3	2 223
	2008	208, 209, 405	146	4.0	69.8	2 224
	2009	207, 210, 405	45	5.6 (44)	66.5	1 878
	2017	207, 208, 209, 210	74	4.6	67.6	1876
Pink Salmon	1999	208, 209	82	2.0 (56)	52.1	1 398
	2000	209	1	1	49.3	907
	2008	209, 403	31	1 (14)	44.3	661
	2009	403	2		47.2	1 089
	2017	208, 209, 403	31	2 (7)	50.5	1283
Rainbow Trout	1999	209	29	2.5 (28)	50.3	1 595 (28)
	2000	208, 209	2	2	74.2	2 381
	2001	208, 209	2	3	61.0	2 041
	2006	209	11	2.5	44.9	1 064
	2008	208, 209, 403	25	3.5 (16)	54.6	1 970
	2009	403	69	4.8 (63)	63.7	2 757
	2017	208, 209, 403	32	2.5	48.5	1549
Smallmouth Bass	1999	208	10	6.3	36.5	809
	2000	207, 210	22	6.5	39.4	1 000
	2001	207, 210	12	5.6	21.9	1 104
	2005	207, 210	52	4.7	39.2	1 010
	2006	207, 208, 209	44	5.7	41.3	1 334
	2007	207, 210, 405	57	4.8 (56)	36.5	909
	2008	208, 209	52	5.3 (51)	40.1	1 225
	2009	207, 210, 405	63	5.7	40.0	1 172
	2017	207, 208, 209, 210	57	4 (2)	39.6	1012 (32)
		207, 208, 209,				

Species	Year	Capture sites	N	Mean Age	Mean Length (cm)	Mean Wt (g)
Walleye	1999	210	205	5.4 (203)	47.1	1 042
	2000	207, 209, 210	78	5.7	48.5	1 135
	2001	207, 209, 210	211	4.1	47.7	1 046
	2005	207, 210	189	5.3	45.9	987
	2006	207, 208, 209	148	4	44.5	1 000
		207, 210, 208.				
	2007	405	259	4.7 (257)	44.7	1 026
	2008	208, 209	183	4.8	46.7	1 095
	2009	207, 210, 405	173	5.7 (172)	46.1	1 083
		207, 208, 209, 210,				
	2017	403	157	5.3 (21)	46.3	1019 (116)
Yellow Perch	1999	207, 209, 210	258	5.5 (255)	21.9 (257)	151 (250)
	2000	207 210	127	3.4	24.2	321
		207, 208, 209,				
	2001	210	100	3.8	23.0	180
	2005	207, 210	150	3.7 (142)	21.2	125
		207, 208, 209,				
	2006	210	160	3.1 (159)	22.2	223
	2007	207, 210, 405	199	3.2 (195)	22.7	198
	2008	208, 209	174	3.3	21.5	135
	2009	207, 210, 405	190	3.5 (189)	21.3	128
	2017	207, 208, 209, 210	170	5.3 (3)	22.9	165 (155)

Appendix 2b: Population dynamics of the St. Marys River fish community 1975 - 2017 (O'Connor et al. 2019)

Population Dynamics of the St. Marys River Fish Community 1975-2017

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Abstract- The St. Marys River fish community was jointly assessed by the member agencies of the St. Marys River Fisheries Task Group under the Great Lakes Fishery Commission in 2017, the 9th such survey since 1975. A gillnet based survey, 44 nets sets each survey year resulted in indices of abundance and population status. Abundance of two cool water species of importance, Walleye and Yellow Perch, were not significantly lower than the 2013 estimates, and have remained stable in the River since 2006. Smallmouth Bass abundance has varied since 2002, with significant peaks in 2006 and 2013. Cisco have maintained stable but lower overall abundance within the River in 2017; however, Northern Pike have continued to increase since 2002, with the highest River abundance reached in 2017. Growth rates, as indicated by length at age at capture, were generally near or below regional averages and may reflect the northern latitude of the St. Marys River. Total annual mortality rates were 59% for Yellow Perch, 49% for Northern Pike, 48% for Smallmouth Bass, 62% for Cisco, and 61% Walleye but were generally deemed within acceptable ranges for these species. Diets varied by species and reflected both piscine prey and invertebrates, especially crayfish. Ruffe were documented for the first time in the Fish Community Survey in 2017, and were reported by anglers in the upper river during the same year. Round gobies continued to be observed in the diets of some predators indicating that they continue to persist in the river fish community. Recommended are timing future surveys with full river-wide creel surveys for maximum information and to increase the frequency of both.

Introduction

The St. Marys River supports a highly diverse fish community reflecting its varied habitat types. Most of the St. Marys constitutes cool water habitat typical of the nearshore Great Lakes environs, but cold water from Lake Superior also results in cold water habitat beneficial for salmon and trout species. The fish community supports recreational, commercial and subsistence fisheries. Recreational fishing effort can be substantial, amounting to as much as one third the total of the Michigan waters of Lake Huron and has been valued at \$8.5 million USD (Godby et al. 2019).

Despite the varied habitat types and high quality water source, the St. Marys River has been the subject of considerable anthropogenic alteration and degradation. The river is channelized throughout much of its reach to accommodate international shipping traffic (Edsall and Gannon 1993). The River was designated as an Area of Concern in the 1987 Great Lakes Water Quality Agreement (GLWQA, 1987). Fishery management challenges also result from the complications of shared resources across multiple fisheries and jurisdictions (Fielder 2002). The St. Marys River constitutes the international boundary water between Michigan and Ontario and includes Native American and Canadian First Nations as well. Fishery management is coordinated through the Great Lakes Fishery Commission's Lake Huron Committee and assessment through its St. Marys River Fisheries Task Group (Fielder 2002). Formed in 1997, representatives of the various management authorities and federal agencies as well as area universities work together for periodic assessment of the fish community. A river fishery assessment plan was developed in 2002 that included the need for and outlined a protocol for a fish community assessment for the St. Marys River (Gebhardt et al. 2002).

The objectives of this survey are to assess and provide information on the abundance, growth, mortality and size structure of important fish populations found in the St. Marys River; to make comparisons to previous surveys; and to comment on the overall current status of certain notable species.

Study Site

The St. Marys River is a connecting channel between Lakes Superior and Huron (Figure 1). The river flows southeasterly about 112 km and empties into Lake Huron at De Tour, Michigan but also drains into Ontario's North Channel through the St. Joseph Channel and Potagannissing Bay. Four large islands divide the river flow into these various channels and the river is bordered on the northeast by Ontario and Michigan on the other side. The river includes a variety of lacustrine reaches; specifically Lake Nicolet, Lake George, Lake Munuscong, and Raber Bay. For practical purposes, and for this study, Potagannissing Bay is also considered part of the St. Marys River. The rapids at Sault Ste. Marie is perhaps one of the most well-known features of this river, although today 93% of the river flow is diverted for hydroelectric generation (Edsall and Gannon 1993). The St. Marys River aquatic habitat includes an expanse of coastal wetlands that provide spawning and nursery habitat for fish (Albert 2003). Duffy et al. (1987) describes in detail the ecological and physical characteristics of the St. Marys River.

Methods

This study followed the fish community assessment procedure recommended by Gebhardt et al. (2002) which in turn was based on the methods used by past surveys (Schorfhaar 1975; Miller 1981; Grimm 1989; Fielder and Waybrant 1998) so as to allow comparability. Multifilament nylon gillnets were used to collect fish in this study. In this survey and since 2002 the nets measured 1.8 m deep by 304.8 m long and were comprised of ten different mesh sizes, each of which is a 30.5 m long

panel. Mesh sizes were; 38.1mm, 50.8 mm, 63.5 mm, 76.2 mm, 88.9 mm, 101.6 mm, 114.3 mm, 127.0 mm, 139.7 mm, and 152.4 mm stretch measure. The survey nets in 1975, 1979, 1987, and 1995 only utilized four mesh sizes; 50.8 mm, 63.5 mm, 76.2 mm and 114.3 mm stretch measure mesh and panels were 30.5 m in length. Nets were fished overnight on the bottom for all surveys.

Field work was jointly conducted by the member agencies of the St. Marys River Fisheries Task Group. They were the Sault Tribe Natural Resources Department (STNRD), Michigan Department of Natural Resources (MDNR), Ontario Ministry of Natural Resources and Forestry (OMNRF), and the United States Fish and Wildlife Service (USFWS). Net set locations were divided throughout the St. Marys River (Figure 1). Data were organized into seven distinct areas based on habitat and geographic regions within the river; Upper River, Lake Nicolet, Lake George, Lake Munuscong, St. Joseph Channel, Raber Bay and Potagannissing Bay (Figure 1, Table 1) for the purpose of some analyses. Many analyses include results from previous surveys for comparison purposes (Schorfhaar 1975; Miller 1981; Grimm 1989; Fielder and Waybrant 1998; Fielder et al. 2004; Fielder et al. 2007; Schaeffer et al. 2011, Schaeffer et al 2016).

The catch from each lift was identified, weighed (round weight) and measured for total length. Five species of special interest, Walleye, Yellow Perch, Smallmouth Bass, Northern Pike, Cisco, had scales or dorsal spines were collected for aging (see Appendix 1 for a complete listing of all the common and scientific names of fishes mentioned in this report). These same species were internally inspected for sex, maturity (according to the methods of Fielder and Waybrant (1998)), and stomach contents. Stomach contents were identified when possible and enumerated. All Walleye stocked into the St. Marys River were marked with oxytetracycline (OTC) prior to release. All collected Walleye otoliths were examined to determine if collected individuals were stocked or wild fish.

Catch-per-unit-of-effort (CPUE) was calculated in two ways: full net; the total number of each species per net lift per 304.8 m of net across all mesh sizes and the second; traditional net; the total number of each species per net lift from four meshes: 50.8 mm, 63.5 mm, 76.2 mm, and 114.3 mm in 122 m net length, which was then extrapolated to 304.8 m. This second method of expressing CPUE allowed a more direct comparison with the pre-2002 surveys ("traditional nets"). The CPUE values of the two different methods were compared for each species to determine if there were differences in CPUE based on the "traditional" and "full" meshes fished. Total species composition was also compared between the two different "nets": full net, comprised of ten individually sized mesh panels vs. the traditional, four individual mesh panels extrapolated to the full net length of 304.8 m. .

Total annual mortality was derived using the Robson-Chapman method (Van Den Avyle and Hayward 1999) on certain species of interest. Age information was also organized by CPUE so as to compare year class strength. Growth rate was expressed as mean length-at-age-at-capture and compared to Michigan averages according to Schneider et al. (2000) and to Lake Huron averages for those species. The Lake Huron data were means of total length from the North Channel of Lake Huron for collections made in similar times of the year (OMNR unpublished data). Survey growth rate averages were also compared to data from past surveys. Condition was expressed as relative weight (Wr; Ney 1999). Growth parameters were further explored via length / weight relationships and Von Bertalanffy growth equations (Van Den Avyle and Hayward 1999) for some species.

Testing for differences of means between two independent samples used the t-test where possible and the Mann-Whitney U (M-WU) test when the assumption of normality could not be met. We assessed the differences in CPUE within and between survey years using non-parametric Kruskal-Wallis (K-W) tests, with Dunn's post-hoc analysis. Some data and means from past surveys were recalculated for reporting and comparison purposes in this report and may differ slightly from those reported by past authors. Length / weight analysis used log transformed data for linear regressions. All statistical tests were performed at the significance level of P \leq 0.05 and followed the methods of Sokal and Rohlf (1981). Analysis was performed using SPSS computer software (SPSS 2001) and R 3.5.0 (R Core Team 2018,), the *ridgeline plots* package (*v.0.5.1*; Wilkie, 2018).

Results

In the 2017 survey, a total of 44 nets were set throughout the river over a 4 week period beginning the end of July through late August (Figure 1, Table 1). A total of 3226 fish representing 30 different species were collected. CPUE was calculated in two ways: traditional and full nets, as described in the Methods section above. For the traditional nets, the catches from four meshes (50.8 mm, 63.5 mm, 76.2 mm and 114.3 mm) were extrapolated to fill the 304.8 m, to match the panels of the historical nets and the length of the full nets (Table 2). For the full nets, CPUE was calculated for each species for the full ten mesh panels (Table 3). Mean CPUE for 2017 was compared (Mann-Whitney) between the two nets types for five species: Northern Pike, Cisco, Walleye, Yellow Perch, and Smallmouth Bass. In 2017, mean CPUE was not significantly different between the full and traditional nets for any of the five species. Mean CPUE between the two net-types for these five species was also compared from 2002 (first year of the full 10 individual mesh size nets) through 2017. Only in 2006, where mean CPUE for Northern Pike was significantly higher for the extrapolated traditional net (M-WU; P=0.013), was there a difference between the two net groups for any of the five species.

While there was no difference between the net types for the five individual species, there was a significant difference in the number of species captured by the two net sets. In 2017, the number of species collected in the full nets was significantly higher (M-WU; P=0.01) than in the traditional nets. This was consistent for 2002, 2006, 2009, and 2013 (M-WU; P<0.001; all years). The full mesh nets also had a higher cumulative species catch and collected five additional species not caught in the traditional nets: Coho Salmon, Creek Chub, Longnose Dace, Muskellunge, and White Crappie (see Appendix 2 for full summary of cumulative net catch by net type). Based on the mean CPUE and catch comparisons between the two nets, main results for individual species and main groups were based on the full nets, unless otherwise indicated.

Individual Species CPUE

Yellow Perch:

Yellow Perch abundance continued to demonstrate an overall stability on a river-wide basis but was down relative to the 2013 survey (Table 3), however, this was not significantly lower (K-W test, P=0.371). When examined by river reach, significant differences in abundance between the reaches were noted (K-W Tests, P=0.005). Yellow Perch abundance declined in 3 reaches and increased in the other 4 (Table 4). There were declines in abundance In the Upper River, Lake Munuscong, and Potagannissing Bay. Yellow Perch abundance in the Upper River was the lowest recorded since the survey began in 1975 with an average CPUE of 6.0 in 2017. Lake Munuscong declined from a mean CPUE of 26.0 in 2013 to 10.5 in 2017, while Potagannissing Bay declined from a historical high mean CPUE of 88.5 in 2013 to 56.2 (Table 4). Lake Nicolet and Raber Bay Yellow Perch abundances were consistent in 2017 with only small increases relative to the 2013 survey. There were larger increases in Lake George where mean CPUE went from 38.3 in 2013 to 50.0 in 2017 and the St. Joseph Channel where mean CPUE went from 6.9 in 2013 to 21.5 in 2017 (Table 4).

Northern Pike:

Northern Pike CPUE has continued to increase since 2002, which was the lowest level measured in the survey series (Table 3). Mean CPUE in 2017 (4.09) was significantly greater (K-W test, P=0.04) than in 2002, 2006 and 2009 (Dunn's Test, P = 0.005, P=0.01, P=0.01, respectively), however, not significantly greater than the 2013 survey mean CPUE (x=2.66) (Dunn's Test, P=0.100). Northern Pike CPUE increased in 2017 in four of the six river reaches (Table 4). Catch was significantly lower in the Upper River (K-W test; P=0.005) compared to lakes Nicolet, George and Munuscong (Dunn's Tests; P=0.013, P=0.028, P<0.001, respectively) and CPUE was significantly lower in Potagannissing Bay compared to Lake Munuscong (M-WU Test, P<0.001). Catches remained similar to the 2013 CPUE in Lake George and Lake Munuscong, with increases in Potagannissing Bay, Raber Bay, and Lake Nicolet in 2017.

Walleye:

Mean CPUE of Walleye (3.41) was lower in 2017 compared to the peak CPUE in 2013 (7.58); however, it was not significantly different (M-WU Test, P=0.483). Walleye CPUE has remained stable since 2006, with an increase from the 2002 survey (2.55) (Table 3). The CPUE in 2002 was significantly lower (K-W test, P=0.015), compared to each of the 2006 – 2017 fishing surveys. Within the fishing locations in the river, mean CPUE was the lowest in Lake Nicolet, however, there was no significant difference between the river locations in 2017 (K-W Test; P=0.124)(Table 4).

Smallmouth Bass:

Smallmouth Bass mean CPUE dropped to 2.84 in 2017, just under half the 6.63 mean CPUE reached in 2013 which was the highest in the time series (Table 3). Smallmouth Bass mean CPUE was variable over the last five surveys (Table 3), with significant differences among the years (K-W test, P=0.001). The peak catches in 2006 and 2013 were significantly greater than catches in 2002, 2009, and 2017. All reaches showed declines in Smallmouth Bass abundance with the exception of the St. Joseph Channel which increased from a mean CPUE of 8.1 in 2013 to 9.0 in 2017 (Table 45). The St. Joseph Channel also had the highest Smallmouth Bass abundance of any reach in 2017. This was a change from 2013 when Lake George had the highest abundance (16.2), which was the highest recorded abundance for any reach in the time series. Overall, the mean CPUE was significantly different between the reaches (K-W Test; P=0.025), with the catches in Lake George, St. Joseph Channel and Raber Bay having the largest abundance (Table 4).

Cisco:

The CPUE of Cisco in the full mesh nets was the second lowest of the time series (mean CPUE = 1.02), however, it was not significantly different over time for 2002 -2017 (K-W test, P=0.831) (Table 3). No Cisco were found in three reaches (Nicolet, Munuscong, and Raber) and Cisco occurred in low abundance in the remaining reaches (Table 4). For the reaches with Cisco, there were no significant differences in the catch CPUE. Early in the time series, the lower most reaches of the St. Marys (Raber and Potagannissing) produced large catches of Cisco, but they have largely been in low abundance since at least 2009.

Other Species:

Mean CPUE was calculated for all of the species collected each year. White Sucker mean CPUE remains high and stable, along with Rock Bass and Brown Bullhead, with no difference among years for the surveys from 2002 through 2017 (K-W tests; P=0.382, P= 0.756, P=0.526) (Table 3). Other species including Lake Whitefish, Burbot, and Menominee have remained stable through the survey years, though at lower CPUEs (K-W tests; P=0.645, P= 0.870, P=0.880)(Table 3).

Aquatic Invasive Species (AIS)

Several AIS were captured during the 2017 survey: Alewife, Rainbow Smelt, White Perch, Sea Lamprey, and new in 2017, Eurasian Ruffe (Table 3). Alewife CPUE has remained stable in all years following their peak CPUE in 2002 (K-W test, P<0.001). Rainbow Smelt and White Perch have remained low and stable in all years (K-W tests, P=0.653, P=0.182), while Ruffe CPUE (0.23) was the first for this survey in the St. Marys River (Table 3).

Species of Concern

A total of 26 Lake Sturgeon were captured in 2017, the highest of any year in the survey series. Mean CPUE was significantly higher in 2017 (0.59) (K-W tests, P<0.001) than any of the other survey years (Table 3). Lake Sturgeon were collected in four of the six netting locations below the Compensating Works: Lake George, Lake Munuscong, Raber and Potagannissing bays, with the majority captured in Lake Munuscong (N=11). Sturgeon ranged in size from 330 mm to 865 mm. The increase in Lake Sturgeon catch in the nets is the highest net CPUE since 1975 (Table 3).

Age, Maturity and Condition

Scales and dorsal spines were collected for aging from Yellow Perch, Walleye, Smallmouth Bass, and Cisco and cleithra for Northern Pike. These fish were also examined for sex, maturity and stomach contents. In addition to aging, walleye otoliths were also collected. These were examined for oxytetracycline marks to determine whether the individual fish was of native or stocked origin.

Yellow Perch:

The 2016 age-1 Yellow Perch year class was not well represented throughout the river, but they may not have fully recruited to the gear yet as has been the case in past surveys. Age-2 fish made up 35% of the Yellow Perch catch river-wide (Table 5). Relative to the MI average the Yellow Perch growth index was +9 in 2017 an increase over the 2013 value of +3 (Table 5).

The total annual mortality rate for Yellow Perch on a river-wide base has been declining since the 2006 survey where it peaked at 0.70 and is down to 0.41 in 2017 (Table 6). Total annual mortality for Yellow Perch declined markedly from 2013 to 2017 in Lake Nicolet, Lake George, and Raber Bay (Table 6). The highest Yellow Perch total annual mortality rate among those reaches that it was calculated for was Lake Munuscong at 0.76 an increase from the 2013 survey mortality rate of 0.63. All size classes in the 2017 maturity schedule for Yellow Perch were above 50% (Table 7). The maturity schedule for Yellow Perch indicates higher proportion of smaller sized females mature relative to the 2013 survey (Chong et al. 2015). Females were fully mature at about 22 cm in total length. Yellow Perch condition, based on mean relative weight, remained high in 2017, similar to the previous years' surveys (Table 8). The condition was lowest in the Yellow Perch caught in the St. Joseph Channel (64) compared with the Yellow Perch from Lake George, where condition was the highest at 102 (Table 8).

Walleye:

Walleye were captured in all age classes 1-14 with the exception of age 10 year class (Table 9). The majority of the fish captured (92%) were in the 1 through 6 age class, with age-2 Walleye having the highest CPUE during the survey (1.98). Mean length-at-age for Walleye in the 2017 survey was slightly below the state of Michigan average. The growth index, which compares length-at-age to the state average, was -1 mm (Table 9). Total annual mortality for walleye in 2017 (39%) stayed fairly consistent with the 2013 (32%) and 2009 (38%) surveys (Table 6). Walleye maturity began at 34 cm, however full maturity for all fish was not achieved until 49 cm in total length (Table 7). Walleye condition increased in 2017, similar to condition measured in 2006 and

previous surveys, increased from the surveys in 2009 and 2013 (Table 8). Walleye condition was relatively stable across all fishing locations in the St. Marys River in the 2017 surveys (Table 8).

Walleye otoliths were examined for oxytetracycline marks. Otoliths were collected from 118 of the 150 fish caught. Of the 118 fish examined, 41 (34.9%) were from hatchery stock. Hatchery stocked fish were found in every reach except Munuscong Bay. Age-1 had the highest percentage of stocked fish with 54%, followed by age-2 at 38%.

Smallmouth Bass:

Smallmouth Bass were captured in all age classes from one though ten, with a mean age of 4 (Table 10). The majority of the fish captured (91%) were aged 2-6. CPUE was highest for age 3 and age 5 (0.8). Smallmouth Bass growth index was lower than the Michigan State average at -14, however, this growth rate was higher than in previous years (Table 10). Smallmouth Bass total annual mortality increased in 2017 to 0.52 from a value of 0.35 in 2013 (Table 6). The Smallmouth Bass size at 50% maturity was difficult to determine given the variability in the data (Table 7). Female Smallmouth Bass achieved 100% maturity by 30 cm which is slightly below the 36 cm Michigan minimum length limit. Smallmouth Bass continue to exhibit a high condition level (92-108) in the St. Marys River in each of the six fishing locations where they were collected. It has remained stable in all of the river wide surveys since 1995 (Table 8).

Northern Pike:

Northern Pike were found in all age classes from 1 - 10, with the mean age from the catch 4 years (Table 11). CPUE was greatest for age 4 fish (1.0). The majority of fish (90%) were in the age range 2 - 6, with a size range of 445 - 655 cm (Table 11). Northern Pike 2017 (-68) growth index in the St. Marys River as compared to the Michigan Stage average dropped from the index calculated in 2013 (-53), to the lowest calculated since 2009 (-71). In 2017, overall length-at-age was approximately 68 mm smaller than the Michigan statewide average lengths-at-age for Northern Pike (Table 11). Northern Pike annual mortality remained relatively stable in 2017 compared to the previous years' surveys in 2013, 2002 and 1995, but lower than the rates calculated in 2006 and 2009 (Table 6). Maturity of female Northern Pike was inconsistent until 63 cm in total length (Table 7). Northern Pike condition has remained stable across the years in which the survey has been conducted (Table 8). Condition was highest for Northern Pike sampled from Lake George (97), however, is not much higher than the lowest condition, from the Upper River (85) (Table 8).

Cisco:

Cisco age structure was dominated by the 2016 and 2015 year classes (Table 12) but in all, ten cohorts were represented. Generally Cisco grew faster than the state of Michigan average or the Ontario North Channel average rates (Table 12) and condition as indicated by W_r was within the range previously observed (Table 8). Cisco totally annual mortality was 0.39 in 2017, also within the range observed in previous surveys (Table 6). Female Cisco were consistently mature after 33cm in 2017 (Table 7). Cisco condition has remained stable throughout the survey period (Table 8), but was highest in Lake George when compared with the other three collection locations (Table 8).

Length/Weight Regressions

Length/weight regression equations and Von Bertalanffy growth equations for five notable species are presented in Appendix 3. Length frequency distributions for these species from the survey catch are presented in the Appendix 4 Figures.

Stomach contents were analyzed for Walleye, Northern Pike, Smallmouth Bass, Yellow Perch, and Cisco. Contents were reported as incidence (percent void and percent with contents) and proportion of occurrence which is the percent of the identified prey items in the total of all prey items consumed by that species (Table 13). The diet of Walleye, at the time of the survey, was dominated by Rainbow Smelt (18.4%), Threespine Stickleback (10.5%) and Cisco at 7.9% (Table 14). Only Walleye were found with Alewife as a food species (5.3%) in the St. Marys River. Crayfish figured prominently in the diet of all other species examined, except Walleve (Table 13). Northern Pike continue to have the most varied diet, with 10 identified fish species utilized as prey (Table 13). Round Goby have become part of the food chain in Walleye (2.6%), Northern Pike (10.2%), and Yellow Perch (7.0%). The proportion of cravfish in the Smallmouth Bass diet declined from 54.9% in 2013 to 36.8% in 2017 coincident with increases in Yellow Perch (13.2%) and Unidentified Insects (15.8%) in their diet (Table 13, Chong et al. 2015). The 2017 diet of Yellow Perch was comparable to previous years (Chong et al. 2015). In 2017, crayfish returned to a great proportion of the diet at 51.1%, similar to the 2006 diet at 60%. This was an increase from the 2013 survey, where by crayfish had fallen to 9.1% of the diet. Almost 94% of the Cisco examined had empty stomachs. For those with food, the identified species were Mayfly and Water Flea each of which comprised 33.3% of the diet. Unidentified fish were the remainder of the stomach contents at 33.3% (Table 13).

Sea Lamprey Wounding

The incidence of Sea Lamprey wounding among all of the species sampled was low (Table 14). Wounds were observed in four species on five different fish. Wounding was only observed on fishes collected from two sampling locations; four fishes in Potagannissing Bay and one in Munuscong Bay, Wounding rates ranged from 0.1% (White Sucker) to 3.8% (Lake Whitefish), however, the overall wounding rate for Cisco was 4.4% (Table 14). Three fish had A wounds (Ebener et al. 2006); White Sucker and Lake Whitefish (A1) and one Cisco (A2), while the remaining two fish, Cisco and Northern Pike each had one B1 wound. The total Cisco wounding rate was slightly higher than the 2017 rate (3.9%) No Sea Lamprey wounds were observed on Walleye, Yellow Perch, or Rock Bass in 2017 (Table 14). When examining the wounding rate based on location capture, wounding rates increased to 0.4% (White Sucker), 1.7% (Northern Pike), 5.3% (Lake Whitefish) and 12.5% (Cisco).

Discussion

Walleye

Walleye fell to the sixth most abundant species (as measured by full mesh net CPUE; Table 3) in the St. Marys River during the 2017 survey with Yellow Perch, White Suckers, Rock Bass, Brown Bullhead, and Northern Pike all having a higher CPUE in 2017. Overall, Walleye CPUE has remained stable in the survey years 2006-2017, after increasing from a survey low in 2002. While the mean CPUE was the lowest in Lake Nicolet for the seven river regions, it was not significantly so. Overall in 2017, Walleye appear to be well distributed throughout the St. Marys River.

The decline in the CPUE for 2017 and the variance over the survey years can largely be attributed to the CPUE in Lake George. While CPUE has remained relatively stable in the other reaches, in Lake George, CPUE was highly variable with peaks in 2009 (26.7) and 2013 (34.2) compared with lower CPUE in the years 2002 (8.8), 2009 (9.6) and 2017 (7.9). Water depth may play a role in net efficiency in the Lake George, as lake levels have been highly variable over the past 10 years.

Walleye CPUE have been traditionally low in Munuscong Bay while at the same time being a popular walleye destination for anglers. Munuscong Bay consists mainly of shallow, warmer water with the deeper cooler water located in or near the shipping channel. Anglers have reported that

walleye are found in or near the shipping channel during the traditional survey time period (mid-July through late August); however, our inability to safely set nets in or near the shipping channel may be contributing to a lower CPUE for Munuscong Bay.

Mean CPUE of Walleye in the St. Marys River community survey in 2017 was 5.11, after the survey high of 11.25 in 2013. Age-1 to age-6 had the highest CPUE during the survey, corresponding to the 2011-2016 year classes with walleye up to age-14 captured. In comparison to the St. Marys River, Saginaw Bay is a shallow productive bay of Lake Huron that is well known for its walleye fisheries. While being less productive the St. Marys, Saginaw Bay had similar CPUE to the St. Marys River from 1998-2004 (Chong et.al 2014). With the decline of alewife in Saginaw Bay (Fielder and Thomas 2014), overall walleye production in Saginaw Bay far exceeds the found in the St. Marys River.

Mean length-at-age for Walleye in the 2017 survey was slightly below the state of Michigan average. The growth index, which compares length-at-age to the state average, was -1mm. The St. Marys is fed by outflow from Lake Superior; the cold water from Lake Superior may be what leads the St. Marys to being less productive than other bodies of water throughout Michigan. Total annual mortality for Walleye in 2017 (39%) stayed fairly consistent with the 2013 (32%) and 2009 (38%) surveys. Mortality is largely attributed to fish angling pressure and predation.

The St. Marys has received stocked Walleye for decades and more consistently since the Walleye stocking protocol was developed in 2008. Hatchery reared fish are OTC marked prior to stocking for identification purposes. River-wide, 34% of the walleye captured were identified as stocked. Stocked fish were found in every reach except Munuscong Bay, with the Upper River having the highest percentage of stocked fish at 92%. The Walleye stocking program began in 2008 and was completed in 2018. The results of this study are currently in preparation for publication.

Rainbow Smelt were the most common identified prey item (32%) followed by Threespine Stickleback (26%) based on identifiable contents of the stomachs of the walleye captured during the survey. This is consistent with what was observed in the 2013 survey. Condition, as measured by mean relative weight increased in 2017 to the 2006 and previous year levels. Relative weights were uniformly high throughout the river in 2017.

Ontario fishing regulations presently include no length limits in the St. Marys River while Michigan maintains a 38 cm minimum length limit on the same species. Common fishing regulations between Ontario and Michigan within the St. Marys River would provide continuity for this species.

Northern Pike

Coastal wetlands provide critical spawning and nursery habitat for Northern Pike. The Lake Superior outflows via the St. Marys River are set by the International Joint Commission (IJC) and controlled through the Compensating Gates at the head of the rapids. Great Lakes water levels, and Lake Superior in particular increased since 2013(ACE 2019), which has led to an increase in St. Marys River water levels and in increased river flows since 2014 and have remained high through 2017. These higher water levels have led to an increased wetted coastal wetland, which in turn have provided more spawning and nursery habitat for Esocids. In 2017, Northern Pike gillnet CPUE continued to improve, reversing the downward trend that began in 2002 (Table 3). Although the CPUE has not returned to pre-2002 levels, it has rebounded to more than double that of its lowest point, measured in 2009. Northern Pike CPUE continued to increase in four river reaches (Lake Nicolet, St. Joseph Channel, Raber Bay, Potagannissing Bay) and in two reaches (Lake

George and Lake Munuscong) CPUE is comparable to the higher CPUE's measured in 2013 (Table 5). In the Upper River, Northern Pike catch has remained scarce since 1995 (Table 5).

An increase in the number of age-5 and younger pike (Table 10) coupled with the lowest total annual mortality rate calculated for the survey series (Table 7), provide support for the hypothesis that the increased higher water levels have provided for improved Northern Pike juvenile production in the St. Marys River. In addition to the younger fish, fish were collected in all age brackets from six to 10, continuing the improvement trends noted in the 2013 survey, which also found reduced mortality rates from previous surveys (Chong et al. 2015).

Northern Pike growth slowed somewhat in 2017 compared with the previous survey, with overall lengths-at-age being approximately 68 mm smaller than the Michigan statewide average lengths-at-age for that species (Table 10). Maturity of female Northern Pike began at 42 cm but was inconsistent until 63 cm in total length (Table 12), an increase from the 2013 survey when consistent sexual maturity was reached by 55 cm (Chong et al. 2015).

While abundance has not rebounded to the peak of 1987, cautious optimism for continued improvement in Northern Pike populations remain as river levels have remained high through 2018 and potentially continue throughout 2019, based on current Lake Superior water levels.

This survey series has admittedly not been particularly effective at tracking Esocid populations, particularly Muskellunge (Schaeffer et al. 2011). Once again, Muskellunge were not captured during this survey, but they do remain an important part of the fish community and provide a popular sport fishery. The 2017 creel survey of the St. Marys River, which was a companion project to this fish community survey, estimated that 214 muskies were caught in the St. Marys River during the open-water season that year (Godby et al. In Progress). The mean age of Northern Pike captured in the sport fishery was 4.6, while the mean age in this fishery-independent survey was 4.0. Mean length in the sport fishery was 676 mm and was 552 mm in this gillnet survey. These differences could point to gear selectivity; i.e., gill nets aren't effectively sampling older/larger Northern Pike. It is also likely that the sport fishery selects for larger (older) pike. There is also a difference in Northern Pike regulations between Michigan and Ontario. Michigan has a 610 mm (24 in) minimum size limit, while Ontario does not have a minimum size limit. In 2017, approximately 12% of the Northern Pike sampled during the creel survey were less than 610 mm. It is important to continue both the fishery-independent survey and the creel survey of the sport fishery in order to get a complete picture of the fish community.

Yellow Perch

Yellow Perch are an important feature of the St. Marys River fishery. Their recreational harvest ranges from 39,241 to a high of 125,000 (Godby et al. 2019) exceeded in harvest only by that of Cisco. Yellow Perch abundance in the St. Marys River decreased in 2017 but the mean CPUE of 31.5 is just below the survey average of 33.2 and the overall trend remains positive since the first survey in 1975. Potagannissing Bay showed the largest decline in abundance but still has the highest mean CPUE among survey reaches at 56.2. Lake George was a close second to Potagannissing Bay with an average CPUE of 50.0. The Upper River was last surveyed in 2006 and had the 3rd highest Yellow Perch mean CPUE that year, and dropped to the lowest value in the St. Marys in 2017 with a mean CPUE of 6.0 which is also the lowest value recorded for this reach.

Growth, as a density dependent indicator of population status relative to carrying capacity of the habitat and available prey base suggests that the Yellow Perch population of the St. Marys River is not depressed. Mean size at age in 2017 is consistent with the 2013 survey with a few older larger

sized Yellow Perch in the catch which is reflected in the lower total annual mortality rate. Relative to the Michigan average for Yellow Perch the growth index for the St. Marys River remains positive.

Generally, the total annual mortality rates were within sustainable levels; however some reachspecific rates are high and consistent with heavy exploitation.

Smallmouth Bass

Smallmouth Bass abundance in the St. Marys River declined in 2017 after reaching a time-series high in 2013 (Table 3). The declines in Smallmouth Bass abundance occurred in all reaches except the St. Joseph Channel (Table 5). The central portion of the river appears to provide good habitat for Smallmouth Bass. The decrease in river-wide mean CPUE of Smallmouth Bass coincides with an increase in the total annual mortality rate (Table 7). Mean age of Smallmouth Bass has declined in three consecutive surveys with fewer older fish in the catch (Table 9). The Smallmouth Bass diet in the St. Mary's River remains relatively simple still depending heavily on crayfish but with a shift towards a few prey fish species including Yellow Perch, and insects (Table 14).

Cisco

Prior to the large scale collapses of many native fish stocks in Lake Huron in the mid Twentieth Century, Cisco were the most abundant pelagic fish in the lake (Koelz 1929) resulting in substantial commercial fishery yields (Baldwin et al. 2009). After collapse, the Lake Huron stock of Cisco only is found in the northern most regions including the St. Marys River (Dobiesz et al. 2005, Ebener 2012). The exact morphotype of the remnant Lake Huron Cisco is not necessarily consistent with the historic artedii form (Eshenroder et al. 2016) and may reflect local adaption since the larger scale collapse.

Cisco in the St. Marys River appear to be less abundant in recent years although overall trends in gillnet CPUE were not significantly different. Cisco in the St. Marys River will concentrate in cooler deeper water in summer months and make something of an upstream migration for spawning purposes (Fielder 2000). Thus their collection in an August survey may reflect distribution as much as trends in abundance. Cisco usage of the St. Marys River might be affected by climate change. Cisco in the St. Marys River are most consistently encountered when spawning (MDNR unpublished data) or in the recreational fishery during the mayfly (*Hexagenia limbatea*) emergence in midsummer (Fielder et al. 2002).

Growth rate of Cisco improved some in 2017 compared to 2013 and was greater than regional averages (Table 12). This may be consistent with lower Cisco abundances and lower population densities resulting in concomitant faster growth rates. The biological metrics including growth rate, mortality, maturity, and condition do not point to a sustainability concern for Cisco, with the exception of the age structure which is dominated by younger fish that could be an indicator of population stress due to exploitation.

Cisco spawning in Potagannissing Bay have become part of an annual gamete collection along with those from the Les Cheneaux Islands performed by the USFWS to provide for culture and reintroduction in central Lake Huron (USFWS unpublished data). It is not readily clear the extent to which St. Marys River Cisco may comingle with those from the Les Cheneaux Islands and the North Channel but Cisco movement in the region is the subject of a new study being conducted by the USGS Great Lakes Science Center.

AIS

The encroaching presence of invasive species was evident within the St. Marys River during this assessment. As the river is home to international shipping, it remains especially vulnerable to invasive species transfer both from within the Great Lakes and from International ports. Twelve invasive and or non-native species were noted including Round Gobies, which were found to be a diet item (Table 13). Of remark was the discovery of Eurasian Ruffe in the upper River (Table 3), likely due to natural dispersal into the river from Whitefish Bay of Lake Superior where they have been observed in recent years (U.S. Geological Survey 2018). Two other Ruffe were captured in Little Lake George during 2017 by an angler (S. Chong, OMNRF, Personal Communication). The current status and potential impacts of this species is unknown, as no Ruffe were captured during follow-up sampling during late 2017 and in 2018 by the USFWS (A.Bowen, USFWS, Personal Communication).

Round Goby were present in the diet of a low number of predators collected during the 2017 assessment (Table 13). However this information has limited use to reflect the abundance of Round Goby in The River because Gobies were not vulnerable to the sampling gear and were likely underrepresented in the diet as indicated by a high reporting of unidentified fish remains. In the 2017 survey, identified Round Goby were found in fish collected from Potagannissing Bay to Lake Nicolet, indicating that they may be present in a large area of the River. Round Goby first appeared as a dietary item during the 2013 assessment (Chong et al. 2015); and they were collected in other River surveys conducted by the USFWS (Schaeffer et al. 2017) and commonly reported by anglers (U.S. Geological Survey 2018). The appearance of Round Goby and Ruffe indicate that invasive species from both the lower and upper lakes can find their way to the St. Marys River.

White Perch, first documented in 2002, were collected in low numbers during the assessment (Table 3). Other invasives have been documented during other efforts conducted within the river, including but not limited to Tubenose Goby, Dreissenids (*Dreissena polymorpha* and *Dreissena bugensis*), and Rusty Crayfish (*Orconectes rusticus*). Crayfish are an important component of the diet of many fish in the St. Marys River, and the presence of Rusty Crayfish and potential invasion of the Red Swamp Crayfish (*Procambarus clarkia*) pose threats for both native crayfish and habitat in the St. Marys River.

Nuisance blooms of the algae, Didymo (*Didymosphenia geminate*), were documented in the upper St. Marys River shortly after the 2013 survey; the blooms have the potential to develop thick mats over rocks that may affect spawning substrate and invertebrate habitat. Didymo is well suited to cold oligotrophic waters and the Lake Superior source makes the St. Marys potentially ideal for colonization.

Despite the presence of invasive species, the fish community of the St. Marys River is relatively healthy; and historically, invasives have remained low in abundance in the River (Pratt and O'Connor 2011, Ripley et al. 2011, and Schaeffer et al. 2017). Even so, concerns remain regarding potential impacts that newly introduced species or other threatening species may have on the St. Marys River fish community.

Lake Sturgeon

Lake Sturgeon were collected in small numbers in the 1975 through 1987 surveys (Table 2) and again in 2002 (Table 3), however, the largest capture of Lake Sturgeon over the time series occurred in 2017. Lake Sturgeon were captured in 67% of the netting areas below the

Compensation Works, in the lacustrine areas of the River, with the greatest number of fish captured in Lake Munuscong. These areas are more likely representative of Lake Sturgeon habitat during summer months, providing feeding and home range habitats for a variety of juvenile Lake Sturgeon. Lake Sturgeon are more likely to be captured in the faster flowing areas of the river, the Upper River and Lake Nicolet, during the spring spawning migration. Our catch of Lake Sturgeon is likely also limited based on mesh size as our largest mesh fished is 14 cm (5.5'), where most nets targeting Lake Sturgeon use mesh sizes 20 cm (8") and greater (Pratt et al 2016). The increase in Lake Sturgeon CPUE to its highest overall and the increase in both the number of reaches and distribution with those reaches for Lake Sturgeon catch are promising for an increase in abundance within the River.

Sea Lamprey Wounding Rates

Overall, Sea Lamprey wounding rates remain low in the St. Marys River, with 0.2% of the total fish collected exhibiting a Sea Lamprey wound. Of the 29 non-lamprey species collected in 2017, four species had individuals with wounds ranging from A1 to B1 (Ebener et al 2006). Of the seven survey locations, marked fishes were only collected in two places in 2017: Potagannissing Bay and Lake Munuscong. When examining the wounding rate based on the location of capture, wounding rates increased to 0.4% (White Sucker), 1.7% (Northern Pike), 5.3% (Lake Whitefish) and 12.5% (Cisco). Sea Lamprey management within the St. Marys River encompasses both assessment of adult and larval populations within the river as well as targeted treatment of larval populations. Assessment and control of the Sea Lamprey population within the St. Marys River remains a priority for DFO-SLCC and USFWS.

Special Concerns

Of special concern for the St. Marys River and much of the Great Lakes is the potential for introduction and spread of high risk species including Asian carp, Northern Snakehead, Golden Mussel (Limnoperna fortunei), and Killer Shrimp (Dikerogammarus villosus, Currie et al. 2017, Herborg et al. 2007, and Sieracki et al. 2014). Researchers have predicted via modeling that Northern Snakehead, Bighead Carp, Black Carp, Grass Carp and Silver Carp would be compatible with most of the USA, Mexico, and southern Canada (Herborg et al. 2007), which would include the St. Marys River. Grass Carp recruitment has been documented in the Lake Erie watershed (Chapman et al. 2013), and dispersal modeling studies based on origins in western Lake Erie (Maumee Bay) indicate that Grass Carp would not likely move into the St. Marys River area quickly due to food resources present within Lake Erie; however if Grass Carp were to establish in southern Lake Michigan, they may disperse into the St. Marys River area within approximately 10 to 20 years based on natural movements (Currie et al. 2017). Furthermore, tagging studies conducted in the St. Marys River and Welland Canal suggest that Grass Carp may move up and downstream through lock areas via natural dispersal (Currie et al. 2017). Modeling has also predicted a potential pathway for the spread of Golden Mussel and Killer Shrimp within the upper Great Lakes vicinity via ballast water discharge (Sieracki et al. 2014).

Information Needs

As noted in past survey reports; continued monitoring of the fish community in the St. Marys River remains essential. The frequency should be increased in accordance with the original St. Marys River Fishery Assessment Plan (Gebhardt et al. 2002), possibly timed with future years of lake wide intensive monitoring sponsored by the US EPA. More information is needed on reproductive success and recruitment of all species. The addition of a trawling or electrofishing survey would be greatly beneficial to the understanding of the fish community within the river. The creel survey

operated jointly between the MDNR and OMNR has been fragmented in most years and has proved difficult to extract the needed information. Creel survey resources should be saved for when a river-wide survey can be conducted and ideally timed to coincide with the fish community survey. The overall management of the St. Marys River fishery resources would greatly benefit from the development of river-wide joint fish community objectives. These objectives would allow the development of management strategies and a better context with which to interpret findings from the Fish Community Index Surveys. The development of common recreational fishing regulations between Ontario and Michigan remains a need. Development of fish community objectives will drive this effort in addition to the continued assessment of the dynamics of the fish community.

Literature Cited

- Albert, D. A. 2003. Between land and lake: Michigan's Great Lakes coastal wetlands. Michigan Natural Features Inventory, Michigan State University Extension Bulletin E-2902. East Lansing. 96p.
- Baldwin, N. A., R. W. Saalfeld, M. R. Dochoda, H. J. Buettner, and R.L. Eshenroder. 2009. Commercial Fish Production in the Great Lakes 1867-2006 [online]. Available from <u>http://www.glfc.org/great-lakes-databases.php</u>.
- Chapman, D. C., J. J. Davis, J. A. Jenkins, P. M. Kocovsky, J. G. Miner, J. Farver, and P. R. Jackson. 2013. First evidence of grass carp recruitment in the Great Lakes Basin. Journal of Great Lakes Research 39(4):547-554.
- Chong, S., A. Bowen, D.G. Fielder, N. Godby, L. O'Connor, and G. Wright. 2015. Population dynamics of the St. Marys River fish community 1975-2013. Great Lakes Fishery Commission, <u>http://glfc.org/pubs/lake_committees/huron/St%20Marys%20FCS%20Report%202013%2</u> <u>0Draft%20V_4.pdf</u>.
- Currie, W. J. S., J. Kim, M. A. Koops, N. E. Mandrak, L. M. O'Connor, T. C. Pratt, E. Timusk, and M. Choy. 2017. Modeling spread and assessing movement of Grass Carp, Ctenopharyngodon idella, in the Great Lakes basin. DFO Canadian Science Advisory Secretariat Research Document. 2016/114. v + 31 p.
- Czypinski, G. D, A. K. Bowen, M. A. Goehle, and B. Browson. 2006. Surveillance for Ruffe in the Great Lakes, 2006. U. S. Fish and Wildlife Service station report. Fishery Resources Office, Ashland, WI. 44 pp.
- Dobiesz, N.E., D.A. McLeish, R.L. Eshenroder, J.R. Bence, L.C. Mohr, M.P. Ebener, T.F. Nalepa, A.P. Woldt, J.E. Johnson, R.L. Argyle, and J.C. Makarewicz. 2005. Ecology of the Lake Huron fish community, 1970–1999. Canadian Journal of Fisheries and Aquatic Sciences 62: 1432–1451.
- Duffy, W. G., T. R. Batterson, and C.D. McNabb. 1987. The ecology of the St. Mary's River, Michigan: an estuarine profile. U. S. Fish Wildlife Service Biol. Rep. 85(7.10). 138 pp.
- Edsall, T. A., and J. E. Gannon 1993. A Profile of St. Marys River. MICHU-SG-93-700 Michigan Sea Grant College Program. Ann Arbor, Michigan.

- Ebener, M.P. 2012. Status of whitefish and Ciscoes. In The state of Lake Huron in 2010. Edited by S.C. Riley. Great Lakes Fisheries Commission Special Publication 13-01: 29-35.
- Ebener, M.P., E.L. King, Jr., T.A. Edsall. 2006. Application of a dichotomous key to the classification of sea lamprey marks on Great Lakes fish. Great Lakes Fish. Comm. Misc. Publ. 2006-02.
- Environmental Protection Agency (EPA). 2017. St. Marys River Area of Concern. https://www.epa.gov/st-marys-river-aoc
- Eshenroder, R. L., P. Vecesi, O. T. Gorman, D. L. Yule, T. C. Pratt, N. E. Mandrak, D. B. Bunnell, and A. M. Muir. 2016. Ciscoes (Coregonous, subgenus Leucichthys) of the Laurentian Great Lakes and Lake Nipigon. Great Lakes Fishery Commission, Miscellaneous Publication 2016-01. 141 p.
- Fielder, D. G. 1998. Lake herring spawning grounds of the St. Marys River with potential effects of early spring navigation. Michigan Department of Natural Resources, Fisheries Research Report 2049. Ann Arbor.
- Fielder, D. G. 2000. Lake herring spawning grounds of the St. Marys River with some potential implications for early spring navigation. North American Journal of Fisheries Management, 20:552-561.
- Fielder, D. 2002. St. Marys River Task Group: The St. Marys River and challenges of multijurisdictional fisheries. Fisheries, 27(9):32-34.
- Fielder, D. G., A. K. Bowen, K. J. Gebhardt, and S. J. Greenwood. 2002. Harvest of fishes in the St. Marys River, May, 1999 through March 2000. Great Lakes Fishery Commission, Miscellaneous publication. <u>http://www.glfc.org/lakecom/lhc/HarvestReport.pdf</u> . Ann Arbor.
- Fielder, D. G., D. J. Borgeson, A. K. Bowen, S. R. Koproski, S. J. Greenwood, and G. M. Wright. 2004. Populations dynamics of the St. Marys River Fish Community 1975-2002. Great Lakes Fishery Commission, <u>http://www.glfc.org/lakecom/lhc/smr2002rpt.pdf</u>. Ann Arbor.
- Fielder, D. G. and seven coauthors. 2007. Population Dynamics of the St. Marys River Fish Community 1975-2006. Great Lakes Fishery Commission, http://www.glfc.org/lakecom/lhc/SMR2006rpt.pdf Ann Arbor.
- Fielder, D. G., A. Liskauskas, L. C. Mohr, and J. Boase. 2008. Nearshore Fish Community, *In* The state of Lake Huron in 2004. *Edited* by J.R. Bence and L.C. Mohr. Great Lakes Fish. Comm. Spec. Pub. 08-01. pp. 47-51.
- Fielder, D. G., and M. V. Thomas, 2014. Status and Trends of the Fish Community of Saginaw Bay, Lake Huron 2005–2011. Michigan Department of Natural Resources, Fisheries Report 03. Lansing.

- Fielder, D.G., and M.V. Thomas. 2006. Fish Population Dynamics of Saginaw Bay, Lake Huron 1998-2004. Michigan Department of Natural Resources, Fisheries Division, Research Report 2083, Ann Arbor.
- Fielder, D. G., and J. R. Waybrant. 1998. Fish population surveys of St. Marys River, 1975-95, and recommendations for management. Michigan Department of Natural Resources, Fisheries Research Report 2048. Ann Arbor.
- Gebhardt, K., D. Fielder, S. Greenwood, H. Robbins, and T. Sutton [Editors]. 2002. St. Marys River Fisheries Assessment Plan. Great Lakes Fisheries Commission, www.glfc.org/pubs/lake_committees/huron/smrfinal.pdf
- Godby, N., T. Claramunt, D.G. Fielder, S. Chong, A. Bowen, and E. Morrow. In progress. A synthesis of sport fishing activity in the St. Marys River, May through October 2017.
- Godby, N., G. Wright, and D. Fielder. 2009. St. Marys River walleye stocking and evaluation plan. Great Lakes Fishery Commission, www.glfc.org/pubs/lake_committees/huron/SMRwalleye42809.doc
- Great Lakes Water Quality Agreement (GLWQA), 1987. Protocolamended by <u>https://legacyfiles.ijc.org/tinymce/uploaded/GLWQA_e.pdf</u>.
- Grimm, K. S. 1989. A fisheries survey of the St. Marys River, Chippewa County, August-October, 1987. Mich. Dept. Nat. Res. Tech. Rep. No. 89-7.
- Herborg, L. M., N. E. Mandrak, B. C. Cudmore, and H. J. MacIsaac. 2007. Comparative distribution and invasion risk of snakehead (Channidae) and Asian carp (Cyprinidae) species in North America. Canadian. Journal of Fisheries and Aquatic Sciences 64:1723-1735.
- King, E. L. Jr., and T. A. Edsall. 1979. Illustrated field guide for the classification of sea lamprey attack marks on Great Lakes lake Trout. Great Lakes Fishery Commission, Special Publication 79-1. Ann Arbor.
- Koelz, W. 1929. Coregonid fishes of the Great Lakes. Bulletin of the U.S. Bureau of Fisheries 1048: 297–643.
- Liston, C. R., and thirteen coauthors. 1986. Limnological and fisheries studies of the St. Marys River, Michigan in relation to proposed extension of the navigation season, 1982 and 1983.
 Department of Fisheries and Wildlife, Michigan State University. U.S. Fish and Wildlife Service contract no. 14-16-0009-79-013.
- Miller, B. R. 1981. A 1979 fisheries survey of the St. Mary's River system in Chippewa County, and comparisons with 1975 results. Mich. Dept. Nat. Res. Tech. Rep. No. 81-1.
- Ney, J. J. 1999. Practical use of biological statistics. Pages 167-191 in C. C. Kohler and W. A. Hubert, editors. Inland fisheries management in North America, 2nd edition. American Fisheries Society, Bethesda, Maryland.

- Pratt, T. C., W. M. Gardner, J. Pearce, S. Greenwood, and S. C Chong. 2014. Identification of a robust Lake Sturgeon (Acipenser fulvescens Rafinesque, 1917) population in Goulais Bay, Lake Superior. Journal of Applied Icthyology 30:1328-1334.
- Pratt, T. C. and L. M. O'Connor. 2011. An assessment of the health and historical changes of the nearshore fish community of the St. Marys River. Journal of Great Lakes Research. 37:61-69.
- Riley, S. C., and E. F. Roseman. 2013. Status of the offshore demersal fish community. In The state of Lake Huron in 2010. Edited by S. C. Riley. Great Lakes Fishery Commission Special Publication 13-01, pp21-28.
- Ripley, M. P., B. Arbic, and G. Zimmerman. 2011. Environmental history of the St. Marys River. Journal of Great Lakes Research 43(2):5-11.
- Schaeffer, J. S., A. K. Bowen, and D. G. Fielder. 2017. Community stability within the St. Marys River fish community: Evidence from trawl surveys. Journal of Great Lakes Research 43(2):399-404.
- Schaeffer, J. S., D. G. Fielder, N. Godby, A. Bowen, L. O'Connor, J. Parrish, G. Wright. 2011. Long-term trends in the St. Marys River open water fish community. Journal of Great Lakes Research, 37:70-79.
- Schaeffer, J. S., M. W. Rogers, D. G. Fielder, N. Godby, A. Bowen, L. O'Connor, J. Parrish, S. Greenwood, S. Chong, G. Wright. 2014. Designing long-term community assessments in connecting channels: Lessons for the Saint Marys River. Journal of Great Lakes Research, 40:15-22.
- Schneider, J. C., P. W. Laarman, and H. Gowing. 2000. Age and growth methods and state averages. Chapter 9 in J. Schneider, editor. Manual of fisheries survey methods II: with periodic updates. Michigan Department of Natural Resources, Fisheries Special Report 25, Ann Arbor.
- Schorfhaar, R. 1975. A Fisheries Survey of the St. Mary's River, 1975. Michigan Department of Natural Resources, Fisheries Division, Marquette. (Unpublished).
- Sieracki, J. L., J. M. Bossenbroek, and W. L. Chadderton. 2014. A spatial modeling approach to predicting the secondary spread of invasive species due to ballast water discharge. PLoS ONE 9(12): e114217. https://doi.org/10.1371/journal.pone.0114217.
- Sokal, R. R., and F. J. Rohlf. 1981. Biometry. Second Edition. W. H. Freeman and Company, San FranCisco, California.
- SPSS. 2001. SPSS for windows. Release 11.0.1. SPSS Inc., Chicago, Illinois.
- Thomas, M.V., and R. Haas. 2007. Status of the fisheries in Michigan Waters of Lake Erie and Lake St. Clair. Michigan Department of Natural Resources, Lake St. Clair Fisheries Research Station. Prepared for GLFC Lake Erie Committee Meeting, March 22, 2007.

- United States Army Corps of Engineers (USACE). 2019. Monthly mean lakewide average water levels. <u>https://www.lre.usace.army.mil/Missions/Great-Lakes-Information/Great-Lakes-Information/Great-Lakes-Information-2/Water-Level-Data/</u> [Accessed: March 2019].
- United States Geological Survey (U.S. Geological Survey). 2018. Nonindigenous Aquatic Species Database, Gainesville, Florida. <u>https://nas.er.usgs.gov/.</u>
- Van Den Avyle, M. V., and R. S. Hayward. 1999. Dynamics of exploited fish populations. Pages 127-166 in C. C. Kohler and W. A. Hubert, editors. Inland fisheries management in North America, 2nd edition. American Fisheries Society, Bethesda, Maryland.
- Wilkie, C. O, 2018. ggridges: Ridgeline Plots in 'ggplot2'. R package version 0.5.1. hhttps://CRAN.R-project.org/package=ggridges

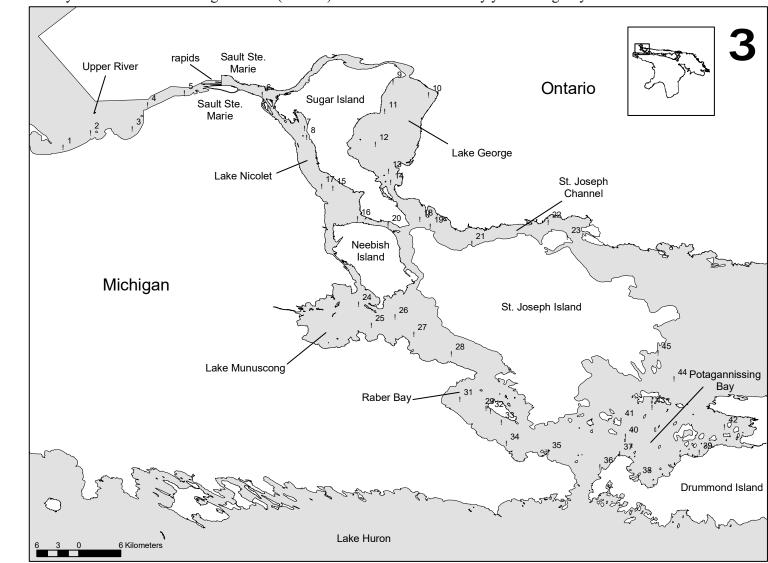


Figure 1. St. Marys River and location of gillnet sets (stations). See Table 1 for effort by year and agency.

! Net set locations

Area	Station numbers	Agency
Upper River	1, 2, 3, 4, 5	MDNR
Lake Nicolet	6, 7, 8, 15, 16, 17, 20	USFWS
Lake George	9, 10, 11, 12, 13, 14	STNRD, OMNRF,
Lake Munuscong	24, 25, 26, 27, 28	MDNR
St. Joseph Channel	18, 19, 21, 22, 23	OMNRF
Raber Bay	29, 31, 32, 33, 34, 35	STNRD
Potagannissing Bay	36, 37, 38, 39, 40, 41, 42, 43, 44, 45	MDNR & OMNRF

Table 1. Net set locations used to define areas within the St. Marys River for the purpose of certain data analyses, along with a list of the agencies that performed the field work in 2017. See Figure 1 for location of each net number.

Table 2. Mean Catch-Per-Unit-of-Effort (CPUE) of all species collected from the St. Marys River 1975 through 2017. Means are based on number per 304.8 m (1000 ft) of gillnet representing the traditional mesh sizes, with standard error of the mean in parentheses. Total nets set were 32 each in 1975 and 1979, 27^b in 1987, 51^c in 1995, 44 in 2002, 2009, 2017, 39 in 2013, and 42 in 2006, although only 34 sets are represented here due to data recording limitations. The St. Joseph Channel portion of the St. Marys was added to the survey series beginning in 2002.

Species ^a	19	75	19	979	19	87 ^b	19	995 °	20)02	2	006	20	009	20)13	20	017
Alewife	1.64	(0.57)	0.23	(0.12)	0.19	(0.11)	15.11	(12.22)	3.92	(3.52)	0.00	(0.00)	0.06	(0.06)	0.39	(0.18)	0.00	(0.00)
Atlantic Salmon	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.09	(0.07)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.07	(0.07)	0.06	(0.06)
Black Crappie	0.03	(0.03)	0.00	(0.00)	0.25	(0.22)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.26	(0.13)	0.06	(0.06)
Bloater	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.06	(0.06)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)
Bluegill	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.06	(0.06)
Bowfin	0.03	(0.03)	0.03	(0.03)	0.40	(0.40)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.06	(0.06)
Brook Trout	0.03	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)
Brown Bullhead	6.41	(3.16)	0.76	(0.50)	6.67	(3.51)	2.56	(1.36)	4.43	(2.28)	3.38	(1.69)	3.52	(2.68)	3.22	(2.18)	10.51	(8.53)
Brown Trout	0.03	(0.03)	0.00	(0.00)	0.03	(0.03)	0.09	(0.07)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)
Burbot	0.05	(0.04)	0.00	(0.00)	0.00	(0.00)	0.05	(0.05)	0.06	(0.06)	0.00	(0.00)	0.17	(0.10)	0.20	(0.15)	0.11	(0.08)
Carp	0.16	(0.08)	0.00	(0.00)	0.03	(0.03)	0.00	(0.00)	0.00	(0.00)	0.07	(0.07)	0.00	(0.00)	0.07	(0.07)	0.00	(0.00)
Channel Catfish	0.00	(0.00)	0.00	(0.00)	0.09	(0.05)	0.00	(0.00)	0.00	(0.00)	0.15	(0.15)	0.00	(0.00)	0.13	(0.13)	0.00	(0.00)
Chinook Salmon	0.00	(0.00)	0.03	(0.03)	0.46	(0.29)	0.08	(0.05)	0.28	(0.12)	0.15	(0.10)	0.06	(0.06)	0.20	(0.11)	0.06	(0.06)
Cisco	14.12	(5.13)	22.40	(11.28)	18.98	(8.34)	9.80	(3.40)	4.38	(2.51)	3.53	(1.84)	10.23	(4.31)	4.08	(2.21)	1.70	(0.59)
Coho Salmon	0.03	(0.03)	0.00	(0.00)	0.00	(0.00)	0.05	(0.05)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)
Freshwater Drum	0.00	(0.00)	0.00	(0.00)	0.03	(0.03)	0.00	(0.00)	0.34	(0.17)	0.59	(0.24)	0.17	(0.10)	0.07	(0.07)	0.00	(0.00)
Gizzard Shad	0.00	(0.00)	0.00	(0.00)	0.12	(0.12)	0.05	(0.05)	0.11	(0.11)	0.00	(0.00)	0.00	(0.00)	0.07	(0.07)	0.00	(0.00)
Lake Sturgeon	0.99	(0.96)	0.03	(0.03)	0.09	(0.05)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.07	(0.07)	0.34	(0.17)
Lake Trout	0.00	(0.00)	0.31	(0.31)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.07	(0.07)	0.17	(0.17)	0.07	(0.07)	0.06	(0.06)
Lake Whitefish	1.15	(0.41)	0.55	(0.25)	2.10	(0.99)	0.73	(0.37)	0.85	(0.41)	0.29	(0.18)	2.33	(1.13)	0.46	(0.21)	0.80	(0.44)
Largemouth Bass	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.07	(0.07)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)
Longnose Gar	0.00	(0.00)	0.03	(0.03)	0.06	(0.04)	0.00	(0.00)	0.06	(0.06)	0.07	(0.07)	0.00	(0.00)	0.07	(0.07)	0.00	(0.00)
Longnose Sucker	0.94	(0.51)	1.07	(0.49)	4.26	(2.46)	2.85	(1.33)	2.10	(1.01)	1.99	(1.26)	2.61	(1.15)	0.13	(0.09)	1.59	(0.79)
Menominee	0.83	(0.44)	0.52	(0.30)	0.00	(0.00)	1.49	(0.55)	0.80	(0.34)	0.22	(0.12)	3.35	(1.80)	0.92	(0.79)	5.23	(2.68)
Muskellunge	0.00	(0.00)	0.68	(0.43)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)

Northern Pike	9.04	(1.77)	8.07	(1.31)	12.69	(2.11)	9.26	(1.64)	2.61	(0.61)	3.82	(0.81)	3.01	(0.75)	5.13	(1.29)	6.99	(1.26)
Pink Salmon	0.00	(0.00)	0.00	(0.00)	2.78	(1.38)	0.55	(0.20)	0.28	(0.15)	0.22	(0.12)	0.06	(0.06)	0.13	(0.09)	0.00	(0.00)
Pumpkinseed	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.97	(0.56)	0.66	(0.66)	0.85	(0.53)	0.00	(0.00)	0.28	(0.19)
Rainbow Smelt	4.97	(2.45)	1.64	(0.69)	1.02	(0.47)	0.86	(0.50)	0.40	(0.21)	0.44	(0.22)	1.65	(1.14)	1.51	(1.06)	2.05	(1.04)
Rainbow Trout	0.03	(0.03)	0.13	(0.07)	0.22	(0.22)	0.00	(0.00)	0.06	(0.06)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)
Redhorse spp.	0.65	(0.29)	0.55	(0.20)	0.62	(0.17)	1.69	(0.53)	0.45	(0.20)	1.25	(0.41)	3.75	(1.19)	1.32	(039)	0.45	(0.17)
Rock Bass	6.20	(2.25)	2.29	(0.67)	11.67	(2.42)	5.57	(1.35)	11.42	(2.77)	14.34	(3.66)	7.84	(1.96)	12.57	(3.56)	7.67	(2.01)
Ruffe	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.06	(0.06)
Sculpin	0.05	(0.04)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)
Sea Lamprey	0.00	(0.00)	0.03	(0.03)	0.00	(0.00)	0.12	(0.09)	0.00	(0.00)	0.00	(0.00)	0.06	(0.06)	0.00	(0.00)	0.57	(0.57)
Smallmouth Bass	0.89	(0.45)	0.26	(0.14)	4.66	(2.23)	3.77	(0.95)	2.27	(0.59)	6.32	(1.76)	1.82	(0.53)	7.76	(2.36)	3.69	(1.17)
Splake	0.34	(0.19)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)
Sucker spp.	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.05	(0.05)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)
Sunfish spp.	0.13	(0.08)	0.13	(0.11)	1.54	(0.89)	0.65	(0.47)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)
Trout-Perch	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.56	(0.56)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)
Walleye	4.27	(1.56)	4.14	(1.73)	7.47	(1.92)	3.92	(0.83)	3.58	(1.04)	11.18	(2.97)	6.02	(1.29)	11.25	(2.88)	5.11	(0.79)
White Bass	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.07	(0.07)	0.23	(0.23)	0.20	(0.15)	0.00	(0.00)
White Crappie	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)
White Sucker	21.48	(3.94)	13.85	(2.20)	25.68	(5.46)	20.00	(2.47)	24.7	(3.93)	17.65	(2.52)	23.07	(3.70)	20.39	(3.84)	22.27	(4.04)
White Perch	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.34	(0.17)	0.74	(0.42)	0.00	(0.00)	0.39	(0.20)	0.11	(0.08)
Yellow Perch	23.02	(6.28)	25.68	(4.93)	49.48	(7.16)	29.97	(5.85)	25.3	(4.50)	37.21	(8.94)	35.34	(7.62)	41.71	(14.95)	31.53	(8.17)
Unknown Sps.	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.11	(0.08)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)	0.00	(0.00)

^a See Appendix 1 for a complete list of common and scientific names of fishes mentioned in this report.

^b Mean CPUEs for 1987 are calculated from a restored data set that lacked five net sets compared to those summarized in Grimm 1987.

^c Mean CPUEs for 1995 included the influence of 3.81 cm (1.5 inch) mesh net on some sets performed in the Raber and Potagannissing area of the river. This effort was incorporated in to the calculation of CPUE but may still have slightly inflated mean CPUE for certain species such as Yellow Perch and Alewife.

Table 3. Mean Catch-Per-Unit-of-Effort (CPUE) of all species collected from the St. Marys River in 2002 - 2017 with all ten mesh sizes included (Expanded mesh) and from the traditional mesh (4 mesh sizes). Means are based number per 304.8 m (1000 ft) of gillnet with standard error of the mean in parentheses. There were 44 total net sets in 2002, 2009, 2017 and 39 in 2013. While 42 nets were set in 2006, however, the traditional mesh CPUE values in 2006 reflect a sample size of 34 net sets, due to data recording limitations.

	200	02	200	06	20	009	201	3	2017		
Species ^a	Expanded mesh	Traditional mesh									
Alewife	10.61 (0.21)	3.92(3.52)	1.12 (0.73)	0.00 (0.00)	0.23 (0.16)	0.06 (0.06)	1.61 (0.72)	0.39 (0.18)	0.18 (0.16)	0.00 (0.00)	
Atlantic Salmon	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.03 (0.03)	0.07 (0.07)	0.02 (0.02)	0.06 (0.06)	
Black Crappie	0.00(0.00)	0.00(0.00)	0.02 (0.02)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.26 (0.15)	0.26 (0.13)	0.02 (0.02)	0.06 (0.06)	
Bloater	0.02 (0.02)	0.06(0.06)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	
Bluegill	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.02 (0.02)	0.06 (0.06)	
Bowfin	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.02 (0.02)	0.06 (0.06)	
Brook Trout	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	
Brown Bullhead	2.59 (1.21)	4.43(2.28)	2.79 (1.13)	3.38 (1.69)	1.89 (1.30)	0.06 (0.06)	3.11 (2.16)	0.00 (0.00)	4.66 (3.57)	10.51 (8.53)	
Brown Trout	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	
Burbot	0.09 (0.04)	0.06(0.06)	0.07 (0.05)	0.00 (0.00)	0.16 (0.06)	0.17 (0.10)	0.24 (0.17)	0.20 (0.15)	0.18 (0.08)	0.11 (0.08)	
Carp	0.05 (0.03)	0.00 (0.00)	0.19 (0.12)	0.07 (0.07)	0.00 (0.00)	0.00 (0.00)	0.05 (0.05)	0.07 (0.07)	0.00 (0.00)	0.00 (0.00)	
Channel Catfish	0.02 (0.02)	0.00 (0.00)	0.31 (0.20)	0.15 (0.15)	0.11 (0.08)	0.00 (0.00)	0.13 (0.07)	0.13 (0.13)	0.09 (0.07)	0.00 (0.00)	
Chinook	0.64 (0.21)	0.28(0.12)	0.29 (0.16)	0.10 (0.08)	0.05 (0.03)	0.06 (0.06)	0.11 (0.06)	0.20 (0.11)	0.05 (0.03)	0.06 (0.06)	
Cisco	2.84 (1.35)	4.38(2.51)	3.62 (1.50)	3.53 (1.84)	6.64 (2.47)	10.23 (4.31)	2.71 (1.51)	4.08 (2.21)	1.02 (0.39)	1.70 (0.59)	
Coho Salmon	0.00 (0.00)	0.00 (0.00)	0.02 (0.02)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.03 (0.03)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	
Freshwater	0.43 (0.18)	0.34(0.17)	1.12 (0.35)	0.59 (0.24)	0.41 (0.15)	0.17 (0.10)	0.37 (0.11)	0.07 (0.07)	0.09 (0.05)	0.00 (0.00)	
Gizzard Shad	0.09 (0.09)	0.11(0.11)	0.02 (0.02)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.03 (0.03)	0.07 (0.07)	0.00 (0.00)	0.00 (0.00)	
Lake Sturgeon	0.02 (0.02)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.11 (0.09)	0.00 (0.00)	0.13 (0.11)	0.07 (0.07)	0.59 (0.27)	0.34 (0.17)	
Lake Trout	0.00 (0.00)	0.00 (0.00)	0.14 (0.09)	0.07 (0.07)	0.16 (0.14)	0.17 (0.17)	0.05 (0.05)	0.07 (0.07)	0.11 (0.11)	0.06 (0.06)	
Lake Whitefish	0.77 (0.35)	0.85(0.41)	0.50 (0.20)	0.29 (0.18)	1.48 (0.66)	2.33 (1.13)	0.42 (0.27)	0.46 (0.21)	0.59 (0.26)	0.80 (0.44)	
Largemouth	0.00 (0.00)	0.00 (0.00)	0.02 (0.02)	0.07 (0.07)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	
Longnose Gar	0.2 (0.02)	0.06(0.06)	0.07 (0.05)	0.07 (0.07)	0.00 (0.00)	0.00 (0.00)	0.11(0.08)	0.07 (0.07)	0.00 (0.00)	0.00 (0.00)	
Longnose	1.20 (0.56)	2.10(1.00)	1.29 (0.59)	1.99 (1.26)	1.61 (0.66)	2.61 (1.15)	0.18 (0.14)	0.13 (0.09)	1.18 (0.68)	1.59 (0.79)	
Menominee	0.36(0.15)	0.80(0.34)	0.86 (0.54)	0.18 (0.11)	1.75 (0.89)	3.35 (1.80)	0.45 (0.35)	0.92 (0.79)	2.55 (1.28)	5.23 (2.68)	
Muskellunge	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.03 (0.03)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	

Table 3 continued.

Northern Pike	1.55(0.33)	2.61(0.61)	1.69 (0.40)	3.82* (0.81)	1.82 (0.37)	3.01 (0.75)	2.66 (0.65)	5.13 (1.29)	4.09 (0.73)	6.99 (1.26)
Pink Salmon	0.39(0.22)	0.28(0.15)	0.14 (0.07)	0.22 (0.12)	0.02 (0.02)	0.06 (0.06)	0.00 (0.00)	0.13 (0.09)	0.00 (0.00)	0.00(0.00)
Pumpkinseed	0.41(0.23)	0.97(0.56)	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.11 (0.07)	0.28 (0.19)
Rainbow Smelt	0.25(0.11)	0.40(0.21)	1.40 (0.51)	0.44 (0.22)	0.84 (0.49)	1.65 (1.14)	1.18 (0.72)	1.51 (1.06)	1.00 (0.50)	2.05 (1.04)
Rainbow Trout	0.02(0.02)	0.06(0.06)	0.00(0.00)	0.00(0.00)	0.00 (0.00)	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00 (0.00)	0.00(0.00)
Redhorse spp.	0.50(0.14)	0.40(0.18)	0.36 (0.20)	0.44 (0.25)	3.07 (1.32)	3.30 (1.21)	0.74 (0.19)	0.86 (0.33)	0.29 (0.13)	0.17 (0.13)
Rock Bass	5.95(1.15)	11.42(2.77)	5.81 (1.32)	14.34 (3.66)	4.14 (1.03)	7.84 (1.96)	7.50 (2.06)	12.57 (3.56)	4.18 (1.09)	7.67 (2.01)
Ruffe	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00 (0.00)	0.00(0.00)	0.00(0.00)	0.00 (0.00)	0.00(0.00)	0.23 (0.23)	0.06 (0.06)
Sculpin	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00(0.00)
Sea Lamprey	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00 (0.00)	0.02 (0.02)	0.06 (0.06)	0.00 (0.00)	0.00 (0.00)	0.05 (0.03)	0.57 (0.57)
Shorthead RH	0.00(0.00)	0.00(0.00)	0.57 (0.22)	0.81 (0.36)	0.30 (0.14)	0.28 (0.12)	0.54 (0.19)	0.46 (0.21)	0.18 (0.08)	0.28 (0.12)
Silver RH	0.02(0.02)	0.06(0.06)	0.00(0.00)	0.00 (0.00)	0.30 (0.14)	0.17 (0.10)	0.05 (0.04)	0.00(0.00)	0.14 (0.06)	0.00(0.00)
Redhorse (all)	0.52 (0.15)	0.45 (0.20)	0.93 (0.28)	1.25 (0.41)	3.66 (1.31)	3.75 (1.19)	1.33 (0.29)	1.32 (0.39)	0.48 (0.14)	0.45 (0.17)
Smallmouth Bass	1.48(0.30)	2.27(0.59)	4.36 (1.21)	6.32 (1.76)	1.73 (0.45)	1.82 (0.53)	6.63 (2.36)	7.76 (2.36)	2.84 (0.83)	3.69 (1.17)
Splake	0.00(0.00)	0.00(0.00)	0.00 (0.00)	0.00(0.00)	0.00 (0.00)	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00(0.00)
Sucker spp.	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00 (0.00)	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00 (0.00)	0.00(0.00)
Sunfish spp.	0.00(0.00)	0.00(0.00)	0.26 (0.22)	0.66 (0.66)	0.39 (0.21)	0.85 (0.53)	0.05 (0.04)	0.00(0.00)	0.00(0.00)	0.00(0.00)
Trout-Perch	0.05(0.03)	0.56(0.56)	0.00(0.00)	0.00(0.00)	0.00 (0.00)	0.00(0.00)	0.00(0.00)	0.00(0.00)	0.00 (0.00)	0.00(0.00)
Unknown	0.05(0.03)	0.11(0.08)	0.00 (0.00)	0.00(0.00)	0.00 (0.00)	0.00(0.00)	0.00 (0.00)	0.00 (0.00)	0.00(0.00)	0.00(0.00)
Walleye	2.55(0.65)	3.58(1.04)	6.07 (1.35)	11.18 (2.97)	4.89 (1.09)	6.02 (1.29)	7.58 (1.81)	11.25 (2.88)	3.41 (0.50)	5.11 (0.79)
White Bass	0.02(0.02)	0.00(0.00)	0.02 (0.02)	0.07 (0.07)	0.30 (0.19)	0.23 (0.23)	0.11 (0.08)	0.20 (0.15)	0.00(0.00)	0.00(0.00)
White Crappie	0.02(0.02)	0.00(0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00(0.00)
White Sucker	18.80(2.09)	24.77(3.93)	17.88 (2.47)	17.65 (2.52)	18.07 (2.84)	23.07 (3.70)	17.39 (3.53)	20.39 (3.84)	16.04 (2.22)	22.27(4.04)
White Perch	0.16(0.09)	0.34(0.17)	0.50 (0.22)	0.74 (0.42)	0.05 (0.05)	0.00 (0.00)	0.26 (0.10)	0.39 (0.20)	0.16 (0.09)	0.11 (0.08)
Yellow Perch	23.43(4.25)	25.34(4.50)	39.92 (7.15)	37.21 (8.94)	37.20 (7.03)	35.34 (7.62)	48.11(12.18)	41.71	29.34 (5.65)	31.53 (8.17)

• * In 2006, Northern Pike CPUE was significantly higher in the traditional net vs. the full net set.

Species	Year	Upper River	Lake Nicolet	Lake George	Lake Munuscong	St. Joseph Channel	Raber Bay	Potagannissing Bay
Yellow Perch	2017	6.0 (3.6)	10.0 (8.8)	50.0 (23.0)	10.5 (2.2)	21.5 (5.9)	44.2 (14.8)	56.2 (30.0)
	2013		9.3 (3.1)	38.3 (13.2)	26.0 (8.8)	6.9 (5.2)	41.2 (13.1)	88.5 (54.4)
	2009	35.0 (32.6)	5.0 (2.3)	81.2 (26.5)	22.5 (3.2)	11.5 (2.3)	61.7 (16.4)	31.8 (18.8)
	2006	40.0 (16.8)	29.5 (12.9)	66.2 (28.2)	25.0 (5.4)	16.5 (5.7)	57.0 (46.0)	1.2 (1.2) ^b
	2002	26.5 (11.1)	20.7 (7.8)	42.5 (20.5)	17.0 (4.6)	54.5 (18.3)	17.9 (7.3)	11.8 (6.0)
	1995	39.0 (17.2)	21.6 (10.2)	42.3 (22.6)	20.3 (2.5)		27.0 (6.8) ^a	29.6 (11.5)
	1987	33.9 (15.9)	30.4 (27.1)	65.0 (19.0)	30.0 (4.9)		41.4 (4.8)	62.5 (16.3)
	1979	43.1 (9.0)	18.9 (9.5)	26.2 (11.0)	9.2 (2.1)		9.8 (5.0)	37.3 (11.7)
	1975	25.3 (16.6)	13.9 (10.0)	31.8 (10.0)	11.2 (6.0)		6.0 (3.6)	33.5 (16.4)
Northern Pike	2017	0.0 (0.0)	6.7 (2.4)	10.0 (2.5)	10.0 (4.3)	7.5 (2.8)	6.7 (2.0)	3.2 (2.1)
	2013		4.3 (3.1)	10.0 (4.5)	11.5 (5.3)	6.9 (2.8)	2.1 (0.8)	0.8 (0.8)
	2009	0.0 (0.0)	0.7 (0.5)	7.08 (2.08)	7.0 (3.2)	4.5 (1.8)	3.8 (1.4)	0.5 (0.5)
	2006	1.0 (0.6)	2.5 (1.4)	4.2 (1.4)	5.0 (2.2)	10.0 (2.8)	1.5 (0.6)	$0.0~(0.0)^{\mathrm{b}}$
	2002	0.0 (0.0)	0.4 (0.4)	21.7 (14.7)	0.0 (0.0)	7.5 (6.3)	0.4 (0.4)	2.2 (1.8)
	1995	2.5 (1.6)	8.1 (3.4)	16.3 (4.5)	18.4 (5.5)		12.8 (3.4)	1.6 (1.2)
	1987	6.9 (5.0)	2.9 (2.1)	27.0 (5.2)	15.6 (3.0)		11.7 (3.2)	8.0 (3.0)
	1979	1.9 (0.3)	4.7 (3.5)	14.3 (3.3)	11.8 (4.6)		6.0 (2.6)	6.5 (1.4)
	1975	4.4 (4.0)	11.7 (7.1)	17.3 (7.8)	9.3 (2.6)		5.0 (3.0)	7.1 (2.4)
Walleye	2017	5.0 (5.0)	2.1 (0.8)	7.9 (1.5)	2.0 (0.5)	5.5 (1.4)	6.7 (1.4)	6.5 (1.8)
	2013		1.8 (0.7)	34.2 (12.9)	0.5 (0.5)	6.2 (2.2)	15.8 (4.5)	8.8 (3.8)
	2009	6.0 (3.0)	1.4 (0.7)	9.6 (5.6)	1.0 (1.0)	6.0 (2.0)	17.9 (3.9)	2.5 (1.2)
	2006	15.5 (6.2)	4.0 (1.7)	26.7 (14.0)	4.2 (1.7)	3.5 (1.9)	8.5 (4.4)	18.8 (6.2) ^b
	2002	2.5 (2.5)	1.1 (0.5)	8.8 (3.6)	1.0 (1.0)	3.0 (1.5)	7.9 (5.6)	1.8 (1.2)
	1995	2.5 (0.8)	5.6 (3.1)	2.0 (6.9)	2.8 (0.9)		3.6 (1.1)	5.4 (2.1)
	1987	1.1 (0.7)	0.8 (0.0)	8.0 (3.5)	3.1 (1.4)		21.9 (8.0)	6.3 (2.4)

Table 4. Mean catch-per-unit-of-effort is number per 304.8 m (1000 ft.) collected from the seven habitat areas of the St. Marys River 1975 - 2017 based on catch from traditional mesh sizes. Standard error of the mean is in parentheses.

Table 4 continued								
	1979	0.0 (0.0)	1.1 (0.7)	4.0 (2.8)	2.9 (1.0)		5.6 (2.8)	6.3 (4.8)
	1975	0.0 (0.0)	4.7 (2.0)	5.0 (4.0)	2.9 (1.8)		2.1 (1.4)	6.5 (4.1)
Smallmouth Bass	2017	0.0 (0.0)	1.1 (0.7)	7.1 (4.8)	3.0 (1.5)	9.0 (4.2)	2.5 (1.3)	3.8 (3.5)
	2013		2.9 (1.8)	16.2 (8.8)	10.5 (3.9)	8.1 (6.9)	4.2 (1.9)	6.8 (6.5)
	2009	1.5 (0.6)	0.0 (0.0)	0.4 (0.4)	2.0 (1.5)	5.0 (2.1)	3.8 (2.1)	1.3 (1.3)
	2006	0.5 (0.5)	4.0 (2.0)	5.0 (1.7)	13.8 (4.6)	16.5 (5.7)	2.5 (1.6)	1.3 (1.3) ^b
	2002	0.0 (0.0)	1.1 (0.7)	4.2 (2.9)	4.5 (1.4)	4.5 (1.8)	2.5 (2.0)	0.8 (0.4)
	1995	0.0 (0.0)	3.1 (3.1)	3.5 (2.0)	8.1 (2.8)		5.9 (4.5)	2.5 (1.0)
	1987	0.6 (0.3)	2.1 (1.2)	15.5 (10.6)	7.9 (5.3)		2.3 (0.4)	0.2 (0.1)
	1979	0.0 (0.0)	0.0 (0.0)	0.0 (0.0)	0.3 (0.3)		0.0 (0.0)	0.6 (0.4)
	1975	0.0 (0.0)	0.0 (0.0)	0.3 (0.2)	1.8 (1.2)		0.0 (0.0)	1.4 (1.1)
Cisco	2017	0.5 (0.5)	0.0 (0.0)	4.6 (1.9)	0.0 (0.0)	4.0 (4.0)	0.0 (0.0)	2.5 (1.0)
	2013		0.4 (0.4)	2.5 (1.3)	2.1 (1.5)	6.9 (6.9)	16.7 (12.8)	1.0 (0.7)
	2009	0.0 (0.0)	2.1 (1.5)	0.0 (0.0)	0.0 (0.0)	2.0 (0.9)	14.2 (7.0)	34.0 (16.8)
	2006	0.0 (0.0)	0.5 (0.5)	0.8 (0.5)	0.0 (0.0)	0.5 (0.5)	22.0 (9.4)	$0.0~(0.0)^{\mathrm{b}}$
	2002	0.5 (0.0)	0.0 (0.0)	0.0 (0.0)	0.0 (0.0)	0.0 (0.0)	0.0(0.0)	3.2 (1.2)
	1995	0.0 (0.0)	13.4 (5.9)	3.5 (3.2)	0.0 (0.0)		11.7 (9.3)	19.2 (9.8)
	1987	0.0 (0.0)	0.8 (0.8)	3.3 (2.9)	0.8 (0.6)		1.2 (1.0)	54.0 (21.1)
	1979	0.0 (0.0)	3.1 (3.1)	0.0 (0.0)	0.0 (0.0)		62.7 (62.4)	39.8 (23.8)
	1975	0.0 (0.0)	9.2 (8.3)	0.0 (0.0)	0.1 (0.1)		42.5 (17.8)	23.0 (11.7)

^a Means from these areas included some efforts of 3.51 c, (1.5 in.) mesh. While compensated for in the calculation of CPUE, the influence of the smaller mesh may have slightly inflated the mean for certain species such as Yellow Perch. ^b Potagannissing Bay mean CPUE values for 2006 reflect only two net sets via the traditional mesh sizes and was probably under-sampled for

^b Potagannissing Bay mean CPUE values for 2006 reflect only two net sets via the traditional mesh sizes and was probably under-sampled for the purpose of this reach specific analysis.

Table 5. Catch-per-unit-of-effort (CPUE) of Yellow Perch by age for 2017 and mean length-at-age at capture for the St. Marys River, August-September, 1979-2017 by river location. For comparison, mean length-at-age is included from past surveys and the Michigan state average length-at-age¹ as well as the Ontario Lake Huron 2006 North Channel average² (ON NC). Unit of effort is one 304.8 m gillnet set. Growth index¹ compares length-at-age to Michigan state average and the 2013 year to the North Channel average. It excludes age groups represented by less than 5 specimens. All lengths and the growth indexes are in mm. CPUE values by age may omit some un-aged fish and therefore may not total to the overall CPUE for this species as reported in Table 3.

Parameter &					<u>Ag</u>	<u>e</u>					Mean age	Mean length	Growth index
Area	1	2	3	4	5	6	7	8	9	10			
Upper River													
Number	1	10	2	2	5	2	1						
CPUE	0.2	0.4	0.6	0.8	1	1.2	1.4						
Frequency (%)	4.3	43.5	8.7	8.7	21.7	8.7	4.3						
<u>Mean length</u>													
2017	140	156	226	187	257	296	285				3.0	204	+10
2013													
2009		149	195	210							3.1	188	+1
2006	159	186	241	251							2.7	219	+40
2002	146	170	222	251	343		361		373	372	3.0	212	+28
1995	-	157	184	200	225	244	269	280	298	354			-7
1987				201	216	224	254	264	305	312			-20
1979			183	201	216	259	272	302	295				-6
MI average	127	160	183	208	234	257	277	292	302				
ON NC 2006	124	173	211	235	243	248	256	276		290			
Lake Nicolet													
Number		11	7	2	11	5	2	1					
CPUE		1.6	1.0	0.3	1.6	0.7	0.3	0.1					
Frequency (%)		28.2	17.9	5.1	28.2	12.8	5.1	2.6					
Mean length													
2017		152	207	214	248	278	311	326			4.0	221	+13
2013		150	170	191							3.3	181	-13
2009		153	171	202							3.3	181	-8
2006	143	164	205	235							2.6	188	+17
2002		148	162	197	238	239	328				3.3	177	-10
1995	170	147	172	209	227	250	275	284					-7
1987				196	221	231	287	295					-7
1979			168	185	221	208	244						-18
MI average	127	160	183	208	234	257	277	292	302				
ON NC 2006	124	173	211	235	243	248	256	276		290			-36

Table 5. Continued.

Parameter &					<u>A</u> g	ge					Mean age	Mean length	Growth index
Area	1	2	3	4	5	6	7	8	9	10			
Lake George			-						-				
Number	2	60	27	42	17	16	18	3	1				
CPUE	0.3	5.6	21.2	7.0	2.0	1.8	1.8	1.0	0.2				
Frequency	1.1	32.3	14.5	22.6	9.1	8.6	9.7	1.6	0.5				
(%)													
Mean length													
2017	145	148	182	216	249	273	271	394	316		3.8	192	+3
2013		151	171	204	280	291	286	287			3.6	170	+8
2009		148	173	217	263	286					3.5	182	+9
2006	156	172	207	246	246	272					2.3	188	+22
2002	155	153	194	222	269	311	318	315			2.8	185	+12
1995		148	169	206	233	247	242	263	256				-15
1987				198	216	256	264	302	323				-10
1979			173	190	203	249	282	282		297			-12
MI average	127	160	183	208	234	257	277	292	302				
ON NC 2006	124	173	211	235	243	248	256	276		290			+4
St. Joseph													
Channel													
Number	2	29	15	17	14	11	5			2			
CPUE	0.4	5.8	3.0	3.4	2.8	2.2	1.0			0.4			
Frequency	2.1	30.5	15.8	17.9	14.7	11.6	5.3			2.1			
(%)													
<u>Mean length</u>													
2017	149	159	174	199	234	261	260			327	3.8	200	-5
2013		148	157	158	183		231				3.6	167	-37
2009		148	153	165	178	190					3.7	162	-42
2006	149	155	174	194	212	283					2.9	167	+0
2002		147	167	217	259	293					3.2	183	+8
1995													
1987													
1979													
MI average	127	160	183	208	234	257	277	292	302				
ON NC 2006	124	173	211	235	243	248	256	276		290			-48

Table 5. Continued.

					<u>Ag</u>	<u>e</u>					Mean	Mean	Growth
Parameter &											age	length	index
Area	1	2	3	4	5	6	7	8	9	10			
Lake													
Munuscong													
Number	6	118	7	15	9	3	1						
CPUE	1.2	23.6	1.4	3	1.8	0.6	0.2						
Frequency (%)	3.8	74.2	4.4	9.4	5.7	1.9	0.6						
<u>Mean length</u>													
2017	140	148	173	205	224	274	207				2.5	162	-4
2013		155	177	194	231						3.2	166	-7
2009		142	172	209	265						3.3	184	+1
2006	155	182	227								2.5	205	+31
2002	153	146	180	208	230		275				2.6	1.66	-6
1995		145	177	213	229	239	256	292	278				-11
1987				196	226	279	292	325					+10
1979		203	193	216	239	284	254						+9
MI average	127	160	183	208	234	257	277	292	302				
ON NC 2006	124	173	211	235	243	248	256	276		290			-26
Raber Bay													
Number	2	37	15	63	12	5	4		1				
CPUE	0.3	6.2	2.5	10.5	2.0	0.8	0.7		0.2				
Frequency (%)	1.4	26.6	10.8	45.3	8.6	3.6	2.9		0.7				
Mean length													
2017	136	148	206	209	238	270	293		305		3.6	194	+6
2013		158	194	238	261						3.0	188	+17
2009													
2006	157	182	207	223	244	273					3.1	204	+20
2002		152	175	203	246	268					3.3	185	-2
1995	137	152	202	227	236	260	268	269					+4
1987		-	165	188	231	251	277	297	307	315			-9
1979		185	196	221	272	262							+17
MI average	127	160	183	208	234	257	277	292	302				- /
ON NC 2006	124	173	211	235	243	248	256	276		290			-3

Table 5 Continued.

					Age	2					Mean age	Mean length	Growth index
Parameter &											C	C	
Area	1	2	3	4	5	6	7	8	9	10			
Potagannissing Bay													
Number		81	162	72	22	17	6		1				
CPUE		8.1	16.2	7.2	2.2	1.7	0.6		0.1				
Frequency (%)		22.4	44.9	19.9	6.1	4.7	1.7		0.3				
<u>Mean length</u>													
2017		154	185	229	301	315	300		379		3.3	190	+28
2013		160	220	230							2.8	190	+20
2009		152	177	204	239	326					4.6	175	+11
2006	143	181	229	263							2.4	202	+37
2002	157	172	196	247	297	175					2.6	189	+32
1995	133	158	167	208	215	243	275	290					-6
1987					231	262	272	307		330			-1
1979			201	224	249	269	302	323	282				+20
MI average	127	160	183	208	234	257	277	292	302				
ON NC 2006	124	173	211	235	243	248	256	276		290			-3
River-wide													
Number	13	346	235	213	90	59	37	4	3	2			
CPUE	0.3	7.9	5.3	4.8	2.0	1.3	0.8	0.1	0.1	0.0			
Frequency (%)	1.3	34.5	23.5	21.3	9.0	5.9	3.7	0.4	0.3	0.2			
<u>Mean length</u>													
2017	141	151	186	216	255	284	277	377	333	327	3.3	190	+9
2013		156	186	208	249	275	280	281					+3
2009		150	172	204	237	251							-6
2006		155	174	220	236	246	280	290					-1
2002	151	153	177	220	258	274	320	315	373	372	3.0	184	+15
1995	140	152	171	211	227	246	260	278	294	354			-7
1987			165	195	223	244	273	296	308	319			-6
1979		196	196	209	229	264	285	302	291	297			+7
MI average	127	160	183	208	234	257	277	292	302				
ON NC 2006	124	173	211	235	243	248	256	276		290			-1

¹From Schneider et al. (2000) ²Ontario MNR, unpublished data

Species	Area, if not total for the river	1995	2002	2006	2009	2013	2017
Yellow						Not	
Perch	Upper River	0.25	0.54	0.70	0.63	sampled	0.59
	Lake Nicolet	0.38	0.70	0.59		0.61	0.39
	Lake George	0.40	0.52	0.43	0.69	0.55	0.42
	C	Not					
	St. Joseph Channel	sampled	0.64	0.50		0.71	
	Lake Munuscong	0.41	0.61	0.78	0.62	0.63	0.76
	Raber Bay	0.44	0.63	0.49		0.71	0.50
	Potagannissing Bay	0.60	0.57	0.96	0.67	0.55	0.59
	River Total	0.38	0.68	0.70	0.64	0.60	0.41
Northern							
Pike		0.58	0.52	0.61	0.72	0.52	0.51
Walleye		0.51	0.49	0.38	0.38	0.32	0.39
Cisco		0.31	0.39	0.40	0.48	0.25	0.38
Smallmouth Bass		0.36	0.37	0.55	0.50	0.35	0.52

Table 6. Comparison of total annual mortality (A) rates for select fish species in the St. Marys River, computed from fish collected in experimental mesh gillnets 1995-2017.

			Species		
		Smallmouth	-		
Length (cm)	Walleye	Bass	Northern Pike	Yellow Perch	<u>Cisco</u>
13				50	
14				79	
15				63	
16				76	
17				73	
18				87	
19		0		77	0
20				94	0
21		100		97	100
22		0		98	
23	0			100	0
24				93	
25		0		100	100
26	0	33		100	
27				100	0
28		100		100	100
29		100		94	67
30	0	87		100	50
31	100	100		92	50
32	33	100		100	0
33	80	100		100	100
34	100	100		100	100
35	25	100		100	100
36	50	100		100	100
37	0	100		100	100
38	0	100		100	100
39	25	100	0		100
40		100			100
41		100			100
42		100	50		100
43	100	100			100
44	80	100			100
45	100	100			100
46	100	100	0		100
47	100	100	50		100

Table 7. Maturity schedule for five notable species expressed as percent maturity of females by length in the St. Marys River. Fish used in the analysis were collected by gillnets in August - September 2017.

			Species		
		Smallmouth	*		
Length (cm)	Walleye	Bass	Northern Pike	Yellow Perch	Cisco
48	67		100		
49	100	100	75		
50	100		100		
51	100		50		
52	100		83		
53	100		100		
54	100		83		
55	100		100	100	
56	100		100		
57	100		100		
58	100		100		
59	100		67		
60	100		100		
61	100		100	100	
62	100		33		
63	100		100		
64	100		100		
65	100		100		
66	100		100		
67			100		
68			100		
69			100		
70			100		
71			100		
72			100		
73			100		
74			100		
75			100		
76			100		
77			100		
78			100		
79			100		
80			100		
81			100		
82			100		
83			100		

Table 7. Continued.

Location	Walleye	Yellow Perch	Smallmouth Bass	Northern Pike	Cisco
Upper River	86	97		85	
Lake Nicolet	91	95	108	94	
Lake George	94	102	101	97	114
Lake Munuscong	84	97	106	96	
St. Joseph Channel	77	64	92	87	79
Raber Bay	87	95	97	96	86
Potagannissing Bay	86	92	98	96	82
River wide 2017	87	93	100	95	90
River wide 2013	56	96	103	94	87
River wide 2009	57	90	112	101	91
River wide 2006	87	91	109	94	84
River wide 2002	90	94	106	87	89
River wide 1995	102	97	106	91	

Table 8. Mean relative weight of select species, by area and river wide, for the St. Marys River, August - September 2017; River wide total values for 1995-2017 are presented for comparison.

Table 9. Catch-per-unit-of-effort (CPUE) of Walleye by age for 2017 and mean length-at-age at capture for the St. Marys River, August-September, 1979-2017. For comparison, mean length-at-age is included from past surveys and the Michigan state average length-at-age¹ as well as the Ontario Lake Huron 2006 North Channel (ON NC) average². Unit of effort is one 304.8 m gillnet set. Growth index¹ compares length-at-age to state average and the 2017 year to the NC average. It excludes age groups represented by less than 5 specimens. All lengths and the growth index are in mm. CPUE values by age may omit some un-aged fish and therefore may not total to the overall CPUE for this species as reported in Table 3.

							А	ge							Mean	Mean	Growth
Parameter	1	2	3	4	5	6	7	8	9	10	11	12	13	14	age	length	index
Number	13	34	21	20	24	24	5	1	1		2	2	1	2			
CPUE	0.3	1.9	0.9	0.6	0.5	0.6	0.1	0.1	0.0		0.1	0.1	0.1	0.1			
Frequency (%)	8.7	22.7	14.0	13.3	16.0	16.0	3.3	0.7	0.7		1.3	1.3	0.7	1.3			
Mean length																	
2017	234	334	381	428	477	509	492	557	582		631	608	586	657	4.2	463	-1
2013	253	335	420	450	450	513	531	576		592					4.0	408	+3
2009		309	394	439	485	529	536	576		592					4.5	440	+2
2006	287	363	391	416	483	520		561							3.0	383	+9
2002	253	312	393	472	530	421	563	552		590	578	660	571	614	4.0	434	+15
1995	209	271	278	363	489	502	560	611		604							-26
1987	240	288	347	407	464	505	549	585	607	660							-17
1979		307	378	447	472	528	513	538									-27
MI average	250	338	386	437	472	516	541	561	582								
ON NC 2006		381	410	471	511	538		635	-	658							-35
average																	
¹ From Schneider et	tal (200)())															

¹From Schneider et al. (2000)

Table 10. Catch-per-unit-of-effort (CPUE) of Smallmouth Bass by age 2017 and mean length-at-age at capture for the St. Marys River, August - September, 1987-2017. For comparison, mean length-at-age is included from past surveys and the Michigan state average length-at-age¹ as well as the Ontario Lake Huron North Channel (ON NC) average². Unit of effort is one 304.8 m gillnet set. Growth index¹ compares length-at-age to state average and excludes age groups represented by less than 5 specimens. All lengths and the growth index are in mm. CPUE values by age may omit some un-aged fish and therefore may not total to the overall CPUE for this species as reported in Table 3.

							A	Age							Mean	Mean	Growth
Parameter	0	1	2	3	4	5	6	7	8	9	10	11	12	13	age	length	index
Nīssasla az		6	10	27	21	22	11	1	1	2	1						
Number		6	12	37	21	33	11	1 <0.1	1 <0.1	2	1						
CPUE		0.1	0.3	0.8	0.5	0.8	0.3	< 0.1	< 0.1	< 0.1	<1.0						
Frequency (%)		4.8	9.6	29.6	16.8	8.8	8.8	0.8	0.8	1.6	0.8						
Mean length																	
2017		190	244	296	317	369				438					4.0	323	-14
2013		148	234	276	349	385	420	430	445	463					4.4	335	-11
2009				271	300	344	363								4.5	313	-44
2006		171	251	282	315	371		391							3.0	273	-18
2002		146	187	222	291	325	376	398	457			457			4.1	281	-61
1995		145		245	263	278	305	340	359								-99
1987				234	268	330	347	371									-72
MI average		178	257	305	356	386	406	434	452	475							
ON NC 2003		128	161	175	256	291	240										+94
average																	

¹From Schneider et al. (2000)

Table 11. Catch-per-unit-of-effort (CPUE) of Northern Pike by age 2017 and mean length-at-age at capture for the St. Marys River, August - September, 1987-2017. For comparison, mean length-at-age is included from past surveys and the Michigan State average length-at-age¹ as well as the Ontario Lake Huron North Channel (ON NC) average². Unit of effort is one 304.8 m gillnet set. Growth index¹ compares length-at-age to state average and the 2017 year to the NC average. It excludes age groups represented by less than 5 specimens. All lengths and the growth index are in mm. CPUE values by age may omit some un-aged fish and therefore may not total to the overall CPUE for this species as reported in Table 3.

Parameter	0	1	2	3	4	5	<u>Age</u> 6	7	8	9	10	11	12	13	Mean age	Mean length	Growth index
Number		5	16	34	42	27	9	4	2	2	1						
CPUE		0.1	0.4	0.8	1.0	0.6	0.2	< 0.1	< 0.1	< 0.1	< 0.1						
Frequency (%)		3.5	11.3	23.9	29.6	19.0	6.3	2.8	1.4	1.4	0.7						
Mean																	
length																	
2017		378	445	503	561	612	655	637	660	763	851				4.0	552	-68
2013			455	525	598	610	685								4.1	583	-53
2009		287	436	520	619										3.0	543	-71
2006	269	429	528	601	642										1.8	491	+13
2002	250	371	455	564	620	669									2.4	477	-34
1995		399	465	538	605	621	722	918		1033							-39
1987		407	468	515	575	672	726	752	754								-39
MI		422	511	579	635	683	732	780									
average																	
ON NC		377	483	580	657	749	706										-60
2002																	
average	• 1	1 (20															

¹From Schneider et al. (2000)

Table 12. Catch-per-unit-of-effort (CPUE) of Cisco by age 2017 and mean length-at-age at capture for the St. Marys River, August - September, 1995-2017. For comparison, mean length-at-age is included from past surveys and the Michigan state average length-at-age¹ as well as the Ontario Lake Huron 2006 North Channel (ON NC) average². Unit of effort is one 304.8 m gillnet set. Growth index¹ compares length-at-age to state average and the 2017 year to the NC average. It excludes age groups represented by less than 5 specimens. All lengths and the growth index are in mm. CPUE values by age may omit some un-aged fish and therefore may not total to the overall CPUE for this species as reported in Table3.

							A	<u>lge</u>							Mean	Mean	Growth
Parameter	0	1	2	3	4	5	6	7	8	9	10	11	12	13	age	length	index
NT 1		10		•				•	•								
Number		10	15	3	2	3	4	3	2	1			l				
CPUE		0.2	0.3	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1			< 0.1				
Frequency (%)		22.7	34.1	6.8	4.5	6.8	9.1	6.8	4.5	2.3			2.3				
Mean length																	
2017		214	304	318	324	401	407	419	416	472			417		3.5	318	+32
2013		196	249	272	269	293	314	351	390	384	388	427			4.7	292	-15
2009		207	260	343	366	379	382	398	404	413					3.7	316	+41
2006		213	232	281	326	387	378	386	377	412					3.2	280	+8
2002		199	240	306	338	374	383	412	416						3.1	292	+26
1995		200	265	330	289	327	379	399	401	412	446						+16
MI average		214	241	267	294	321	347	374	400								
ON NC 2006			265	263	329	292	358	377	372	388	372	390	374	393			-17
average																	
¹ From Schneider e	t al (2	000)															

¹From Schneider et al. (2000)

	Walleye	Northern Pike	Smallmouth Bass	Yellow Perch	Cisco
Incidence					
No. stomachs examined	141	180	122	716	32
% void	73.0	72.8	68.9	62.0	93.8
Percent of Occurrence					
Unidentified fish remains	44.7	44.9	28.9	15.8	33.3
Crayfish		16.3	36.8	51.1	
Alewife	5.3				
Rainbow Smelt	18.4	4.1			
Mayfly					33.3
Logperch		4.1			
Unidentified zooplankton					
Menominee		2.0			
Trout-Perch		6.1			
Yellow Perch	7.9	10.2	13.2	2.2	
Cisco					
Sculpin	2.6	2.0			
Johnny Darter	5.3	2.0			
Unidentified insects	2.6	2.0	15.8	26.1	
Ninespine Stickleback					
Threespine Stickleback	10.5	10.2			
Stickleback sp.			2.6	0.4	
Snails		2.0	2.6		
White Sucker		2.0			
Round Goby	2.6	10.2		7.0	
Vegetation		2.0	2.6	0.4	
Water Flea					33.3

Table 13. Incidence and percent of occurrence of food items (based on stomach content identification) for select species from the St. Marys River, August – September 2017. Note percent occurrence may total more than 100% due to multiple food species in stomach.

Species	Ν	A1	A2	A3	A4	B1	B2	B3	B4	Total
Walleye	150	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Yellow Perch	1291	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Cisco	45	0.0	2.2	0.0	0.0	2.2	0.0	0.0	0.0	4.4
Lake Whitefish	26	3.8	0.0	0.0	0.0	0.0	0.0	0.0	0.0	3.8
Northern Pike	180	0.0	0.0	0.0	0.0	0.6	0.0	0.0	0.0	0.6
White Sucker	706	0.1	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.1
Rock Bass	184	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0

Table 14. Percent of sea lamprey wounds by species exhibiting wounding from the St. Marys River, August - September 2017. N denotes sample size of specimens examined for wounds. Wounds scored according to Ebner et al. (2006).

Appendix 1. Common and scientific names of fishes and other aquatic organisms mentioned in this report.

Appendix 1 continued.

White Perch	Morone americana
White Sucker	Catostomus commersoni
Yellow Perch	Perca flavescens

		Cumula	tive Full Me	esh Nets		Cumulative Traditional Mesh Nets					
Species	2002	2006	2009	2013	2017	2002	2006	2009	2013	2017	
Alewife	467	47	10	61	8	69		1	6		
Atlantic Salmon				1	1				1	1	
Black Crappie		1		10	1				4	1	
Bloater	1					1					
Bluegill				2	1					1	
Bowfin					1					1	
Brown Bullhead	114	117	85	118	205	78	46	62	49	185	
Burbot	4	3	7	9	8	1		3	3	2	
Carp	2	8		2			1		1		
Channel Catfish	1	13	5	5	4		2		2		
Chinook Salmon	28	12	2	4	2	5	2	1	3	1	
Cisco (Lake Herring)	125	152	292	103	45	77	48	180	62	30	
Coho Salmon		1		1							
Creek Chub			9								
Freshwater Drum	19	47	18	14	4	6	8	3	1		
Gizzard Shad	4	1		1		2			1		
Lake Sturgeon	1		5	5	26				1	6	
Lake Trout		6	7	2	5		1	3	1	1	
Lake Whitefish	34	21	65	16	26	15	4	41	7	14	
Largemouth Bass		1					1				
Longnose dace			1								
Longnose Gar	1	3		4		1	1		1		
Longnose Sucker	53	54	71	7	52	37	27	46	2	28	
Moxostoma sp.	22	15	135	29	7	7	6	58	13	3	

Appendix 2. Total catch and cumulative species by net set type: Full Mesh Nets (all 10 mesh panels) and Traditional Mesh Nets (4 meshes, extrapolated to 304.8 m) for the surveys years 2002 to 2017. High-lighted are new species for each year.

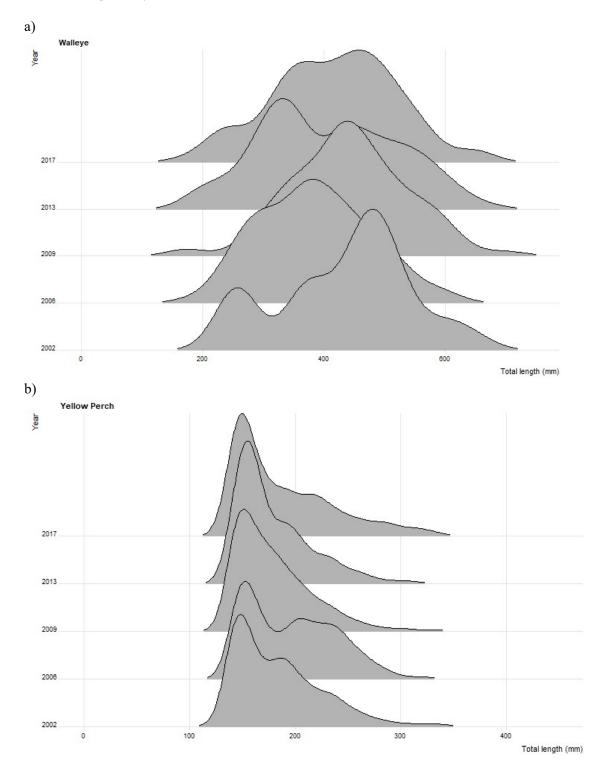
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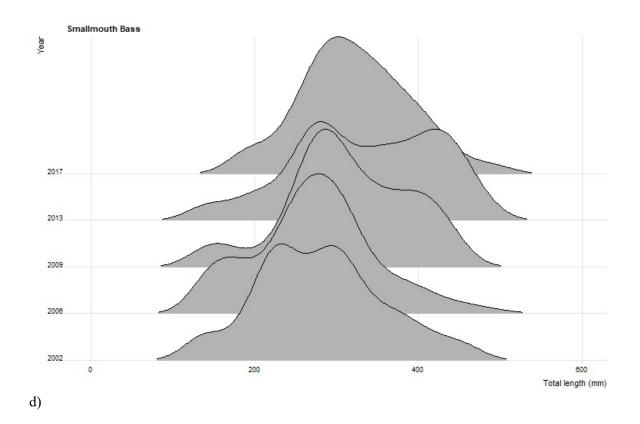
Muskellunge				1						
Northern Pike	68	71	80	101	180	46	52	53	78	123
Pink Salmon	17	6	1	6		5	3	1	2	
Pumpkinseed	18	11	17	2	5	17	9	15		5
Rainbow Smelt	11	59	37	45	44	7	6	29	23	36
Rainbow Trout	1					1				
Rock Bass	262	245	182	285	184	201	195	138	191	135
Round Whitefish (Menonimee)	16	36	77	17	112	14	3	59	14	92
Ruffe					10					1
Sea Lamprey			1		2			1		1
Shorthead Redhorse		24	13	21	8		11	5	7	5
Silver Redhorse	1		13	2	6	1		3		
Smallmouth Bass	65	183	76	252	125	40	86	32	118	65
Trout-perch	2					1				
Unknown fish species	2	1				2				
Walleye	112	254	215	288	150	63	152	106	171	90
White Bass	1	1	13	4			1	4	3	
White Crappie	1									
White Perch	7	21	2	10	7	6	10		6	2
White Sucker	827	751	795	661	706	436	240	407	310	392
Yellow Perch	1031	1677	1637	1828	1291	446	506	622	634	555
Total Catch	3318	3842	3871	3917	3226	1585	1421	1873	1715	177
Species Count	32	31	29	34	30	27	25	24	29	26
Cumulative Total	32	37	40	43	45	27	33	34	37	40
New Species	0	5	3	3	2	0	6	1	3	3

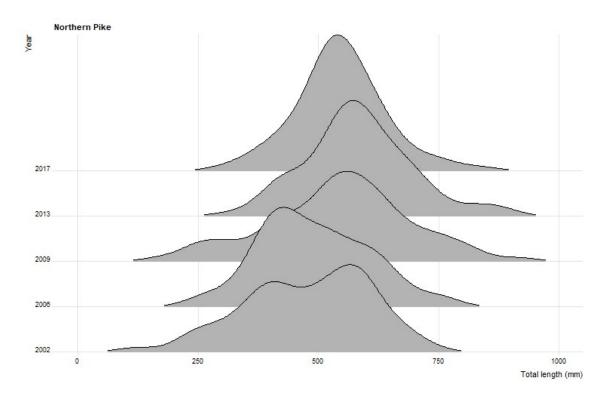
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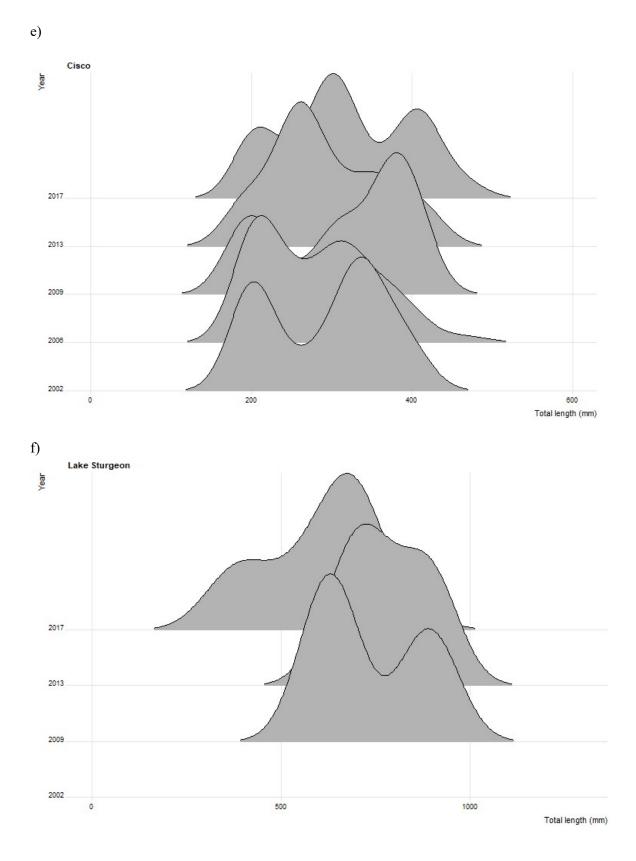
Species	Length/Weight Equation	Len/Wt r ²	Von Bertalanffy Equation	К	Γ∞	t _o
Walleye	log(wt)=3.223 log(len)-5.626	0.98	$L_t = 610 [1 - e^{-0.2708(t-0.26)}]$	0.2708	610	0.26
Yellow Perch	log(wt)=3.118 log(1en)-5.170	0.90	$L_t = 348 [1 - e^{-0.2293(t-0.26)}]$	0.2293	348	0.26
Smallmouth Bass	log(wt)=3.240 log(1en)-5.435	0.96	$L_t = 545 [1 - e^{-0.1703(t+1.51)}]$	0.1703	545	-1.51
Northern Pike	log(wt)=3.044 log(1en)-5.355	0.94	$L_t = 721 [1 - e^{-0.2537(t+1.91)}]$	0.2537	721	-1.91
Cisco	log(wt)=3.291 log(1en)-5.759	0.87	$L_t = 464 [1 - e^{-0.3348(t+2.32)}]$	0.3348	464	-2.32

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Appendix 3a: St. Marys River Area of Concern: Coastal Wetland Habitat Assessment Report (Darwin, 2016)

St. Marys River Area of Concern: Coastal Wetland Habitat Assessment Report

August 2016





Environment and Climate Change Canada Environnement et Changement climatique Canada

St. Marys River Area of Concern: Coastal Wetland Habitat Assessment Report

August 2016

Environment and Climate Change Canada – Canadian Wildlife Service

Executive Summary

The St. Marys River is a 112 km connecting channel between Lake Superior and Lake Huron. In 1985, the International Joint Commission (IJC) identified the St. Marys River as one of the 43 Areas of Concern (AOC) on the Great Lakes. The condition of wildlife populations, a subcomponent under Beneficial Use Impairment (BUI) #3 (*Degradation of Fish and Wildlife Populations*), is currently listed as "requires further assessment" while BUI #14 (*Loss of Fish and Wildlife Habitat*) is currently listed as "impaired". Given that coastal wetlands provide a number of ecological functions as well as wildlife habitat, it is important to gain a better understanding of their current state within the AOC and surrounding area. Building upon work started in 2012 (see Environment Canada 2013, 2014, 2015), a subset of coastal wetlands on the Canadian side of the St. Marys River, both within and outside the AOC, were visited and surveyed for water quality, submerged aquatic vegetation, aquatic macroinvertebrates, amphibians (frogs and toads), and marsh breeding birds in 2015 to assess the condition of coastal wetland habitat and biotic communities.

Using the Water Quality Index (WQI), water quality within St. Marys River coastal wetlands ranged from moderately degraded to very good, and indicated that water quality in AOC versus non-AOC wetlands was similar. The degradation of water quality appears to be primarily a result of increased turbidity which may be a result of natural processes or anthropogenic causes. An Index of Biotic Integrity (IBI), a multimetric index indicating the ability of a habitat to support and maintain a balanced, integrated, adaptive biological system having the full range of elements expected in a region's natural habitat, was used to report on the condition of the submerged aquatic vegetation (SAV) community. The SAV IBI demonstrated variation among sites, with wetland condition ranging from poor to excellent. From 2015 data, it was determined that no suitable IBIs could be developed for macroinvertebrate, marsh bird, or amphibian communities due to the lack of a strong response to disturbance. The IBIs developed in previous work (Environment Canada 2014) did not respond well with 2015 data, and therefore cannot be used to report on the condition of these biotic communities. Biotic communities lacking a strong response to disturbance, combined with wetlands within the AOC and outside the AOC exhibiting comparable WQI values, suggest that the Canadian side of St. Marys River can be considered no longer impaired with respect to the status of the Loss of Fish and Wildlife Habitat BUI's first criterion regarding coastal wetland wildlife habitat conditions. Further, this also suggests the marsh bird and amphibian (frogs and toads) coastal wetland communities can be considered not impaired with respect to the status of the Degradation of Fish and Wildlife Populations BUI.

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Cover Photos: Environment and Climate Change Canada – Canadian Wildlife Service

1. Introduction

The St. Marys River is a 112 km connecting channel between Lake Superior and Lake Huron. In 1985, the International Joint Commission (IJC) identified the St. Marys River as one of the 43 Areas of Concern (AOC) within the Great Lakes basin. On the Canadian side, the AOC extends from the head of the river at Whitefish Bay to Quebec Bay and includes the waters partway surrounding St. Joseph Island (Figure 1). The area was listed because of historical problems associated with pollution from industrial sources, partially treated municipal and private sewage, and physical alterations to the river and its shoreline (St. Marys River Binational Public Advisory Committee 2015).

A standardized set of impairments called Beneficial Use Impairments (BUIs) were created by the IJC as a measure to assess the health of the Great Lakes. The BUIs cover a wide range of environmental and ecological concerns. The Remedial Action Plan (RAP) process seeks to restore these beneficial uses through various remedial and monitoring actions recommended in the Stage 2 RAP report. In the St. Marys River AOC, nine of the fourteen beneficial uses defined in the Great Lakes Water Quality Agreement (GLWQA) are listed as impaired including degradation of fish and wildlife populations and loss of fish and wildlife habitat.

The condition of wildlife populations, a subcomponent under BUI #3 (*Degradation of Fish and Wildlife Populations*), is currently listed as "requires further assessment" and does not contain delisting criteria at the time of this report. BUI #14 (*Loss of Fish and Wildlife Habitat*) is currently listed as "impaired", and the delisting criteria are as follows: i) coastal wetland wildlife habitat conditions within the Area of Concern are comparable to those of suitable reference sites, as assessed using an index of biotic integrity; ii) rapids habitat conditions are enhanced through feasible conservation and restoration measures identified in the Stage 2 Remedial Action Plan; and iii) the closely linked "Degradation of Fish Populations" BUI is no longer deemed impaired (St. Marys River Binational Public Advisory Council 2015).

Given that coastal wetlands provide a number of ecological functions as well as wildlife habitat, it is important to gain a better understanding of their current state within the AOC and surrounding area. Standardized methodologies for surveying in coastal wetlands have been developed (Environment Canada and Central Lake Ontario Conservation Authority (herein, EC and CLOCA) 2007). Using these methodologies, a subset of coastal wetlands in the St. Marys River, both within and outside the AOC, were surveyed from 2012 to 2015 to collect information on water quality, submerged aquatic vegetation (SAV), aquatic macroinvertebrates (2013-2015 only), marsh breeding birds, and amphibians (2013-2015 only). The results of these surveys can be used to provide components of information necessary for BUI assessments.

Numerous attempts to develop disturbance gradients with multiple years of data (Cvetkovic and Midwood 2014, 2015a, 2015b) in the St. Marys River have had limited success. Compared to other AOCs in the Great Lakes, the St. Marys River demonstrates both a narrower range of disturbance and higher overall water quality. Indices of Biotic Integrity (IBIs) are ideally

developed based on a wide continuum of disturbance that elicits a response in both degraded and pristine sites within a region. Most sites in the St. Marys River, however, exhibit relatively undisturbed condition. The absence of a broad range of disturbance necessary to capture change and variation in the St. Marys River results in the inability to create suitable IBIs for this area, in particular for the marsh breeding bird, aquatic macroinvertebrate, and amphibian biotic communities. The SAV community does show response to disturbance and may have limited use in reporting on wetland condition. The lack of wide and well-distributed disturbance and the subsequent difficulty in developing IBIs that are sensitive to change provide evidence of coastal wetland habitat and biotic communities in the Canadian side of the St. Marys River AOC being no longer impaired with respect to criterion (i) of BUI #14: *Loss of Fish and Wildlife Habitat* and provides information to support a not impaired assessment with respect to coastal wetland bird and amphibian (frogs and toads) populations in BUI #3: *Loss of Fish and Wildlife Populations*.

2. Purpose of Report

The purpose of this document is to briefly describe the sampling methodologies and report on the condition of coastal wetlands, where available. These assessments allow for comparisons of coastal wetland quality between survey years as well as between AOC and non-AOC coastal wetlands on the Canadian side of the St. Marys River. This report also suggests that there is a narrow range of relatively minimally disturbed coastal wetlands in the AOC, which supports the assertion that coastal wetlands are no longer impaired with respect to criterion (i) of BUI #14 and coastal wetland bird and amphibian (frogs and toads) populations in BUI #3.

3. Surveyed Wetlands

Since it was not feasible to survey all coastal wetlands within the St. Marys River, a subset of sites that collectively provide a geographic spread throughout the river and represent the geomorphic types and sizes of coastal wetlands present on the Canadian side was selected. More details on site selection and description of sites are provided in Environment Canada, 2013. Six AOC and five non-AOC wetlands (one added in 2013¹) were surveyed from 2012 to 2015 for water quality and biotic communities (Figure 1) for a total of 43 site-years:

- AOC sites:
 - Carpin Beach
 - Echo Bay
 - Lake George
 - Maskinonge Bay
 - Pumpkin Point
 - West Shore, St. Joseph Island

¹ An addition non-AOC wetland was added in 2013 to improve AOC/non-AOC comparisons.

- Non-AOC sites:
 - Anderson Creek (added in 2013)
 - Desbarats Wetland
 - Stobie Creek
 - Hay Bay Wetland
 - Joe Dollar Bay Wetland

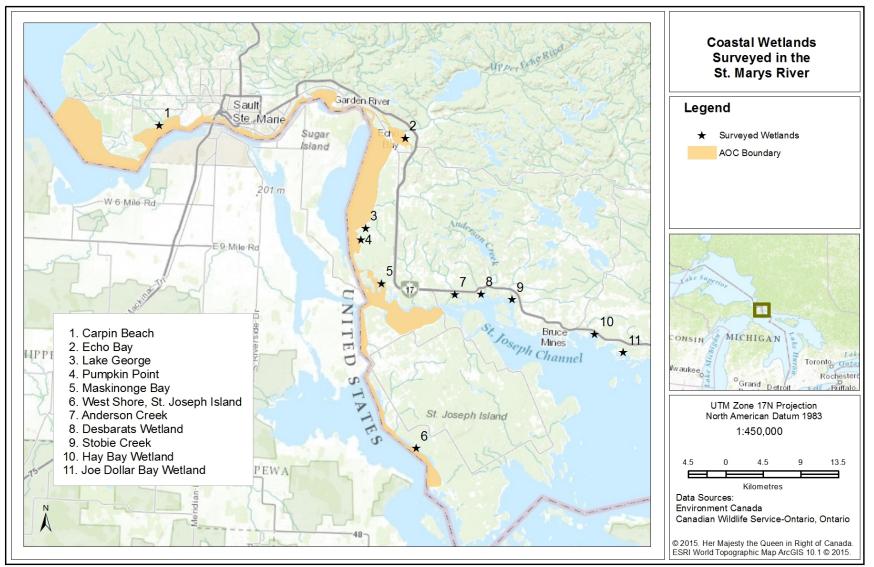
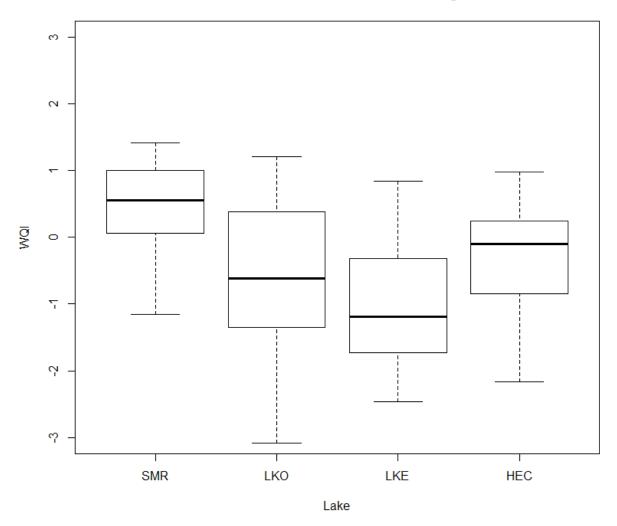


Figure 1: Location of coastal wetlands sampled in the St. Marys River from 2012-2015.

4. Disturbance and Wetland Condition in the St. Marys River

IBIs can be used to monitor the condition of coastal wetlands and have been used regionally (e.g., Durham Region's Coastal Wetland Monitoring Program) and in other AOCs (e.g., Bay of Quinte, Wheatley Harbour, Detroit River, St. Clair River, and Thunder Bay), as well as in lake-wide basin assessments (e.g., Environment Canada's Coastal Habitat Assessment and Monitoring Project). Previous efforts to develop IBIs specific to the St. Marys River (Cvetkovic and Midwood 2014, 2015a, 2015b) have had limited success, with only the SAV community showing a response to disturbance while marsh breeding birds, amphibians, and aquatic macroinvertebrates showed weak to no sensitivity to disturbance. The most common gradient of disturbance used in previous work is the Water Quality Index (WQI; Equation 1), a tool for determining coastal wetland water quality in the Great Lakes (Chow-Fraser 2006). The WQI was developed as a relative ranking tool to report on coastal wetland water quality in the Great Lakes Basin, further discussed in section 5. WQI scores fit into six categories which correspond with values ranging from -3 to +3, with lower scores indicating higher degradation, and are therefore useful in quantitatively determining the range of disturbance in an area.

Figure 2 highlights the distinct differences in the range of disturbance, as measured by the WQI, in the St. Marys River versus other Canadian areas in the lower Great Lakes (i.e., Lake Ontario, Lake Erie, the Huron-Erie Corridor), with the St. Marys River demonstrating a much narrower range of disturbance as well as overall better water quality.



WQI values in the lower Great Lake regions

Figure 2: Boxplots of the mean WQI values corresponding to each lake/region, where SMR = St. Marys River, LKO = Lake Ontario, LKE = Lake Erie, and HEC = Huron-Erie Corridor. Boxplot "whiskers" indicate the maximum/minimum values, and the top and bottom of the boxplot indicate the 25th and 75th percentiles, respectively.

The development of IBIs includes, in part, examining the response of biotic community attributes over a continuum of disturbance. As site disturbance increases, the likelihood of observing sensitive taxa (e.g., species) decreases. For example, in lower Great Lakes, it is well known that as disturbance increases, overall coverage of SAV decreases. In the St. Marys River region, the disturbance continuum is truncated, wherein most sites experience relatively little disturbance. Without more highly disturbed sites (such as in the lower Great Lakes), associations between disturbance and biotic attribute cannot be statistically discerned. While there has been success in developing IBIs in the lower Great Lakes, the limited range of disturbance in the St. Marys River has resulted in the inability to develop suitable IBIs for this

geographic area. Although several potential IBIs have been developed in the recent past (Cvetkovic and Midwood 2014, 2015a, 2015b), attempts at validation with 2015 data resulted in strong indication that these potential IBIs did not respond well to disturbance as predictors of wetland condition. Consequently, it was determined that IBIs as indicators of change in biotic communities, with the exception of the SAV community, cannot be developed for the St. Marys River. Although this may be frustrating from an IBI development standpoint, it is very encouraging from an impaired beneficial use perspective.

5. Water Quality

Methodology

Water quality was measured using both *in situ* probes and chemical analyses. *In situ* water quality determination included four parameters (pH, conductivity [µS/cm], temperature [°C], and turbidity [NTU]) and was collected using a Hydrolab MS5[™] multiprobe at mid depth of the water column adjacent to emergent vegetation.

Multiprobe sampling was conducted at all water quality stations. The four parameters measured are used to calculate the Water Quality Index (WQI; Equation 1), a tool for determining coastal wetland water quality in the Great Lakes (Chow-Fraser 2006).

Equation 1:

WQI = (-1.37 * log TURB) - (1.58 * log COND) - (1.63 * log TEMP) - (2.37 * log pH) + 9.27

where TURB = turbidity, COND = conductivity, and TEMP = temperature

Water samples for additional nutrient parameters (Table 1; Table 2) were collected in 2012 at four of the six stations at each wetland and include Total Nitrate Nitrogen (TNN), Total Ammonia Nitrogen (TAN), and Total Phosphorus (TP). TNN and TAN were analyzed in a field lab within five hours of sampling using colorimetry (Hach DR890 Colorimeter); samples for TP were stabilized through acidification and later analyzed by Environment Canada's National Laboratory for Environmental Testing (NLET; Burlington, Ontario). From 2013 and onward, nitrate and ammonia values were analyzed at the site level using a composite water sample collected from each station at each wetland. Individual samples of TP in 2013-2015 were collected and analyzed using the same methodology as in 2012.

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Parameter	Units	Relationship with Increased Disturbance
In Situ		
Turbidity	NTU	$ m \uparrow$ turbidity from algae, suspended sediments, and bioturbation
Conductivity	μS/cm	↑ conductivity from agricultural, industrial, urban inputs (e.g., fertilizer salts and road salt)
Temperature	°C	↑ temperature from industrial/urban runoff and riparian vegetation removal
рН	рН	\uparrow in pH from photosynthesis affects nutrient availability
Nutrient		
Total Nitrate Nitrogen	mg/L	↑ nitrates from agricultural/urban runoff and wastewater and industrial discharge
Total Ammonia Nitrogen	mg/L	↑ ammonia from agricultural and industrial wastes; and sewage and septic leachate
Total Phosphorus	mg/L	↑ phosphorus from agricultural runoff, urban stormwater, and industrial discharge

Table 1: Water quality parameters measured in coastal wetlands including parameter relationships with increased disturbance.

Disturbance Variable	Description
Total Phosphorus	The concentration (mgL ⁻¹) of all forms of phosphorus dissolved in the sample. This is an important indicator of enrichment in surface waters.
Ammonia	The concentration (mgL ⁻¹) of ammonia nitrogen in the sample. Ammonia can be toxic to aquatic organisms and is released into waterways by many industries, primarily municipal wastewater treatment plants.
Nitrate	The concentration of nitrate nitrogen (mgL ⁻¹) in the sample. The primary sources of nitrates in the environment are sewage, fertilizer, and manure.
Turbidity	A measure of the degree to which light traveling through a water column is scattered by the suspended organic (including algae) and inorganic particles measured in Nephelometric Turbidity Units (NTU).
Conductivity	A measure of the dissolved ions in water measured in microSiemens per centimetre (μScm ⁻¹) or milliSiemens per centimetre (mScm ⁻¹). Conductivity is a good indicator of urban run-off, especially from road salt.

Table 2: Descriptions of water quality parameters used to score and rank water quality.

Ranking Water Quality

The WQI was developed as a relative ranking tool to report on coastal wetland water quality in the Great Lakes Basin. WQI scores fit into six categories which correspond with values ranging from -3 to +3 (Table 3).

WQI Score	Qualitative Descriptor
+3 to +2	Excellent
+2 to +1	Very good
+1 to 0	Good
0 to -1	Moderately degraded
-1 to -2	Very degraded
-2 to -3	Highly degraded

 Table 3: Water Quality Index (WQI) score and associated category based on Chow-Fraser (2006).

Results

In 2015, coastal wetland WQI qualitative descriptors (Table 3) varied from moderately degraded to very good with the majority of sites ranked in either good or very good condition (Table 5). Non-AOC wetlands varied from moderately degraded to very good, while AOC wetland condition was good or very good. Impaired water quality from the WQI is typically the result of elevated turbidity such as seen at Hay Bay Wetland, with a turbidity value greater than 15 NTUs and a WQI descriptor of moderately degraded (Table 4).

Total phosphorus (TP) values in 2015 were at or below the Provincial Water Quality Objective limit of 0.03 mg/L (Table 6) for all wetlands except Maskinonge Bay, which had a total phosphorus level of 0.07 mg/L. The higher level of TP at Maskinonge Bay is likely due to an output source in the vicinity of the wetland, which has consistently been the source of high levels of phosphorus during all sampling years at this wetland. Overall, total ammonia nitrogen

values in 2015 were 0.03 mg/L or less, and total nitrate nitrogen values varied from 0.02 mg/L to 0.22 mg/L. As noted in the Methods section, these parameters were collected from a composite water sample in 2013-2015 compared to individual samples in 2012, which may account for the slight differences observed among years; values from 2014 to 2015 appear to be comparable among the same wetlands.

Statistical comparisons (paired t-tests) of the WQI and each water quality parameter were performed for AOC versus non-AOC wetlands for each sampling year, and then for the mean of all sampling years. No statistical difference was observed between wetlands located in the AOC versus outside the AOC for the WQI or any water quality parameter for any year.

Wetland Name	Turbidity (NTU)				Co	Conductivity (µS/cm)			Water Temp (°C)				рН			
	2012	2013	2014	2015	2012	2013	2014	2015	2012	2013	2014	2015	2012	2013	2014	2015
AOC sites																
Carpin Beach	6.3	7.2	5.7	13.5	130.5	125.5	110.0	107.8	23.4	18.6	18.0	21.1	7.3	7.2	7.3	7.2
Echo Bay	4.5	8.9	7.2	6.1	115.2	84.8	84.2	78.1	25.7	22.0	21.0	21.4	8.5	8.3	7.3	7.1
Lake George	50.6	38.7	14.3	7.6	150.0	129.1	111.8	103.7	23.5	21.7	19.0	19.8	7.8	7.7	7.3	7.5
Pumpkin Point	51.3	44.9	16.5	12.1	123.1	116.9	94.0	101.6	29.1	26.7	18.4	20.0	9.1	9.0	8.1	8.2
Maskinonge Bay	1.7	1.6	2.9	1.9	110.3	105.8	106.8	104.0	24.0	24.7	18.4	22.9	8.5	8.0	7.6	7.7
West Shore, St. Joseph Island	37.9	69.3	137.1	14.5	190.0	164.9	136.2	114.8	22.9	22.7	17.1	23.0	8.5	8.1	7.9	7.6
Non-AOC sites																
Anderson Creek	-	12.8	6.6	4.3	-	133.8	120.3	103.5	-	21.8	17.5	21.2	-	7.4	7.7	7.4
Desbarats Wetland	3.2	3.6	3.2	2.8	129.3	99.3	99.5	102.3	25.5	22.9	19.0	19.9	8.1	8.0	7.5	7.3
Stobie Creek	2.5	2.8	2.8	2.7	152.8	113.6	98.8	106.4	30.7	25.5	17.5	21.3	9.2	9.1	7.4	7.5
Hay Bay Wetland	31.9	19.2	12.2	17.3	195.1	135.0	129.5	136.9	24.3	25.6	16.2	21.2	8.2	8.6	7.7	7.9
Joe Dollar Bay Wetland	8.8	2.3	2.4	2.3	156.1	135.1	133.5	145.0	26.4	25.2	19.2	21.6	8.4	8.2	7.6	7.7

Table 4: Mean water quality parameters for selected coastal wetlands in the St. Marys River Area of Concern (AOC) and non-AOC sites in the St. Marys River. Wetlands are ordered from west to east.

		WQI										
	2012	2013	2014	2015	Mean	Descriptor*						
AOC Sites												
Carpin Beach	0.55	0.69	0.92	0.33	0.62	Good						
Echo Bay	0.63	0.56	0.86	1.02	0.77	Good						
Lake George	-0.85	-0.51	0.33	0.70	-0.08	Moderately degraded						
Pumpkin Point	-1.03	-0.84	0.28	0.34	-0.31	Moderately degraded						
Maskinonge Bay	1.30	1.36	1.28	1.40	1.34	Very good						
West Shore, St. Joseph Island	-0.90	-1.11	-1.15	0.13	-0.76	Moderately degraded						
Non-AOC sites												
Anderson Creek	-	0.15	0.74	0.99	0.63	Good						
Desbarats Wetland	0.79	0.99	1.26	1.33	1.09	Very good						
Stobie Creek	0.56	0.85	1.42	1.24	1.02	Very good						
Hay Bay Wetland	-0.82	-0.36	0.38	-0.08	-0.22	Moderately degraded						
Joe Dollar Bay Wetland	0.01	0.97	1.22	1.09	0.82	Good						

Table 5: Water Quality Index (WQI) score and descriptor for selected coastal wetlands in the St. Marys River Area of Concern (AOC) and non-AOC sites in the St. Marys River.

* based upon mean WQI value for years sampled

Table 6: Additional water quality collected for selected coastal wetlands in the St. Marys River Area of Concern (AOC) and non-AOC wetlands in the St. Marys River. TP=Total Phosphorus, NH₃-N=Total Ammonia Nitrogen, NO₃-N=Total Nitrate Nitrogen.

Wetland Name			NO ₃ -N	(mg/L)		TP (mg/L)						
	2012*	2013	2014	2015	2012*	2013	2014	2015	2012	2013	2014	2015
AOC sites												
Carpin Beach	0.03	0.04	0.04	0.03	0.13	0.16	0.18	0.16	0.02	0.02	0.02	0.02
Echo Bay	0.00	0.02	0.01	0.01	0.15	0.02	0.03	0.02	0.02	0.03	0.02	0.02
Lake George	0.03	0.02	0.01	0.02	0.05	0.06	0.10	0.14	0.04	0.03	0.02	0.02
Pumpkin Point	0.06	0.02	0.01	0.01	0.03	0.05	0.25	0.22	0.03	0.03	0.02	0.02
Maskinonge Bay	0.02	0.01	0.04	0.03	0.10	0.01	0.06	0.05	0.18	0.17	0.22	0.07
West Shore, St.	0.01	0.03	0.01	0.03	0.10	0.03	0.18	0.13	0.03	0.04	0.04	0.02
Joseph Island												
Non-AOC sites												
Anderson Creek	-	0.03	0.01	0.02	-	0.05	0.20	0.07	-	0.05	0.02	0.03
Desbarats Wetland	0.00	0.02	0.01	0.02	0.16	0.05	0.08	0.08	0.04	0.03	0.02	0.02
Stobie Creek	0.00	0.02	0.08	0.03	0.15	0.03	0.13	0.14	0.04	0.03	0.01	0.02
Hay Bay Wetland	0.08	0.02	0.02	0.02	0.05	0.12	0.24	0.20	0.02	0.03	0.02	0.02
Joe Dollar Bay	0.00	0.02	0.03	0.01	0.18	0.04	0.09	0.11	0.02	0.02	0.01	0.01
Wetland												

*2012 levels of NH₃-N and NO₃-N were the mean of replicate samples collected at each wetland.

Discussion

Based on the collected water quality data over the past four years, there is no apparent difference in coastal wetland AOC sites versus non-AOC sites: mean values ranged from moderately degraded to very good in the AOC and from moderately degraded to very good in non-AOC sites. Wetlands in the St. Marys River show a general improvement in WQI scores from 2012 to 2015. Exceptions include West Shore, St. Joseph Island, which experienced degradation in WQI from 2012-2014 but improvement in 2015, and Maskinonge Bay, which experienced relatively stable scores.

Sites with degraded water quality are typically wetlands with higher turbidity, which may be a result of natural processes (e.g., resuspension of fine sediments) or anthropogenic causes (e.g., inputs from the watershed) (Table 4). While turbidity values decreased in 2015 and were the lowest recorded values measured during all sampling years for Pumpkin Point, Lake George, and West Shore, St. Joseph Island, turbidity at these sites remained high when compared to other wetlands. Carpin Beach's turbidity values in 2015 were slightly elevated compared to values measured in previous years. Higher turbidity values at Lake George, Pumpkin Point, and West Shore, St. Joseph Island may be a result of wave action from large vessels resuspending fine mineral (e.g., clay, silt) substrates into the water column. The high turbidity at Hay Bay Wetland may be a result of impacts from the aggregate quarry located to the north of the wetland. The passage of large vessels in the river increases wave action and the resulting turbidity may negatively affect emergent wetlands (Kauss 1991). Other wetlands along the shipping channel such as Echo Bay, Carpin Beach and Maskinonge Bay have, to some degree, a level of protection from wave action from the passage of large vessels. For example, Echo Bay is protected from wave action as it has an opening of approximately 30 metres at the base of the wetland into the river.

Mean water temperature (Table 4) increased slightly in 2015 from 2014 values. In most cases, 2015 temperatures remained lower than 2013 values. This drop in water temperature, combined with a drop in pH, likely contributed to the improved WQI scores seen in 2014 and 2015. Water depth per station was on average 92 cm in 2015, 71 cm in 2014, and 46 cm in 2013; these increased depths may have influenced the colder water temperatures seen in 2014 and 2015. It should be noted that water depth is not measured at precisely the same points from year to year, but in most cases within the general vicinity of the same water station. Nonetheless, the increase in water depth was apparent at most sites, and combined with lower water temperatures, may have slowed SAV community growth, as discussed in Section 5.

Conductivity levels recorded in St. Marys River (average: 109 μ S/cm) in 2015 are below levels observed through surveys conducted by Environment and Climate Change Canada – Canadian Wildlife Service in other Great Lakes coastal wetlands. For example, surveys conducted as part of the Coastal Habitat Assessment and Monitoring Program in the Huron-Erie Corridor, which includes the Detroit River and St. Clair River AOCs, showed an average conductivity value of 325 μ S/cm in 2015 (Environment and Climate Change Canada, unpublished data). The average conductivity reading for St. Marys River AOC sites was 102 μ S/cm and 119 μ S/cm for non-AOC sites. This suggests that inputs from agricultural, industrial, and urban inputs in the St. Marys River are less than in other areas.

With the exception of Maskinonge Bay, total phosphorus levels were 0.03 mg/L or less. The Ministry of Environment and Energy (1999) presents an interim provincial water quality objective indicating that to avoid nuisance concentrations of algae in lakes, average total phosphorous concentrations should not exceed 0.02 mg/L. In 2015, nine of the eleven sites meet this objective, improving from only two of eleven sites meeting this objective in 2013.

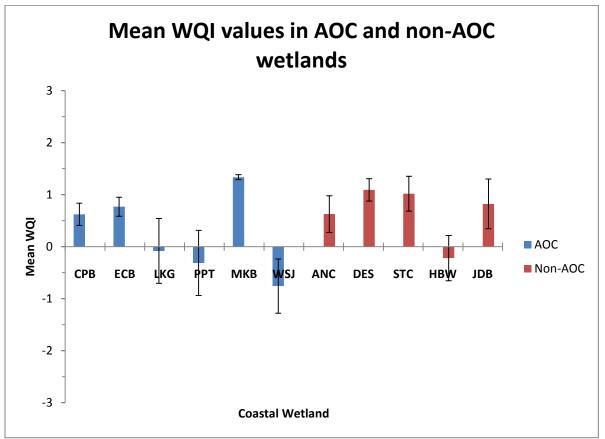


Figure 3: Mean WQI values for sampled wetlands during all years for comparisons between sites within the AOC and outside the AOC. CPB = Carpin Beach, ECB = Echo Bay, LKG = Lake George, PPT = Pumpkin Point, MKB = Maskinonge Bay, WSJ = West Shore, St. Joseph's Island, ANC = Anderson Creek, DES = Desbarats Wetlands, STC = Stobie Creek, HWB = Hay Bay Wetland, JDB = Joe Dollar Bay Wetland.

Figure 3 demonstrates the variation in mean WQI values over all sampled years, with wetlands both inside and outside the AOC demonstrating a range of values. In addition, the highest (best) scoring wetland, Maskinonge Bay, is within the AOC, and Hay Bay Wetland, which is outside of the AOC, scores below the majority of sites within the AOC. The WQI values of AOC sites showed no statistical difference from those of non-AOC sites, suggesting that values are comparable and overall water quality in the St. Marys River AOC can be considered not impaired with respect to criterion (i) of BUI #14.

6. Submerged Aquatic Vegetation Community

Methodology

The submerged aquatic vegetation (SAV) community was surveyed by sampling a one-metre square quadrat at 20 random locations in the open water basin of each wetland. Quadrat locations were randomly generated in a Geographic Information System (GIS) using ArcGIS 10.1 (ESRI 2012) prior to sampling. Within each quadrat, total areal coverage and species-specific coverages for submerged and floating-leaved species were recorded.

Submerged aquatic vegetation species were grouped into two plant guilds based on growing tolerance (i.e., turbidity tolerant and turbidity intolerant) and native designation (EC and CLOCA 2004; Grabas et al. 2012). Species were also assigned a coefficient of conservatism (Oldham et al. 1995): values range from 0 to 10 where higher scores are given to species having lower disturbance tolerance and greater fidelity to a certain habitat. Three metrics were shown to significantly respond to disturbance: SNAT (number of native species), CC (Coefficient of conservatism), and PCOV (total coverage).

Metrics were standardized into a range from 0 to 10 (EC and CLOCA 2004; Grabas et al. 2012). They were then added, multiplied by 10 and divided by the total number of metrics to create an IBI with scores between 0 and 100 with higher IBI scores representing better SAV community condition. EC and CLOCA (2004) identified five classes in which minimum detectable differences could be distinguished and associated with a qualitative descriptor (Table 7).

IBI Score	Qualitative Descriptor
81-100	Excellent
61-80	Very good
41-60	Good
21-40	Fair
0-20	Poor

 Table 7: Index of biotic integrity (IBI) score and associated category based on EC and CLOCA (2004).

Results

All surveyed wetlands in 2015 had one or more native species present (see Appendix 2 for full list of plant species observed; Tables 8 and 9). Pumpkin Point, West Shore, St. Joseph Island, and Hay Bay Wetland scored zero in total cumulative cover. Carpin Beach scored zero in the coefficient of conservatism standardized metric. The average of total cumulative coverage ranged from 2.9% (Hay Bay Wetland) to 66.2% (Maskinonge Bay).

In 2015, by percent cover and number of quadrats, Fern Pondweed (*Potamogeton robbinsii*), Tape Grass (*Vallisineria americana*), Stonewort (*Chara* sp.), Canada Waterweed (*Elodea canadensis*), and Richardson's Pondweed (*Potamogeton richardsonii*) were the five most common SAV species observed. Tape Grass, Canada Waterweed, Stonewort, and Richardson's Pondweed were the most ubiquitous species, found at ten of the eleven wetlands surveyed. Fern Pondweed and Tape Grass are turbidity intolerant, while Canada Waterweed is turbidity tolerant. All of the most common species are also native species. After experiencing a decline in IBI scores for the majority of sites in 2014, most wetlands remained stable or showed an increase in IBI score in 2015. Sites that had higher IBI scores in previous years (e.g., 80 or above) appeared to remain somewhat stable (e.g., Maskinonge Bay, Desbarats Wetland, Stobie Creek). Carpin Beach's IBI score was the lowest for all wetlands surveyed in 2015, while Stobie Creek's IBI score was the highest. Statistical comparisons (paired t-tests) of each SAV parameter were performed for AOC versus non-AOC wetlands for each sampling year, and then for the mean of all sampling years. The parameters of SNAT (number of native species) and FQI (floristic quality index, calculated by multiplying the mean coefficient of conservatism by the square root of native species per quadrat; Oldham et al., 1995) showed statistical differences in AOC versus non-AOC wetlands during 2013 and 2014, but not for other years or for the mean of all sampling years.

Discussion

The SAV IBI appeared to show differences between AOC and non-AOC sites in 2015 as well as in previous survey years. AOC sites ranged from poor to excellent (ranging in IBI scores from 5.5 to 88.9), with four of six sites ranked as fair or poor. West Shore, St. Joseph Island, Pumpkin Point, and Carpin Beach ranked in the poor category. Maskinonge Bay, however, ranked in the excellent category. Similarly, the non-AOC sites ranked from poor to excellent (ranging in IBI scores from 18.0 to 89.1), with one site ranked as poor and two of five sites ranked as excellent.

Increased water depth and lower water temperatures in 2015 likely affected the wetland plant community. Increased depth at some quadrats may have limited the visibility to determine percent coverage, including species such as Stonewort or Slender Naiad (*Naja flexilis*) which may cover the bottom of the substrate. Both species were among the most common species found in 2013, but only Stonewort was among the most common species found in 2015. Additionally, the wetland plant community may have experienced limited growth rates and abundance due to increased water depth. Zhu et al. (2012) studied the growth response of five SAV species to differing water depths, and found that increased water depth inhibited relative growth rate and biomass in all species, with Coontail (*Ceratophyllum demersum*) demonstrating the highest adaptive response to flooding. Furthermore, a decline in SAV at deep waters was suggested to be a result of root anchorage inhibition and subsequent facilitated uprooting in deep water conditions (Zhu et al. 2012).

Temperature has been shown to influence aquatic plant biomass, with 45-1160% more biomass produced in a warm year than a cool one (a difference of 1-6°C) in boreal lakes (Rooney and Kalff 2000). Bartleson et al. (2014) found Tape Grass had faster growing rates at high light and temperatures greater than 20°C, and slowest growth rates at 13°C, which may suggest a change in species dynamics due to cooler temperatures. Tape Grass was recorded in 2013 and 2015 surveys as one of the most common species, but was not one of the most common species in 2014 surveys during the lowest recorded temperatures, although it remained abundant.

The numerous and different factors influencing wetland plant community appear to be nuanced. It is therefore difficult to observe a pattern in solely four years of data, but

fluctuations are apparent in the St. Marys River, likely as a result of variability in water levels. Statistical differences were observed in two SAV parameters in two years (2013 and 2014), during the aforementioned change in water levels and temperatures. However, these differences were not observed across all years or for the mean of all years, indicating that the overall area is not impaired with respect to the SAV community, but nonetheless that the SAV IBI may have limited use in detecting change. Sites are affected both inside and outside the AOC, which emphasizes the importance of delisting criteria that accounts for natural variation.

	SNAT			CC			PCOV					
Wetland Name	2012	2013	2014	2015	2012	2013	2014	2015	2012	2013	2014	2015
AOC sites												
Carpin Beach	3.33	4.50	2.16	1.50	5.08	6.69	0.38	0.00	1.86	2.46	0.02	0.14
Echo Bay	10.0	6.33	3.83	10.0	8.03	7.94	6.31	8.96	9.22	6.00	1.32	2.38
Lake George	5.83	5.66	2.66	4.00	4.99	7.46	1.17	3.03	4.91	3.49	0.93	0.15
Pumpkin Point	5.33	4.16	1.17	2.16	7.60	3.79	0.00	1.06	3.08	2.02	0.00	0.00
Maskinonge Bay	10.0	10.0	10.0	10.0	9.10	9.67	10.0	10.0	8.95	9.50	8.79	6.67
West Shore, St.	2.33	2.16	0.83	2.00	1.46	2.17	0.00	0.25	0.54	0.00	0.00	0.00
Joseph Island												
Non-AOC sites												
Anderson Creek	-	10.0	6.83	10.0	-	8.77	8.19	10.0	-	4.71	1.10	1.19
Desbarats Wetland	10.0	10.0	10.0	10.0	9.04	10.0	9.27	9.69	10.0	10.0	6.19	4.62
Stobie Creek	8.99	10.0	10.0	10.0	8.55	9.46	8.84	9.63	7.64	9.94	7.06	7.09
Hay Bay Wetland	5.33	5.00	4.33	2.50	3.50	5.19	2.31	1.69	3.29	1.25	0.28	0.00
Joe Dollar Bay	6.49	9.82	4.33	5.66	4.35	7.38	4.96	4.68	4.91	6.28	4.39	3.45
Wetland												

Table 8: Submerged aquatic vegetation community standardized metrics (out of 10) for selected coastal wetlands in the St. Marys River Area of Concern (AOC) and non-AOC sites in the St. Marys River.

SNAT Number of native species

CC Coefficient of conservatism

PCOV Total cumulative coverage

Table 9: IBI scores (out of 100) and means for the condition of the submerged aquatic vegetation (SAV)
community for selected coastal wetlands in the St. Marys River Area of Concern (AOC) and non-AOC
sites in the St. Marys River.

			SAV	SAV IBI	
Wetland Name	2012	2013	2014	2015	Mean
AOC sites					
Carpin Beach	34.3	45.5	8.5	5.5	23.4
Echo Bay	90.8	67.6	38.2	71.1	66.9
Lake George	52.4	55.4	15.9	23.9	36.9
Pumpkin Point	53.4	33.2	3.9	10.8	25.3
Maskinonge Bay	93.5	97.3	96.0	88.9	93.9
West Shore, St. Joseph Island	14.4	14.4	2.8	7.5	9.8
Non-AOC sites					
Anderson Creek	-	78.3	53.7	70.6	67.5
Desbarats Wetland	96.8	100.0	84.9	81.0	90.7
Stobie Creek	83.9	98.0	86.3	89.1	89.3
Hay Bay Wetland	40.4	38.1	23.1	14.0	28.9
Joe Dollar Bay Wetland	52.5	78.3	45.6	46.0	55.6

7. Aquatic Macroinvertebrates

Methodology

For each wetland, three replicate sub-samples of approximately 150 nektonic and epiphytic aquatic macroinvertebrates (\geq 500 µm) were taken by sweep-netting through the water column in the emergent communities. Macroinvertebrates were identified to the lowest taxonomic group possible. While the target number of aquatic macroinvertebrates in each sample is 150, in some instances samples in previous years contained far fewer than 150 individuals.

A full list of aquatic macroinvertebrates can be found in Appendix 3.

Results

Although Cvetkovic and Midwood (2015a) developed an invertebrate IBI based on the WQI, it was later determined that none of the candidate metrics were suitable for creating an IBI, and also that there were no single metrics that could be used to assess the aquatic macroinvertebrate condition (Cvetkovic and Midwood 2015c).

Discussion

The difficulty in developing an aquatic macroinvertebrate IBI reiterates the fact that relatively little disturbance occurs in the St. Marys River. Without a large continuum of disturbance, the biotic communities, both inside and outside the AOC, are similarly in relatively undisturbed condition. Note that this does not imply that any IBIs developed would not able to differentiate the degraded sites from those in better condition; rather, there appears to be little difference in condition among sites, and consequently that severe degradation is no longer occurring in this AOC. This similarity in condition suggests that the aquatic macroinvertebrate biotic community can be considered no longer impaired.

8. Breeding Bird Community

Methodology

Breeding marsh bird communities were surveyed using a modification to the Marsh Monitoring Program (MMP) protocol (Meyer et al. 2006) to report on site-level or specific AOC wetland bird communities. The primary purpose of the MMP is to assess population trends of common marsh bird species across broad geographic scales and/or long timeframes. Bird survey stations were identified using aerial photographs and set up at least 250 metres apart. Only those that had at least 50% marsh habitat (i.e., non-woody emergent plants) within the sampling radius (100 m) were surveyed. Marsh bird surveys were conducted using a 15 minute point count as follows: five minutes of passive surveying followed by five minutes of call broadcasting for secretive species (e.g., Yellow Rail (*Coturnicops noveboracensis*), Least Bittern (*Ixobrychus exilis*), Sora (*Porzana carolina*), Virginia Rail (*Rallus limicola*), Common Moorhen (*Gallinula chloropus*) / American Coot (*Fulica americana*), and Pied-billed Grebe (*Podilymbus podiceps*)) followed by five minutes of passive surveying. The protocol was modified slightly so that only marsh birds were recorded rather than all birds.

Results

All 11 wetlands were surveyed for birds in 2015. Echo Bay and Lake George were the only sites that had area-sensitive marsh-nesting obligates present during the surveys. Specifically, Lake George had Black Tern (*Chlidonias niger*) and American Bittern (*Botaurus lentiginosus*) present, while Echo Bay had Black Tern present. A full list of species recorded during the survey within 100 metres of the station can be found in Appendix 3.

Discussion

Area sensitive marsh nesting obligates such as Black Tern and American Bittern are species that are known to prefer larger wetlands and are less likely to be found in smaller wetland sites. For example, MMP data suggests that Black Terns require permanent marshes of at least 50 hectares in size to reproduce successfully (McCracken, no date). Of the wetlands surveyed, the following are over 50 hectares: Echo Bay (587 ha), Lake George (155 ha), Maskinonge Bay (71.5 ha), Desbarats Wetland (89.7 ha), and Hay Bay Wetland (158.5 ha) although only a small portion (12 ha) of the wetland is accessible for surveys. To date, area sensitive marsh nesting obligate species have only been in Echo Bay and Lake George. Smaller wetlands may not be large enough to consistently support populations of area-sensitive marsh nesting obligates, and in some cases, wetlands over the threshold limit still may not be able to support these species for various other reasons (e.g., ratio of vegetation to open water). Water level changes are also likely to impact the available suitable habitat within the wetlands.

Although Cvetkovic and Midwood (2015a) previously developed potential marsh breeding bird IBIs in the St. Marys River AOC, validation with 2015 data proved unsuccessful wherein the developed IBIs responded poorly to the disturbance gradient. Thus, the potential IBIs were considered unsuitable to determine sensitivity of the wetland sites to disturbance. Again, the lack of sensitivity to disturbance suggests that sites are in relatively undisturbed condition. As

sites inside and outside the AOC are in comparable condition, the marsh bird biotic community can be considered not impaired.

9. Amphibians

Methodology

Surveys for amphibians (frogs and toads) were conducted following the Marsh Monitoring Program (Bird Studies Canada 2000). Amphibian survey stations are separated by at least 500 metres and visited at night on three separate surveys. Each amphibian station is surveyed for three minutes and one of three Call Level Codes is used to categorize the intensity of calling activity for each species.

Amphibian community condition may be determined using an IBI. However, at present no IBI has been developed for St. Marys River coastal wetlands. Cvetkovic and Midwood (2015a, 2015b) determined that none of the nine metrics tested were potential candidates to develop a suitable IBI for this region. Although a potential metric was identified in previous work (Cvetkovic and Midwood 2014), an additional year of data rendered the potential metric unsuitable for IBI development. Furthermore, there were no clear relationships between amphibian metrics and disturbance gradients.

Results

Eleven wetlands were surveyed for amphibians in 2015, while ten (excluding Anderson Creek) were surveyed in 2013-2014. As described above, no amphibian IBI was developed due to insufficient response of metrics to disturbance.

Using combined 2013-2015 amphibian data, Green Frog (*Rana clamitans melanota*) and Spring Peeper (*Pseudacris crucifer*) were present in every site, followed by Wood Frog (*Lithobates sylvaticus*), Gray Treefrog (*Hyla versicolor*), American Toad (*Anaxyrus americanus*), and Northern Leopard Frog (*Lithobates pipiens*) at ten of eleven sites. Mink Frog (*Lithobates septentrionalis*) was recorded in eight sites and Bullfrog (*Lithobates catesbeianus*) was only recorded once, in Desbarats in 2013. Desbarats had the highest mean species richness (2013-15) of all sites, recording all eight species, while Carpin Beach and Stobie Creek had the fewest, recording only five species. Species richness for AOC wetlands ranged from five to seven species and six to eight for non-AOC wetlands. A full list of species recorded during 2015 surveys within 100 metres of the station can be found in Appendix 4.

Discussion

The continued difficulty in creating an amphibian IBI suggests that the disturbance gradients for the region are not sufficient to detect strong responses from the amphibian community attributes. Nonetheless, without a sharp delineation between sites inside and outside the AOC with respect to the disturbance gradients used here, the amphibian community in the St. Marys River may be considered not impaired.

In an additional caveat, the amphibian community metrics were based on expected (within species range) species at each wetland. The list of expected species was set the same for all surveyed wetlands to aid in IBI analysis given the relatively small geographic range and limited

number of other amphibian surveys in the area. Although necessary for analysis, the standardization of expected species across all sites may have diminished any subtle species or guild differences found among wetlands, therefore reducing the ability to develop an IBI that accurately detects changes and responses in the amphibian community.

At this time, with no clear response of the amphibian community to different disturbance gradients, it is recommended that the St. Marys River be considered not impaired for this biotic community.

10. Summary

Water Quality and Biotic Community

Based on data collected in the four surveyed years, there is no clear pattern in water quality or the biotic community for AOC and non-AOC sites. Both areas show a range of condition for water quality and biotic communities. Statistical comparisons showed no statistical difference between wetlands located in the AOC versus outside the AOC for any water quality parameter for any year, suggesting that the water quality in the St. Marys River is comparable and therefore not impaired. While the SAV parameters of SNAT and FQI showed statistical differences were not observed across all years, indicating that the overall area is not impaired with respect to the SAV community.

Development of IBIs

An IBI was developed for the SAV community, but IBIs could not be developed for marsh bird, aquatic macroinvertebrate, and amphibian communities in the St. Marys River. Small sampling sizes combined with the lack of a clear gradient of disturbance likely contribute to the difficulty in developing appropriate IBIs.

Water temperatures dropped drastically in the St. Marys River during 2014 when compared to 2012 and 2013, and generally remained low in 2015. Lower conductivity and lower pH were also observed. Cvetkovic and Midwood (2015a) suggest that interannual variability may be greater than among-site variability, and therefore grouping data by wetland (as done in this report) may mask changes in water quality better observed when grouped by year if differences are mitigated by the multi-year data when averaged by wetland.

The increase in water levels observed in the St. Marys River from 2012 to 2013, 2013 to 2014, and 2014 to 2015 may have had significant impacts on the biotic communities measured here. Water levels in Lake Michigan-Huron increased approximately 1.0 metre from January 2013 to December 2014 (Gronewold et al. 2015). Comparing data gathered in the St. Marys River from a single year may be more effective at analyzing change than combining wetland data from several years, which may mask yearly effects. Further, amphibian and bird metric responses were found to be weaker during low water levels in IBI development in Lake Ontario and Lake Erie, such that broad-scale effects were more apparent during low water levels (Crewe and Timmermans 2005).

Wetlands were sampled in the St. Marys River from 2012 to 2015, totalling 43 site years. Despite this, development of an amphibian IBI has consistently proved unsuccessful, and other biotic communities generally showed weak to no response to disturbance. Validation of potential IBIs developed in 2014 proved successful with only the SAV community; potential IBIs for the marsh bird and invertebrate community did not show a strong response to disturbance, which rendered them unsuitable for reporting on community condition. Accordingly, IBIs cannot be used in the St. Marys River to assess wetland condition in most biotic communities.

The exception of the SAV IBI in responding to disturbance and its ability to detect change in wetland condition suggests that this IBI is tied strongly to the geophysical parameters of the WQI, although it represents only a portion of overall wetland condition. Therefore, using IBIs to assess overall wetland condition in the St. Marys River remains unsuitable, resulting in a strong indication that the area is not impaired with respect to the marsh breeding bird, aquatic macroinvertebrate, and amphibian biotic communities.

The narrow range of WQI values and overall high water quality in the area, combined with the fact that IBIs cannot be developed for several biotic communities, suggest that the range of disturbance in the St. Marys River is no longer strong enough to warrant a corresponding response in the biota. Therefore, with sites condition inside and outside the AOC being comparable, it is recommended that the St. Marys River be considered no longer impaired for criterion (i) of BUI #14: *Loss of Fish and Wildlife Habitat* as well as coastal wetland bird and amphibian (frogs and toads) populations in BUI #3: *Loss of Fish and Wildlife Populations*.

Literature Cited

- Bartleson, R.D., M.J. Hunt, and P.H. Doering. Effects of temperature on growth of *Vallisneria americana* in a sub-tropical estuarine environment. Wetlands Ecol Manage (2014) 22:571–583.
- Bird Studies Canada. 2000. The Marsh Monitoring Program Quality Assurance Project Plan. Prepared for the U.S. Environmental Protection Agency, Great Lakes National Program Office. West Jackson Boulevard, Chicago, Illinois.
- Chow-Fraser, P. 2006. Development of the Wetland Water Quality Index (WQI) to assess effects of basin-wide land-use alteration on coastal marshes of the Laurentian Great Lakes. In: *"Coastal Wetlands of the Laurentian Great Lakes: Health, Habitat and Indicators"* Eds. Simon, T.P. and Stewart, P.M. Authorhouse, Bloomington, Indiana.
- Crewe, T.L. and S.T.A. Timmermans. 2005. Assessing Biological Integrity of Great Lakes Coastal Wetlands Using Marsh Bird and Amphibian Communities Project # WETLAND3-EPA-01 Technical Report. Bird Studies Canada, Port Rowan, Ontario. 88 pp.
- Cvetkovic, M. and J. Midwood. 2014. Developing a Site Disturbance Gradient and Coastal Wetland Indices of Biotic Integrity for St. Marys River AOC. Freshwater Inquiry Network. Prepared for Environment Canada, Canadian Wildlife Service – Ontario. Toronto, Ontario, Canada.
- Cvetkovic, M. and J. Midwood. 2015a. Refining a Site Disturbance Gradient and Coastal Wetland Indices of Biotic Integrity for St. Marys River AOC. Freshwater Inquiry Network. Prepared for Environment Canada, Canadian Wildlife Service (Ontario).
- Cvetkovic, M. and J. Midwood. 2015b. Coastal wetland indicators in St. Marys River Area of Concern: Addendum to Midwood and Cvetkovic (2015) Report. Environment Canada, Canadian Wildlife Service – Ontario. Toronto, Ontario, Canada.
- Cvetkovic, M. and J. Midwood. 2015c. St Marys River Macroinvertebrate IBI Exploration. Environment Canada, Canadian Wildlife Service – Ontario. Toronto, Ontario, Canada.
- Environment Canada. 2013. St. Marys River Area of Concern: Coastal Wetland Habitat Scoping Report. Environment Canada, Canadian Wildlife Service (Ontario).
- Environment Canada. 2014. St. Marys River Area of Concern: Coastal Wetland Habitat Assessment Report. Environment Canada, Canadian Wildlife Service (Ontario).
- Environment Canada. 2015. St. Marys River Area of Concern: Coastal Wetland Habitat Assessment Report. Environment Canada, Canadian Wildlife Service (Ontario).

Environment Canada and Central Lake Ontario Conservation Authority. 2004. Durham Region Coastal Wetland Monitoring Project: Year 2 Technical Report. Downsview, ON: ECB-OR.

Environment Canada and Central Lake Ontario Conservation Authority. 2007. Durham Coastal Wetland Monitoring Project: Methodology Handbook. Environment Canada, Canadian Wildlife Service (Ontario). <u>http://www.cloca.com/lwc/monitoring_coastalact.php</u>.

Environmental Systems Research Institute (ESRI). 2012. ArcGIS 10.1. Redlands, California.

- Grabas G.P., E.A. Blukacz-Richards, and S. Pernanen. 2012. Development of a submerged aquatic vegetation community index of biotic integrity for use in Lake Ontario coastal wetlands. *Journal of Great Lakes Research*. 30(2):243-250.
- Grabas, G.P., T.L. Crewe and S.T.A. Timmermans. 2008. Bird Community Indicators. Chapter 7 *In* Great Lakes Coastal Wetlands Monitoring Plan, pp.116-142. Great Lakes Wetlands Consortium, March 2008. <u>www.glc.org/wetlands/documents/finalreport/ch7.pdf</u>
- Gronewold, A. D., A. H. Clites, J. Bruxer, K. W. Kompoltowicz, J. P. Smith, T. S. Hunter, and C. Wong (2015), Water levels surge on Great Lakes, Eos, 96, doi:10.1029/2015EO026023. Published on 17 March 2015.
- Kauss, P.B. 1991. Biota of the St. Marys River: habitat evaluation and environmental assessment. *Hydrobiologia*. 219: 1-35.
- McCraken, J. 'Troubled Waters: What is Happening to Marsh Bird Populations?' Bird Studies Canada. Accessed 23 April 2015. <<u>http://www.bsc-eoc.org/blterns.html</u>>
- Meyer, S.W., J.W. Ingram and G.P. Grabas. 2006. The Marsh Monitoring Program: evaluating marsh bird survey protocol modifications to assess Lake Ontario coastal wetlands at a site-level. Technical Report Series 465. Canadian Wildlife Service, Ontario Region, Ontario.
- Ministry of Environment and Energy. 1999. Water Management Policies Guidelines Provincial Water Quality Objectives of the Ministry of Environment and Energy. Ministry of Environment and Energy. 32 pp.
- Oldham, M. J., W. D. Bakowsky and D. A. Sutherland. 1995. Floristic Quality Assessment System for Southern Ontario. Ontario Ministry of Natural Resources, Natural Heritage Information Centre: Peterborough. 69 pp.
- Rooney, N., and Kalff, J. 2000. Inter-annual variation in submerged macrophyte community biomass and distribution: the influence of temperature and lake morphometry. Aquat. Bot. 68: 321–335.

- St. Marys River Binational Public Advisory Council. 2015. Stage 2 Remedial Action Plan Implementation Annex for the Canadian Waters of the St. Marys River Area of Concern.
- Uzarski, D.G., Brady, V.J., and M.C. Cooper. 2014. GLIC: Implementation Great Lakes Coastal Wetland Monitoring. U.S. EPA. 96 pp.
- Zhu, G., W. Li, M. Zhang, L. Ni, and S. Wang. 2012. Adaptation of submerged macrophytes to both water depth and flood intensity as revealed by their mechanical resistance. Hydrobiologia 696: 77-93.

Appendices

Appendix 1: List of taxa recorded on submerged aquatic vegetation surveys for 2015 showing nativeness, turbidity tolerance ($\sqrt{}$ = tolerant, X = intolerant) and coefficient of conservatism (for vascular species).

Common Name	Genus/Species	Native	Turbidity- Tolerant	Coefficient of Conservatism
Algae	Algae sp. (fil. underwater)	V		
Algae	Algae sp. (fil. surface)	V		
Bladderwort	<i>Utricularia</i> sp.	V		
Brittlewort	Nitella sp.	V		
Broad-leaved Arrowhead	Sagittaria latifolia	V		4
Canada Waterweed	Elodea canadensis	V	v	4
Claspingleaf pondweed	Potamogeton perfoliatus	V		7
Common Bladderwort	Utricularia vulgaris	V		4
Common burreed	Sparganium eurycarpum	V		3
Common Three-square	Schoenoplectus pungens	V		6
Coontail, Hornwort	Ceratophyllum demersum	V	v	4
Eurasian Water Milfoil	Myriophyllum spicatum		V	
Fern Pondweed	Potamogeton robbinsii	V	Х	7
Flat-leaved Bladderwort	Utricularia intermedia	V		
Flat-stemmed Pondweed	Potamogeton zosteriformis	V	Х	5
Floating-leaved Burreed	Sparganium fluctuans	V		9
Floating-leaved Pondweed	Potamogeton natans	V		5
Greater Duckweed	Spirodela polyrhiza	V		4
Hardstem Bulrush	Schoenoplectus acutus	V		6
Lakebank sedge	Carex lacustris	V		5
Leafy Pondweed	Potamogeton foliosus	V	V	4
Marsh St. John's Wort	Triadenum virginicum	V		
Narrow-leaved Cattail	Typha angustifolia	V		3
Needle Spikerush	Eleocharis acicularis	V		
Northern Water Milfoil	Myriophyllum sibiricum	V	Х	6
Pickerelweed	Pontederia cordata	V		7
Purple Loosestrife	Lythrum salicaria			
Pygmy Water Lily	Nymphaea tetragona	V		
Richardson's, Clasping Leaved Pondweed	Potamogeton richardsonii	V		5
Seven-angled pipewort	Eriocaulon aquaticum	V		9
Slender Naiad	Najas flexilis	V	Х	5
Slender Pondweed	Potamogeton pusillus	V	v	5
Softstem Bulrush	Schoenoplectus tabernaemontani	٧		5
Spike-Rush	Eleocharis smallii	V		6
Star Duckweed	Lemna trisulca	V		4
Stonewort, Muskgrass	Chara sp.	V		
Tape Grass, Water Celery	Vallisneria americana	V	Х	6
Threeway sedge	Dulichium arundinaceum	V		7
Variable-leaved Pondweed	Potamogeton gramineus	V		4
Vasey's pondweed	Potamogeton vaseyi	V		8

Common Name	Genus/Species	Native	Turbidity- Tolerant	Coefficient of Conservatism
Water horsetail	Equisetum fluviatile	V		7
Watershield	Brasenia schreberi	V		7
Water Smartweed	Polygonum amphibium	V		5
Water Star-grass	Heteranthera dubia	V	v	7
Water-Marigold	Megalodonta beckii	V	Х	8
White Water Lily, Fragrant Water Lily	Nymphaea odorata	V		5
Whorl-leaf Water Milfoil, Bracted Water Milfoil	Myriophyllum verticillatum	V		7
Wild Rice	Zizania palustris	V		9
Yellow Pond Lily, Bullhead Lily	Nuphar lutea ssp. variegata	V		4

Appendix 2: List of aquatic macroinvertebrate species identified to the lowest taxonomic unit possible from 2015 samples.

Phylum	Class	Order	Family	Genus/Species
Annelida	Clitellata	Arhynchobdellida	Erpobdellidae	
		,	·	Alboglossiphonia
Annelida	Clitellata	Rhynchobdellida	Glossiphoniidae	heteroclita
		,	·	Glossiphonia
Annelida	Clitellata	Rhynchobdellida	Glossiphoniidae	complanata
Annelida	Clitellata	Rhynchobdellida	Glossiphoniidae	Helobdella sp.
Annelida	Clitellata	Rhynchobdellida	Glossiphoniidae	Helobdella stagnalis
Annelida	Clitellata	Rhynchobdellida	Glossiphoniidae	Helobdella triserialis
Annelida	Oligochaeta			
Arthropoda	Arachnida	Hydracarina		
Arthropoda	Crustacea	Amphipoda		
Arthropoda	Crustacea	Amphipoda	Gammaridae	Gammarus fasciatus
Arthropoda	Crustacea	Amphipoda	Gammaridae	Gammarus sp.
Arthropoda	Crustacea	Amphipoda	Hyalellidae	Hyalella azteca
Arthropoda	Crustacea	Decapoda	Cambaridae	Orconectes sp.
Arthropoda	Crustacea	Isopoda	Asellidae	Caecidotea sp.
Arthropoda	Crustacea	Isopoda	Asellidae	Lirceus sp.
Arthropoda	Insecta	Coleoptera	Gyrinidae	Dineutus sp.
Arthropoda	Insecta	Coleoptera	Gyrinidae	Gyrinus sp.
Arthropoda	Insecta	Coleoptera	Haliplidae	Haliplus sp.
Arthropoda	Insecta	Coleoptera	Haliplidae	Peltodytes sp.
Arthropoda	Insecta	Coleoptera	Hydrophilidae	Tropisternus sp.
Arthropoda	Insecta	Collembola		
Arthropoda	Insecta	Diptera	Ceratopogonidae	
Arthropoda	Insecta	Diptera	Ceratopogonidae	Bezzia/Palpomyia
Arthropoda	Insecta	Diptera	Chironomidae	
Arthropoda	Insecta	Diptera	Chironomidae	Tanypodinae sp.
Arthropoda	Insecta	Diptera	Sciomyzidae	
Arthropoda	Insecta	Ephemeroptera	Baetidae	
Arthropoda	Insecta	Ephemeroptera	Baetidae	Callibaetis sp.
Arthropoda	Insecta	Ephemeroptera	Baetidae	Cloeon dipterum
Arthropoda	Insecta	Ephemeroptera	Baetidae	Procloeon/Centroptilum
Arthropoda	Insecta	Ephemeroptera	Caenidae	Caenis sp.
Arthropoda	Insecta	Ephemeroptera	Ephemerellidae	
Arthropoda	Insecta	Ephemeroptera	Ephemerellidae	Eurylophella sp.
Arthropoda	Insecta	Hemiptera	Belostomatidae	
Arthropoda	Insecta	Hemiptera	Corixidae	
Arthropoda	Insecta	Hemiptera	Corixidae	Hesperocorixa sp.
Arthropoda	Insecta	Hemiptera	Corixidae	Palmacorixa sp.
Arthropoda	Insecta	Hemiptera	Corixidae	Sigara sp.
Arthropoda	Insecta	Hemiptera	Corixidae	Trichocorixa sp.
Arthropoda	Insecta	Hemiptera	Gerridae	
Arthropoda	Insecta	Hemiptera	Gerridae	Gerris sp.
Arthropoda	Insecta	Hemiptera	Mesoveliidae	Mesovelia sp.
Arthropoda	Insecta	Hemiptera	Nepidae	Ranatra sp.
Arthropoda	Insecta	Hemiptera	Notonectidae	Notonecta sp.
Arthropoda	Insecta	Hemiptera	Pleidae Granski da s	Neoplea sp.
Arthropoda	Insecta	Lepidoptera	Crambidae	Acentria sp.

Phylum	Class	Order	Family	Genus/Species
Arthropoda	Insecta	Lepidoptera	Crambidae	Elophila sp.
Arthropoda	Insecta	Megaloptera	Sialidae	Sialis sp.
Arthropoda	Insecta	Odonata	Aeshnidae	
Arthropoda	Insecta	Odonata	Aeshnidae	Aeshna sp.
Arthropoda	Insecta	Odonata	Aeshnidae	Anax sp.
Arthropoda	Insecta	Odonata	Aeshnidae	Basiaeshna sp.
Arthropoda	Insecta	Odonata	Calopterygidae	Calopteryx sp.
Arthropoda	Insecta	Odonata	Coenagrionidae	
Arthropoda	Insecta	Odonata	Coenagrionidae	Enallagma sp.
				Enallagma/Coenagrion
Arthropoda	Insecta	Odonata	Coenagrionidae	sp.
Arthropoda	Insecta	Odonata	Coenagrionidae	lschnura sp.
Arthropoda	Insecta	Odonata	Corduliidae	
Arthropoda	Insecta	Odonata	Corduliidae	Epitheca (Tetragoneuria)
Arthropoda	Insecta	Odonata	Gomphidae	Stylurus sp.
Arthropoda	Insecta	Odonata	Lestidae	Lestes sp.
Arthropoda	Insecta	Odonata	Libellulidae	Leucorrhinia sp.
Arthropoda	Insecta	Odonata	Libellulidae	Leucorrhinia/Sympetrum
Arthropoda	Insecta	Odonata	Libellulidae/Corduliidae	
Arthropoda	Insecta	Trichoptera	Helicopsychidae	Helicopsyche sp.
Arthropoda	Insecta	Trichoptera	Hydroptilidae	Agraylea sp.
Arthropoda	Insecta	Trichoptera	Hydroptilidae	Oxyethira sp.
Arthropoda	Insecta	Trichoptera	Leptoceridae	
Arthropoda	Insecta	Trichoptera	Leptoceridae	Ceraclea sp.
Arthropoda	Insecta	Trichoptera	Leptoceridae	Mystacides sp.
Arthropoda	Insecta	Trichoptera	Leptoceridae	Oecetis sp.
Arthropoda	Insecta	Trichoptera	Leptoceridae	Triaenodes sp.
Arthropoda	Insecta	Trichoptera	Limnephilidae	Anabolia sp.
Arthropoda	Insecta	Trichoptera	Limnephilidae	Limnephilus sp.
Mollusca	Bivalvia	Veneroida	Pisidiidae	
Mollusca	Bivalvia	Veneroida	Pisidiidae	Musculium sp.
Mollusca	Gastropoda	Basommatophora	Ancylidae	
Mollusca	Gastropoda	Basommatophora	Ancylidae	Ferrissia sp.
Mollusca	Gastropoda	Basommatophora	Lymnaeidae	
Mollusca	Gastropoda	Basommatophora	Lymnaeidae	Lymnaea stagnalis
Mollusca	Gastropoda	Basommatophora	Lymnaeidae	Pseudosuccinea sp.
Mollusca	Gastropoda	Basommatophora	Lymnaeidae	Stagnicola sp.
Mollusca	Gastropoda	Basommatophora	Physidae	Physa/Physella
Mollusca	Gastropoda	Basommatophora	Planorbidae	
Mollusca	Gastropoda	Basommatophora	Planorbidae	Gyraulus sp.
Mollusca	Gastropoda	Basommatophora	Planorbidae	Helisoma/Planorbella
Mollusca	Gastropoda	Basommatophora	Planorbidae	Promenetus sp.
Mollusca	Gastropoda	Mesogastropoda	Valvatidae	Valvata sp.
Mollusca	Gastropoda	Mesogastropoda	Valvatidae	Valvata tricarinata
	Gastropoda	Neotaenioglossa	Hydrobiidae	
Mollusca	Gastropoua	Neotaemogiossa	nyaroonaac	

Appendix 3: List of species recorded within 100 m on marsh breeding bird surveys in 2015.

Species Common Name	Scientific Name	Marsh User	Forager	
Alder Flycatcher	Empidonax alnorum	NA	AF	
American Bittern	Botaurus lentiginosus	AEMNO	WF	
American Black Duck	Anas rubripes	MNG	WF	
American Crow	Corvus brachyrhynchos	NA	NAF	
American Goldfinch	Carduelis tristis	NA	NAF	
American Redstart	Setophaga ruticilla	NA	NAF	
American Robin	Turdus migratorius	NA	NAF	
American Wigeon	Anas americana	NA	WF	
Bald Eagle	Haliaeetus leucocephalus	NA	AF	
Bank Swallow	Riparia riparia	NA	AF	
Barn Swallow	Hirundo rustica	NA	AF	
Belted Kingfisher	Ceryle alcyon	NA	AF	
Black Tern	Chlidonias niger	AEMNO	AF	
Black-capped Chickadee	Parus atricapillus	NA	NAF	
Blue Jay	Cyanocitta cristata	NA	NAF	
Blue-winged Teal	Anas discors	NA	WF	
Canada Goose	Branta canadensis	MNG	WF	
Caspian Tern	Sterna caspia	NA	AF	
Cedar Waxwing	Bombycilla cedrorum	NA	NAF	
Common Goldeneye	Bucephala clangula	NA	WF	
Common Grackle	Quiscalus quiscula	NA	NAF	
Common Loon	Gavia immer	MNG	WF	
Common Merganser	Mergus merganser	NA	WF	
Common Moorhen	Gallinula chloropus	EMNO	WF	
Common Raven	Corvus corax	NA	NAF	
Common Tern	Sterna hirundo	NA	AF	
Common Yellowthroat	Geothlypis trichas	MNG	NAF	
Double-crested Cormorant	Phalacrocorax auritus	NA	WF	
European Starling	Sturnus vulgaris	NA	NAF	
Gadwall	Anas strepera	NA	WF	
Great Blue Heron	Ardea herodias	NA	WF	
Green-winged Teal	Anas crecca	NA	WF	
Hairy Woodpecker	Picoides villosus	NA	NAF	
Hermit Thrush	Catharus guttatus	NA	NAF	
Herring Gull	Larus argentatus	NA	AF	
Hooded Merganser	Lophodytes cucullatus	NA	WF	
Killdeer	Charadrius vociferus	NA	NAF	
Le Conte's Sparrow	Ammodramus leconteii	MNG	NAF	
Mallard	Anas platyrhynchos	NA	WF	
Marsh Wren	Cistothorus palustris	EMNO	NAF	
Merlin	Falco columbarius	NA	AF	
Northern Flicker	Colaptes auratus	NA	NAF	
Northern Rough-winged Swallow	Stelgidopteryx serripennis	NA	AF	
Northern Shoveler	Anas clypeata	NA	WF	
Northern Waterthrush	Seiurus noveboracensis	NA	NAF	
Osprey	Pandion haliaetus	NA	AF	
Pied-billed Grebe	Podilymbus podiceps	EMNO	WF	
Pine Warbler	Dendroica pinus	NA	NAF	
Red-winged Blackbird	Agelaius phoeniceus	MNG	NAF	

Species Common Name	Scientific Name	Marsh User	Forager
Ring-billed Gull	Larus delawarensis	NA	AF
Sandhill Crane	Grus canadensis	MNG	WF
Song Sparrow	Melospiza melodia	NA	NAF
Sora	Porzana carolina	EMNO	NAF
Swamp Sparrow	Melospiza georgiana	MNO	NAF
Tree Swallow	Tachycineta bicolor	NA	AF
Turkey Vulture	Cathartes aura	NA	AF
Virginia Rail	Rallus limicola	EMNO	NAF
Wilson's Snipe	Capella gallinago delicata	MNG	NAF
Wood Duck	Aix sponsa	NA	WF
Yellow Warbler	Dendroica petechia	NA	NAF
AEN	INO Area Sensitive Emergent Ma	rsh Nesting Obligat	e
AN	INO Area Sensitive Marsh Nestin	g Obligate	
EN	1NO Emergent Marsh Nesting Ob	ligate	
Ν	1NG Marsh Nesting Generalist		
N	1NO Marsh Nesting Obligate		
	AF Aerial Forager		
	NAF Non-Aerial Forager		
	WF Water Forager		

Appendix 4: List of species recorded within 100 m on amphibian surveys in 2015.

Species Common Name	Scientific Name
American Toad	Anaxyrus americanus
Gray Treefrog	Hyla versicolor
Green Frog	Rana clamitans melanota
Mink Frog	Lithobates septentrionalis
Northern Leopard Frog	Lithobates pipiens
Spring Peeper	Pseudacris crucifer
Wood Frog	Lithobates sylvaticus

Appendix 3b: An Assessment of Breeding Populations of Common Terns and Black Terns in the St. Marys River Area of Concern, Ontario (Hughes et al. 2014)

An Assessment of Breeding Populations of **Common Terns and Black Terns in the** St. Marys River Area of Concern (Ontario)



Environment Canada K.D. Hughes, D.J. Moore, and P.A. Martin October 2014



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Purpose

The purpose of this summary is to report on the results of recent assessments of breeding populations of Common Terns (*Sterna hirundo*) and Black Terns (*Chlidonias niger*) within the St. Marys River Area of Concern (AOC) (Ontario) following the recommended action put forward in the Stage 2 Remedial Action Plan report (Environment Canada *et al.* 2002). These assessments are based on nest count surveys conducted in the AOC and are supplemented with historical breeding data for these species where available. Population trends for other colonial waterbirds based on decadal surveys are also reported to provide a broader historical context of trends in diversity and abundance of breeding colonial waterbirds in the St. Marys River AOC.

Methods

Since the 1970s, Canadian Wildlife Service (CWS) biologists have conducted four decadal surveys of colonial waterbirds nesting on the Canadian side of the Great Lakes. In the St. Marys River (Ontario), these surveys were completed in 1978/80 (1st survey), 1989 (2nd survey), 1999/2000 (3rd survey) and with the most recent decadal survey (i.e., 4th survey) in 2007/2008 (Blokpoel and Tessier 1996, 1997, 1998; Morris *et al.* 2003, 2010, 2011, 2012; Rush *et al.* in review; CWS unpublished). During these surveys, all nests of colonial waterbirds were counted individually using the total count methods of Blokpoel and Tessier (1996). The results of decadal surveys of colonial waterbirds in the North Channel of Lake Huron are also reported to provide a comparison of trends downstream and beyond the influence of the AOC. For the purpose of reporting on colonial waterbird trends, the North Channel is defined as the area bounded by the contiguous shoreline from McNab Reef in St. Joseph Lake, which is northeast of Campement d'Ours Island, to Frazer Bay, located southeast of McGregor Point on the mainland. The southern boundary of the North Channel includes Manitoulin Island, Cockburn Island and the international boundary.

In addition to the decadal surveys conducted within the AOC, additional surveys were conducted from 2010-2013 as part of two separate Environment Canada (EC) studies of Common Terns in the region. Nest counts from these studies are included to examine more recent trends in abundance in the AOC. One of these studies was conducted in 2011 and 2012 to assess the reproductive status of Common Terns and Herring Gulls breeding within the St. Marys River AOC (Hughes *et al.* 2014). Noteworthy is that in 2011 the only breeding Common Tern colony in the AOC was at an island NNW of Hay Point near the southwestern shoreline of St. Joseph Island. This colony was abandoned early in the breeding season and subsequently a colony located at North Sister Rock, just beyond the AOC boundary in St. Joseph Lake, was selected as the alternate AOC colony. For consistency purposes in this report, this nesting site as well as four other small tern nesting sites (i.e., McNab Reef, Hurt Rock, islet 200m SW of mouth of Sucker Creek, and South Sister Rock) are considered to be part of the AOC. This cluster of nesting sites, consisting of small rocks, are in close proximity to one another and are situated approximately 6 kilometres beyond the eastern AOC boundary at Quebec Bay. Surveys of these Common Tern nesting sites as well as other sites downstream were conducted from 2010-2013 as part of another EC study of breeding site tenacity and productivity of Common Terns on the North Channel (D.J. Moore, CWS

unpublished). Similar to that reported above, temporal trends of Common Tern populations in the St. Marys River AOC will be compared to trends reported for tern populations in the North Channel.

As part of the CWS decadal survey of Great Lakes coastal marshes conducted in 2010, marsh habitat within a 5 kilometre band along the St. Marys River shoreline was surveyed for Black Terns, as well as other marsh-nesting waterbirds. Nest counts were determined by either counting individual nests or by halving the maximum number of adult terns counted during a series of sweeps with binoculars and/or a spotting scope. Historical breeding data for Black Terns in the AOC are provided where available. These include CWS decadal surveys of coastal marshes (Graham *et al.* 2002) as well as evidence of breeding status based on data collected for the first and second Ontario Breeding Bird Atlases in the St. Marys River (Cadman *et al.* 1987, 2007).

Results and Discussion

Historical Context – Colonial Waterbirds in the St. Marys River AOC (Ontario)

As part of the 4th decadal CWS Great Lakes colonial waterbird survey, four species including the Herring Gull (*Larus argentatus*), Common Tern, Ring-billed Gull (*Larus delawarensis*) and the Great Blue Heron (*Ardea herodias*) were found nesting on the St. Marys River (Ontario) (CWS unpublished; Table 1a). In total, 882 nests (=breeding pairs) were found at eight nesting sites situated on natural habitat that consisted of islands, reefs, shoals, and the mainland (Figure 1). Herring Gulls were the most abundant species nesting at six sites with colonies ranging in size from 42 nests at one of two islands south of Pumpkin Point to 159 nests at the island west of Chene Island in the northern portion of the AOC. Ring-billed Gulls were the next most abundant species and were found nesting at the shoal northeast of Whitestone Reef (262 nests) and the island west of Chene Island (83 nests). Common Terns were found nesting at two sites with colonies on the island NNW of Hay Point near the southwestern shoreline of St. Joseph Island (68 nests) and a nearby shoal west of Hay Point (10 nests). These two Common Tern nesting sites were not visited in 2008 as part of the survey for nesting terns in the St. Marys River; however, they were visited in 2010 and counts at these two sites are considered part of the 4th decadal survey. A single Great Blue Heron nest was found on Whitestone Reef in 2008.

Over the four CWS Great Lakes colonial waterbird decadal surveys on the St. Marys River, numbers of Herring Gull nests have remained high at over 450 nests (Blokpoel and Tessier 1996, 1997; Morris *et al.* 2003; CWS unpublished; Table 1a). Nest numbers decreased by -36.0% between the 3rd and 4th decadal surveys from 716 nests in 1999/2000 to 458 nests in 2007/2008. Ring-billed Gull nests which were relatively fewer in number during the first three decadal surveys (ranging from 0 nests to 14 nests) increased by two orders of magnitude from two nests found in the 3rd decadal survey to 345 nests in the 4th decadal survey (Blokpoel and Tessier 1996, 1997; Morris *et al.* 2011; CWS unpublished). Common Terns nested at two sites, Whitestone Reef and the island NNW of Hay Point, in the AOC in the 1st decadal survey (70 nests) and the 2nd decadal survey (159 nests; Blokpoel and Tessier 1996, 1997). While no nesting of Common Terns was reported in the AOC during the 3rd decadal survey in 1999, numbers of Common Tern nests increased in the 4th decadal survey (78 nests; CWS unpublished). The apparent abandonment of nesting sites by Common Terns in the AOC during the 3rd decadal survey is likely related to the low site fidelity shown by this species (see below), particularly evident in an area where

only three nesting sites have been historically reported. Great Blue Heron nests were found in low numbers at two different sites in two of the four decadal surveys: at an island south of Pumpkin Point in the 2nd decadal survey (3 nests) and at Whitestone Reef in the 4th survey in 2008 (1 nest; Blokpoel and Tessier 1998; CWS unpublished). Overall, the total number of colonial waterbird nests on the St. Marys River (Ontario) increased by +22.8% from 718 nests in 1999/2000 to 882 nests in 2007/2008, a finding largely driven by the dramatic increase in nesting Ring-billed Gulls in the AOC.

Table 1. Census data of colonial waterbird nests (=breeding pairs) on the Canadian side of the St. Marys River (a) during the 1st (1978/80), 2nd (1989), 3rd (1999/2000) and 4th (2007/08) decadal surveys as part of the Great Lakes colonial waterbird surveys conducted by the Canadian Wildlife Service (Blokpoel and Tessier 1996, 1997, 1998; Morris *et al.* 2003, 2010, 2011, 2012; CWS unpublished). Census data for colonial waterbirds on the North Channel of Lake Huron during the four decadal surveys are shown in (b). Percent change in nest numbers are based on comparisons between the 3rd and 4th surveys; NC denotes that rate cannot be calculated.

Species		Percent			
Species	1978/80	1989	1999/2000	2007/08*	Change
Herring Gull	539	612	716	458	-36.0%
Ring-billed Gull	0	14	2	345	+17150%
Common Tern	70	159	0	78	NC
Great Blue Heron	0	3	0	1	NC
Totals	609	788	718	882	+22.8%

a) St. Marys River (Ontario):

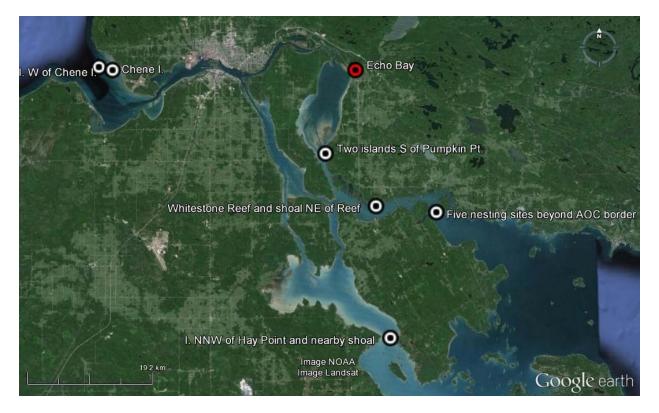
* Includes census data for two Common Tern nesting sites that were surveyed in 2010.

b)	North	Channel:
~,	1.01.01	enannen

Species		Percent			
	1980	1989/91	1997-2001	2008/09	Change
Herring Gull	5,551	3,706	4,291	3,377	-21.3%
Common Tern	2,680	1,360	1,954	1,924	-1.5%
Ring-billed Gull	53,645	48,405	35,117	23,898	-31.9%
Great Blue Heron	206	82	108	43	-60.2%
Caspian Terns	547	603	338	377	+11.5%
Double-crested Cormorant	159	2,177	6,716	7,380	+9.9%
Black-crowned Night Heron	0	0	9	48	+433%
Great Black-backed Gull	0	0	1	0	-100%
Totals	62,788	56,333	48,534	37,047	-23.7%

Population trends of colonial waterbirds breeding in the North Channel of Lake Huron during the four decadal surveys are provided in Table 1b as a comparison to trends downstream of the St. Marys River AOC (Blokpoel and Tessier 1996, 1997, 1998; Morris *et al.* 2003, 2010, 2011, 2012, Morris *et al.* in press; Rush *et al.* in review; CWS unpublished). Seven species were reported breeding in the North Channel during the 4th decadal survey in 2008/09. In addition to the four species breeding within the AOC, Caspian Tern (*Hydroprogne caspia*), Double-crested Cormorant (*Phalacrocorax auritus*), and Black-crowned Night-Heron (*Nycticorax nycticorax*) were also found nesting. This region has historically

Figure 1. Locations of nesting sites for colonial waterbirds (white circles) and Black Terns (red circle) within the St. Marys River Area of Concern (Ontario) during surveys conducted in 2007/08/10. Also shown is the general location of the five Common Tern nesting sites (McNab Reef, Hurt Rock, Islet SW of mouth of Sucker Creek, North Sister Rock and South Sister Rock) situated just beyond the AOC boundary and considered within the AOC for the assessment of Common Tern populations in this report.



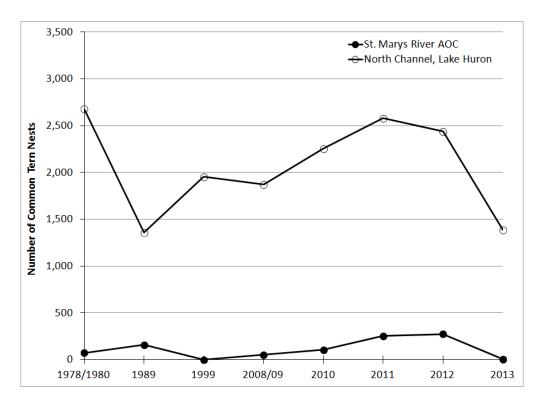
supported a rich diversity of colonially nesting waterbirds owing to its large expansive area consisting of numerous islands, reefs and rocks ideal as nesting habitat with the surroundings waters providing an abundant food supply. Over 37,000 nests were counted at 129 nesting sites during this survey period. Between the 3rd and 4th decadal surveys, increases in nest numbers were found for Caspian Terns (+11.5%), Double-crested Cormorants (+9.9%), and Black-crowned Night-Heron (+433%) in the North Channel. Similar to trends in the AOC, a decrease in nest numbers was found for Herring Gulls (-21.3%) while, in contrast to the AOC, decreases in nest numbers of Common Terns (-1.5%), Ring-billed Gulls (-31.9%), and Great Blue Heron (-60.2%) were found in the North Channel between the two last surveys. While a single Great Black-backed Gull (*Larus marinus*) nest was found in 1999, no nesting was reported by this species in this region in 2009. In the North Channel, a steady decrease in numbers of breeding colonial waterbirds has been found since the first decadal survey with an overall decrease in nest numbers of colonial waterbirds by -23.7% between the 3rd and 4th decadal surveys.

Trends in Breeding Populations of Common Terns

Based on the results of the four CWS Great Lakes decadal surveys and supplemented with data from recent surveys conducted by EC from 2010-2013, the total nesting population of Common Terns in the St. Marys River AOC generally ranged in size from a total of 53 nests in 2008 to 273 nests in 2012 (Figure

2). These counts include the additional five small nesting sites situated just beyond the AOC boundary as cited in the Methods (Figure 1). In two years, 1999 and 2013, nest numbers in the AOC were notably low with 1 and 3 nests, respectively, at single nesting sites. While a total of eight different nesting sites were occupied in the AOC during these eight surveys, numbers of nesting sites in each survey year ranged from one to five. The largest nesting colonies and corresponding years reported were: island NNW of Hay Point (1980, 1989, 2010: range=39-129 nests), McNab Reef (1999: 1 nest, 2008: 48 nests), South Sister Rock (2011: 109 nests), North Sister Rock (2012: 129 nests), and Hurt Rock (2013: 3 nests). Note that nest counts for two sites (i.e., island NNW of Hay Point and the nearby shoal west of Hay Point) in 2010 are grouped with census data for other colonies surveyed in that year. As a result, the total nest count may be underestimated in 2008/09 since these two colonies were not visited in that period. A complete listing of the eight nesting sites and corresponding nest numbers for the eight survey periods is provided in the Appendix.

Figure 2. Total nest counts of Common Terns in the St. Marys River AOC (Ontario) and the North Channel of Lake Huron for the four decadal surveys, 1978/80, 1989, 1999, and 2008/09 and annual surveys conducted from 2010-2013 as part of two complementary EC studies of Common Terns in the region. Note that the first four time-points on the x-axis are not evenly spaced time periods.



Breeding populations of Common Terns downstream in the North Channel were consistently higher than breeding populations in the St. Marys River AOC in every survey year (Figure 2). Colony sizes were larger and nesting sites were more numerous with total numbers of nesting sites ranging from 13 sites in 2010 to 28 sites in 1980. The largest nesting colonies and corresponding years reported were: island NE of Thessalon Dock (1980: 661 nests), island SW of Henry Island (1989, 2008, 2013: range=307-534 nests),

Batture Island (1999: 478 nests) and E island of the Cousins Islands (2010, 2011, 2012: range=525-1,142 nests). Nest numbers in this region were the highest in 1980 (2,677 nests), decreased in 1989 (1,360 nests), slowly increased reaching a peak in 2011 (2,582 nests) and then decreased in 2012 (2,440 nests) and 2013 (1,386 nests). While general declines in breeding populations of Common Terns have been reported throughout the Canadian Great Lakes over the four decadal surveys (Morris *et al.* 2012), declines in nest numbers were not as pronounced at colonies on Lake Huron (including the North Channel, as shown here) as those found on the lower Great Lakes.

Despite differences in the relative sizes of the breeding populations in the St. Marys River AOC and North Channel, there are some similarities in the patterns of abundance and site occupancy of Common Terns between the two regions which contributed to inter-year fluctuations in nest numbers observed. Specifically, nesting sites were not consistently occupied from year to year and there appeared to be considerable turnover in nesting sites between years. In the AOC, five of eight nesting sites (63%) were occupied in only one or two years of eight survey years and the remaining three sites were occupied in either three (South Sister Rock), four (North Sister Rock) or five (island NNW of Hay Point) survey years. This pattern of low site fidelity was effectively demonstrated in a broad EC 2010-2012 study of Common Terns in the North Channel where comparisons of nesting sites showed considerable turnover between consecutive study years: 14-59% of sites were abandoned, 9-55% of sites were newly colonized and 31-42% of sites remained active (D.J. Moore, CWS unpublished). These findings were further supported by the longer term decadal survey data for Common Terns in the North Channel (D.J. Moore, CWS unpublished). Morris *et al.* (2012) found similar evidence of shifts in site occupancy with high rates of abandonment and high rates of colonization of new sites at tern colonies on the Canadian Great Lakes (based on the decadal survey), Newfoundland and New Brunswick.

These results highlight that low site fidelity in Common Terns is an important life history trait which must be considered in interpreting trends in abundance of Common Terns both at the local level (i.e., nesting colony) and the larger regional area. Inter-year fluctuations in nest numbers would be more pronounced in regions such as the AOC where Common Terns breed in relatively low numbers and at few nesting sites. In addition, the nearby proximity of the North Channel as well as the U.S. side of the St. Marys River with ample suitable habitat may facilitate selection of these regions as alternate nesting sites. This strategy may be beneficial for reproductive success under conditions of stress such as when predators are present. In 2011, significant predation of tern eggs early in the breeding season at the Hay Point colony in the EC Common Tern reproduction study was suspected to have resulted in abandonment of the colony which led to the subsequent selection of the next nearest colony for study at North Sister Rock. Predation was also severe at some North Channel nesting sites from 2010-2013 (CWS unpublished). Other general stressors known to influence productivity of Common Terns include adverse weather and water level fluctuations which were responsible for egg and chick losses at some North Channel nesting sites in 2010-2013 (CWS unpublished). Periods of high water levels in the northern Great Lakes have also been associated with loss of Common Tern nesting sites which would also impact population trends (Schugart and Scharf 1983). Contaminant levels in eggs of Common Terns collected from AOC colonies in 2011 and 2012 were not sufficiently high to impact reproductive success and development (Hughes et al. 2014). Based on the evidence from nest count surveys conducted from

1980 to 2013, trends in populations of nesting Common Terns in the AOC are likely related to factors consistent with the life history strategies of the species and that are not specific to influences in the AOC.

Trends in Breeding Populations of Black Terns

Based on the results of the CWS survey of marsh-nesting terns conducted in 2010, a large colony of Black Terns (88 nests) was found at Echo Bay, a large coastal wetland on Lake George (Figure 1). No other breeding colonies of Black Terns were found at coastal marshes along the St. Marys River (Ontario) shoreline. Additional evidence of Black Terns breeding at Echo Bay was also found in 2012 (point count surveys; Environment Canada 2013) and in 2013 when approximately 20 breeding pairs (nests) were counted (T. Hoar, pers. comm.).

Table 2 provides a summary of available data reporting breeding status of Black Terns at sites within the AOC, just beyond the AOC border and on the North Channel shown for comparisons purposes. During the CWS decadal survey conducted in 2001, one Black Tern colony (16 nests) was found on St. Joseph Island (Graham *et al.* 2002). This survey did not include Echo Bay or Pumpkin Point. No nests were found in surveys of sites beyond the AOC border and the North Channel in 2001 and 2010 although only a portion of the North Channel was surveyed in 2010. Overall it is not possible to report on temporal trends in abundance of Black Terns in the AOC since only two CWS surveys were conducted and some sites were not surveyed in 1991, surveys did not include areas north of McGregor Bay near Little Current, Lake Huron and thus did not include the St. Marys River (Austen *et al.* 1996). Using data collected for the two Breeding Bird Atlases, nesting was confirmed at sites within the AOC during the periods 1981-1985 and 2001-2005 (Cadman *et al.* 1987, 2007). Based on the very limited data reported here, there is no evidence to suggest that breeding status for Black Terns nesting within the AOC differs from those nesting at sites downstream in the North Channel.

The Black Tern is a species of Special Concern in Ontario. Long term trends of Black Terns based on data collected by the Marsh Monitoring Program from 1995 to 2007 indicate a significant Great Lakes basin decline for Black Terns at a rate of -11.4% per year (Archer and Jones 2009). These trends are supported by Breeding Bird Survey data from 1970-2012 that showed population declines for Black Terns in Ontario at a rate of -5.54% per year and regionally in the Boreal Hardwood Transition Zone (Bird Conservation Region 12), which includes regions along the St. Marys River and North Channel shorelines, at a rate of -6.43% per year (Environment Canada 2014). As reviewed by Burke (2012), Black Tern populations in Ontario have disappeared from many traditional nesting sites and have declined at existing sites, particularly in southern Ontario where they were historically most abundant. The destruction and/or degradation of wetlands are among the most important factors contributing to population declines of this species (Dunn and Agro 1995). Habitat quality is also important as Black Terns prefer to nest in a hemi-marsh, i.e., a wetland with 50:50 open water and emergent vegetation. The extensive areas of large coastal wetlands on the St. Marys River provide nesting habitat for Black Terns compared to the sites further downstream in the North Channel which may be less suitable although confirmed breeding was reported on Manitoulin Island (Table 2; Cadman *et al.* 2007). Other

Table 2. Summary of available data reporting breeding status of Black Terns at sites within the St. Marys River AOC, beyond the AOC boundary and in the North Channel from 1981 to 2010. Shown are results of nest counts for surveys conducted by CWS and breeding evidence from the Ontario Breeding Bird Atlas (OBBA) with survey years/periods as indicated. "NS" denotes that the site was not surveyed.

Regions	OBBA1 square	OBBA2 square	Site Name	CWS Survey [#] 2001	CWS Survey 2010	OBBA1 [^] 1981-85	0BBA2 [^] 2001-05
St. Marys R. AOC	16GG25	16GS25	Echo Bay	NS	88	Possible	Possible
St. Marys R. AOC	16GG24	16GS24	Pumpkin Point	NS	0	Confirmed	Confirmed
St. Marys R. AOC	16GG21	16GS21	St. Joseph I. (Everens Pt./Sunset Pt.)	16	0	Possible	Confirmed
St. Marys R. AOC	16GG22	16GS22	St. Joseph I. (Reed Pt./Court Pt.)	0	0	Confirmed	Not Breeding
St. Marys R. AOC	16GG31	16GS31	St. Joseph I. (Outlook/Hay Point)	0	0	Not Breeding	Not Breeding
Beyond AOC boundary	17KB70	17KM70	St. Joseph I. (South A-D)	0	0	Probable	Not Breeding
Beyond AOC boundary	17KB81	17KM81	St. Joseph I. (Milford Haven)	NS	0	Confirmed	Probable
North Channel, Lake Huron	17KB73	17KM73	Desbarats	0	0	Confirmed	Probable
North Channel, Lake Huron	17LB91	17LM91	Spanish Marsh	0	NS	Confirmed	Not Breeding
North Channel, Lake Huron	17MA38	17ML38	Manitoulin Island	0	NS	Possible	Confirmed

CWS decadal survey of Great Lakes coastal marshes (2001) conducted by Bird Studies Canada (Graham et al. 2002)

^ Breeding evidence for the two Ontario Breeding Bird Atlases (Cadman *et al.* 1987, 2007) is categorized as confirmed, probable, possible, or not breeding.

potential stressors include decreased fluctuations in water levels (which can impact the extent of emergent vegetation that is likely more an issue in the lower Great Lakes), increasing numbers of invasive species (that can affect wetland structure) and climate change (Burke 2012). Differences in survey methodology and regions surveyed and few data collection years combined with large-scale population declines make an assessment of trends within the AOC difficult. However, based on the most recent CWS survey of coastal wetlands along the St. Marys River (Ontario) in 2010, the low population size of nesting Black Terns within the AOC is likely reflective of low densities reported throughout region, Great Lakes and Ontario.

Summary

Recent surveys of Common Terns and Black Terns by CWS on the St. Marys River (Ontario) indicate that these two colonial waterbird species are breeding within the AOC. Common Terns have nested consistently at sites situated both within and just beyond the official AOC boundary in eight survey years. While large inter-year fluctuations in nest numbers were evident at these nesting sites, these patterns are likely related to factors consistent with the life history strategies of this species that are not specific to influences within the AOC. A colony of Black Terns has consistently been reported at Echo Bay, a large coastal wetland, over the last few years. Based on the limited data available, there is no evidence to suggest that breeding status for Black Terns nesting within the AOC differs from that outside of AOC. In addition, the low population size of Black Terns is reflective of low density throughout Ontario rather than conditions specific to the AOC. In general, population declines have been reported for both of these species in the Great Lakes basin. During the most recent CWS decadal survey, four colonial waterbird species were found on the St. Marys River including a new species, the Great Blue Heron, which was not found nesting on this side of the River in the previous survey. Overall, the total number of colonial waterbird nests on the St. Marys River (Ontario) increased by +22.8% from 718 nests in 1999/2000 to 882 nests in 2007/08, a finding largely driven by the dramatic increase in nesting Ringbilled Gulls in the AOC.

References

Archer, R.W. and K.E. Jones. 2009. *The Marsh Monitoring Program Annual Report, 1995-2007. Annual indices and trends in bird abundance and amphibian occurrence in the Great Lakes basin.* Prepared for Environment Canada-Canadian Wildlife Service. Bird Studies Canada. 39 pp.

Austen, M.J., H. Blokpoel, and G.D. Tessier. 1996. Atlas of colonial waterbirds nesting on the Canadian Great Lakes, 1989-1991. Part 4. Marsh-nesting terns on Lake Huron and the lower Great Lakes system in 1991. Canadian Wildlife Service Technical Report Series Number 217. Canadian Wildlife Service, Environmental Conservation Branch, Ontario Region.

Blokpoel, H. and G.D. Tessier. 1996. *Atlas of colonial waterbirds nesting on the Canadian Great Lakes, 1989-1991. Part 1. Cormorants, gulls and island-nesting terns on Lake Superior in 1989.* Canadian Wildlife Service Technical Report Series No. 181. Canadian Wildlife Service, Environmental Conservation Branch, Ontario Region. 104 pp.

Blokpoel, H. and G.D. Tessier. 1997. *Atlas of colonial waterbirds nesting on the Canadian Great Lakes, 1989-1991. Part 2. Cormorants, gulls and island-nesting terns on Lake Huron in 1989*. Canadian Wildlife Service Technical Report Series No. 259. Canadian Wildlife Service, Environmental Conservation Branch, Ontario Region. 154 pp.

Blokpoel, H. and G.D. Tessier. 1998. *Atlas of colonial waterbirds nesting on the Canadian Great Lakes, 1989-1991. Part 5. Herons and egrets in 1991.* Canadian Wildlife Service Technical Report Series No. 272. Canadian Wildlife Service, Environmental Conservation Branch, Ontario Region. 36 pp.

Burke, P.S. 2012. *Management plan for the Black Tern (Chlidonias niger) in Ontario*. Ontario Management Plan Series. Prepared for the Ontario Ministry of Natural Resources (OMNR). Peterborough, Ontario. vi +47 pp.

Cadman, M.D., P.F.J. Eagles, and F.M. Helleiner. 1987. *Atlas of the breeding birds of Ontario*. Federation of Ontario Naturalists and the Long Point Bird Observatory, University of Waterloo Press. 617 pp.

Cadman, M.D., D.A. Sutherland, G.G. Beck, D. Lepage, and A.R. Couturier. 2007. *Atlas of the breeding birds of Ontario, 2001-2005*. Bird Studies Canada, Environment Canada, Ontario Field Ornithologists, Ontario Ministry of Natural Resources, and Ontario Nature. Toronto. xxii + 706 pp.

Dunn, E.H. and D.J. Agro. 1995. Black Tern (*Chlidonias niger*). In *The Birds of North America*, No. 147. (A Poole and F. Gill, eds). The Academy of Natural Sciences, Philadelphia, and The American Ornithologists' Union, Washington D.C.

Environment Canada, United States Environmental Protection Agency, Ontario Ministry of the Environment, and Michigan Department of Environmental Quality. 2002. *The St. Marys River Area of Concern remedial strategies for ecosystem restoration.* 140 pp.

Environment Canada. 2013. *St. Marys River Area of Concern: coastal wetland habitat scoping report*. Environment Canada, Canadian Wildlife Service. March 2013. 35 pp.

Environment Canada. 2014. North American Breeding Bird Survey-Canadian trends website: <u>http://www.ec.gc.ca/ron-bbs</u>. Data-version 2012. Environment Canada, Gatineau, Quebec, K1A 0H3.

Graham, D., S. Timmermans, and J. McCracken. 2002. *A comparison of abundance of colonial marsh birds between 1991 and 2001 in the Canadian portions of Lakes Huron, St. Clair, Ontario and Erie*. Report submitted to Environment Canada, Environmental Conservation Branch, Ontario Region. August 2002.

Hughes, K.D., D. Crump, K. Williams, and P.A. Martin. 2014. *Assessment of the wildlife reproduction and deformities beneficial use impairment in the St. Marys River Area of Concern (Ontario).* February 2014. Environment Canada. 32 pp.

Morris, R.D., D.V. Weseloh, and J.L. Shutt. 2003. Distribution and abundance of nesting pairs of Herring Gulls (*Larus argentatus*) on the North American Great Lakes, 1976 to 2000. *J. Great Lakes Res.* 29(3): 400-426.

Morris, R.D., D.V. Weseloh, F.J. Cuthbert, C. Pekarik, L.R. Wires, and L. Harper. 2010. Distribution and abundance of nesting common and Caspian terns on the North American Great Lakes, 1976 to 1999. *J. Great Lakes Res.* 36: 44-56.

Morris, R.D., D.V. Weseloh, L.R. Wires, C. Pekarik, F.J. Cuthbert, and D.J. Moore. 2011. Population trends of Ring-billed Gulls breeding on the North American Great Lakes, 1976 to 2009. *Waterbirds* 34(2): 202-212.

Morris, R.D., C. Pekarik, and D.J. Moore. 2012. Current status and abundance trends of Common Terns breeding at known coastal and inland nesting regions of Canada. *Waterbirds* 35(2): 194-207.

Shugart, G.W. and W.C. Scharf. 1983. Common Terns in the northern Great Lakes: current status and population trends. *J. Field Ornithol.* 54(2): 160-169.

Appendix. Nest counts of Common Terns at three sites within the boundary of the St. Marys River AOC (Ontario) as designated by the 41K site ids and the five nesting sites just beyond the AOC border as designated by the 41J site ids. Counts provided are from the four decadal surveys conducted in 1980, 1989, 1999 and 2008 and annual surveys conducted from 2010-2013 as part of two separate EC studies of Common Terns is the region. "-" indicates a site not previously identified and "NS" denotes that the site was not surveyed.

SiteID	Site Name	1978/80	1989	1999	2008	2010*	2011	2012	2013
	Census	1	2	3	4				
41K027	Whitestone Reef	31	30	0	0	NS	0	0	NS
41K028	Island NNW of Hay Point	39	129	0	NS	68	18	34	0
41K028A	Nearby shoal west of Hay Point	-	-	-	NS	10	0	0	0
41J001	McNab Reef	0	0	1	48	0	0	0	0
41J002	Hurt Rock	0	0	0	0	0	57	0	3
41J002A	Islet 200m SW of mouth of Sucker Creek	-	-	-	0	0	3	0	0
41J004	North Sister Rock	3	0	0	5	0	68	129	0
41J004A	South Sister Rock	-	-	-	0	27	109	110	0
	Total Number of Nests	73	159	1	53	105	255	273	3
	Total Number of Colonies	3	2	1	2	3	5	3	1

* Nest counts for all sites surveyed in 2010 are grouped together and include counts for the two nest sites 41K028 and 41K028A that are considered part of the official 4th decadal CWS survey completed in 2007/08.

Appendix 3c: Assessment of the Wildlife Reproduction and Deformities Beneficial Use Impairment in the St. Marys River Area of Concern, Ontario (Hughes et al. 2014)





Assessment of the Wildlife Reproduction and Deformities Beneficial Use Impairment in the St. Marys River Area of Concern (Ontario)



Environment Canada K.D. Hughes, D. Crump, K. Williams, and P.A. Martin February 2014



Environment Environnement Canada Canada



ABSTRACT

Reproduction and development were examined in herring gulls (Larus argentatus) and common terns (Sterna hirundo) breeding within the St. Marys River Area of Concern (Ontario) in 2011 and 2012. Freshly-laid eggs were collected from colonies within the Area of Concern (AOC) as well as outside of the AOC, artificially incubated in the laboratory and assessed for embryonic viability, incidence of embryonic deformities, contaminant burdens and other biochemical endpoints. Productivity was determined at the colonies when chicks were > 21 days old and chicks were examined for morphological deformities as well as other biological endpoints. Overall, embryonic viability of herring gulls and common terns was high at AOC colonies and herring gull productivity at AOC colonies was within the range required to maintain a stable population. Common tern productivity at AOC colonies, while low, was consistent with rates for common terns within the region and was largely attributable to external stressors, such as predation and severe weather events. No morphological deformities were found in field surveys of juveniles of either species (based on sample sizes of 13-63 chicks); however, a low incidence of embryonic deformities (i.e., 4-8% based on sample sizes of 15-30 embryos) was observed for both species collected from AOC colonies and artificially incubated in the lab. The significance of this finding is unknown at this time. Comparable burdens of non-ortho PCBs, 2,3,7,8-TCDD, and TEQs between deformed and normal embryos from AOC colonies of both species rule out the possibility that embryonic deformities were associated with elevated exposure to these compounds. Importantly, contaminant burdens (e.g., PCBs, 2,3,7,8-TCDD, and mercury) in gull and tern embryos from the St. Marys River AOC (Ontario) were comparable and not notably elevated compared to burdens at respective reference colonies in the two study years. Finally, concentrations of PCBs, other organochlorines, PBDEs, dioxins/furans and mercury were not sufficiently elevated in embryos to adversely impact the reproductive success and development of herring gulls and common terns foraging in the St. Marys River AOC.

INTRODUCTION

The St. Marys River is approximately 112 kilometres in length and is an important and major waterway in the Great Lakes interconnecting Lake Superior and the North Channel of Lake Huron. The St. Marys River Area of Concern (AOC) is one of 43 Great Lakes AOCs which were initially identified by Canada, the United States and the International Joint Commission (IJC) as specific locations where local environmental degradation had severely impacted the area's ability to support aquatic life. Historical discharges of pollutants from local steel and pulp and paper industries, a tannery and manufactured gas plant, and municipal storm sewers and wastewater treatment plants impaired water quality and contaminated sediment along parts of the St. Marys River (OMOE and MDNR 1992). Contaminants of concern included PAHs, mercury and other heavy metals, and polychlorinated biphenyls (PCBs) which contributed to exceedences of water quality objectives, sediment quality guidelines, fish consumption guidelines and impacted biota (OMOE and MDNR 1992; EC *et al.* 2002). The St. Marys River, as a connecting channel, is one of five Great Lakes AOCs jointly shared by Canada and the United States. As directed by Annex 2 of the 1987 Protocol to the Canada-U.S. Great Lakes Water Quality Agreement (GLWQA), a Remedial Action Plan (RAP) for the St. Marys River was developed collaboratively by Canadian and U.S. partners to address environmental concerns that are specific to the Ontario and

Michigan portions of the river, respectively. Implementation of the remedial actions continues. The border of the Canadian portion of the St. Marys River AOC (Ontario) extends from its head at Gros Cap in Whitefish Bay downstream to St. Joseph Island via Lake George to Quebec Bay in the St. Joseph Channel and downstream to Hay Point on the western shore of St. Joseph Island (Figure 1).

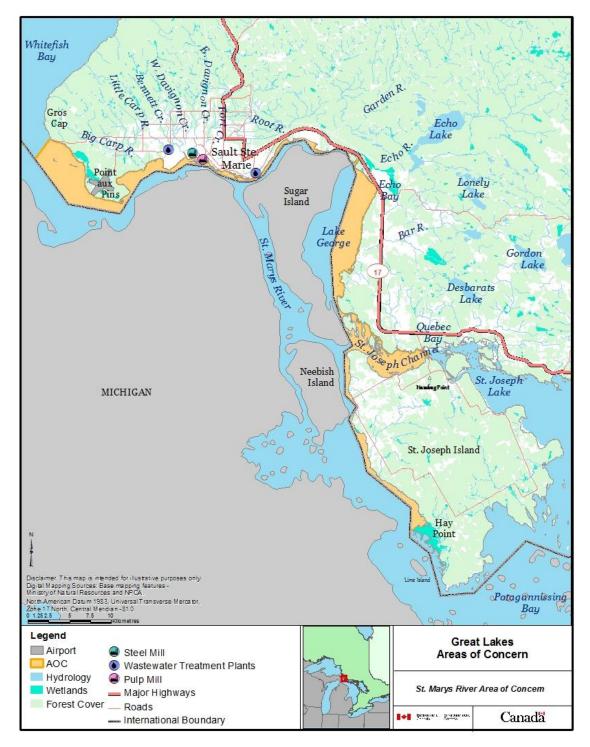


Figure 1. The St. Marys River Area of Concern as defined by the boundary in Ontario.

Fourteen beneficial use impairments (BUIs), caused by a detrimental change in the chemical, physical or biological integrity of the Great Lakes system, were used by the Canadian and U.S. federal governments to identify AOCs and then as a framework for directing remediation efforts. One of these BUIs, "bird or animal deformities or reproduction problems", relates to contaminant exposure or other anthropogenic environmental stressors on reproductive success or deformity rates in wildlife. Based on reports from the 1980s, there were no bird or animal deformities or reproductive problems in the St. Marys River AOC (with limited contaminants data available for wildlife) and this BUI was not impaired (OMOE and MDNR 1992). However, the status of the BUI in the AOC was subsequently changed to "requires further assessment" in the 1990s/early 2000s with the recommendation that reproductive assessments of herring gulls (*Larus argentatus*) and common terns (*Sterna hirundo*) be completed within the AOC (EC *et al.* 2002). It was recommended that deformities be assessed in common terns following evidence of deformities found in three common tern chicks - cross bills - at a colony on Lime Island, Michigan, in 1998. Following this, studies of the potential effects of contaminants on reproduction and development in aquatic-feeding wildlife were initiated by federal, state and provincial agencies to more fully evaluate and assess the current status of this BUI in the St. Marys River AOC.

Fish-eating wildlife, such as colonial waterbirds, are important indicators of exposure to persistent contaminants in the aquatic environment (Fox and Weseloh 1987). As top predators, they occupy a high trophic level in the aquatic food web and therefore can accumulate high levels of contaminants which may in turn adversely affect their reproductive health and development. Two colonial waterbird species which breed and forage within the St. Marys River AOC were selected for assessment purposes. The herring gull is a long-lived, primarily fish-eating colonial waterbird that from the time it reaches breeding age is a year-round resident in the Great Lakes basin. The common tern, as an obligate piscivore, is an excellent bio-indicator for tracking potential problems relating to PCB contamination since they are very sensitive to the dioxin-like effects of certain PCBs (Nisbet 2002). In 2011 and 2012, breeding colonies of these two species were studied by Environment Canada (EC) in the St. Marys River AOC (Ontario). Eggs were collected for artificial incubation in the laboratory to assess embryonic viability, incidence of embryonic deformities, contaminant burdens and biochemical endpoints (i.e., fatty acids and stable isotopes). Under controlled laboratory conditions, this method assesses the effects of embryonic exposure to potentially high levels of contaminants during critical periods of development. Reproduction and development were also assessed in wild populations with visits to colonies to monitor productivity and examine chicks for morphological deformities as well as measure additional endpoints (i.e., stress hormone, thyroxine levels) relating to growth and development which could be influenced by increased contaminant exposure. In combination with the results of extensive reproduction and development studies conducted in Michigan's AOCs (MDEQ 2012), the results of this study will be used to assess the current status of the wildlife reproduction and deformities BUI in this binational AOC.

METHODS

Two herring gull colonies within the St. Marys River AOC (Ontario), Hay Point (46°07'N, 83°59'W) and Pumpkin Point (46°23'N, 84°07'W), and one downstream reference colony at Double Island (46°10'N, 82°52'W) in the North Channel of Lake Huron were selected for study purposes. The common tern colony at Hay Point was the AOC colony used in this study. A tern colony on North Sister Rock (46°18'N,

83°54'W) was selected as an alternate AOC colony following abandonment of the Hay Point colony (likely due to predation) early in the breeding season of 2011. This colony is in St. Joseph Lake and is approximately 6 km beyond the AOC boundary at Quebec Bay. A tern colony on Cousins Island (46°04'N, 82°49'W) in the North Channel was selected as the reference colony.

Visits to each colony were made at two times in the breeding season: 1) egg laying (late April for gulls and late May for terns) and 2) when chicks were ≥ 21 days old (mid-May for gulls and mid-June for terns) in 2011 and 2012 to assess reproduction and various parameters of health. In 2011, an additional visit in May (i.e., immediately post-hatch) was conducted to assess deformity rates but this was not performed in 2012 in order to minimize disturbance associated with visits to the colony. During the first visit, 15-30 freshly-laid eggs (i.e., not incubated) were collected from one-egg nests at each colony for artificial incubation in the laboratory at the National Wildlife Research Centre (NWRC) in Ottawa. Embryonic viability, incidence of embryonic deformities, contaminant burdens, fatty acid profiles and stable isotope signatures were determined. In addition, a thorough nest count was conducted and contents were recorded. Individual nest enclosures (~1m in diameter and 16″ high) were constructed around ten to twelve 3-egg nests at each colony. As a measure of colony health, egg measurements for up to thirty 3-egg clutches were recorded (in millimetres) and egg volume calculated as:

Egg volume (cm³) = $K_{sp} x$ (length x breadth²)/1000

where K_{sp} =0.476 and 0.502 for herring gulls and common terns, respectively.

Total clutch volume was determined as the sum volume of the three eggs in the clutch. Intraclutch variation in egg size was calculated as the difference in volume between the largest and smallest egg in the clutch divided by the largest egg size (i.e., volume) and multiplied by 100. During the second visit, when chicks in enclosed nests were \geq 21 days old, productivity was calculated as:

Productivity = no. of \geq 21-day-old chicks/no. of enclosed nests

Enclosed nests that had been abandoned or where there was evidence that chicks had escaped were not included in estimates of productivity. Body measurements of 15 chicks, including mass, tarsus, wing cord, and culmen length (only measured in 2012), were also recorded and chicks were banded with a stainless steel USFW band. A 2 ml blood sample was collected from the brachial vein of chicks using a heparinized syringe to examine thyroxine concentrations in plasma (see below for details). In addition, two secondary covert feathers were collected to quantify corticosterone concentrations as a measure of stress over time in herring gull chicks in 2011 and 2012 and in common tern chicks in 2011 (see below). Chicks were examined for morphological deformities during the post-hatch visit in 2011 and when chicks were \geq 21 days old in 2012. An opportunistic deformity survey was also performed in 2011 on as many chicks as possible from nests outside of the enclosures.

Due to the general unpredictability of common terns during the nesting season (e.g., lack of fidelity to nesting colonies), there were challenges associated with determining productivity using the methods reported above. This was notably evident following abandonment of the Hay Point common tern colony early in the breeding season of 2011 where there was evidence of significant predation of tern eggs. Given these challenges, only one visit was made to Hay Point, North Sister Rock, and Cousins Island in late May for the purpose of collecting freshly-laid eggs for artificial incubation in the laboratory in 2012.

As part of a separate EC study of breeding site tenacity and productivity of common terns on the North Channel, productivity estimates are available for colonies at North Sister Rock and Cousins Island in 2011 and 2012 (D. Moore and D.V. Weseloh, EC unpublished), which will be reported here. In addition, egg measurements for 3-egg clutches of common terns at study sites were collected as part of this intensive EC study and will be included here. In 2011 and 2012, eggs were also measured at one additional AOC tern colony on South Sister Rock (46°18'N, 83°55'W) which is adjacent to North Sister Rock and hence just outside of the AOC boundary.

Artificial Incubation of Eggs:

Unincubated eggs were collected in the field, transported to NWRC in insulated coolers with foam inserts, gently cleaned and placed in a Petersime incubator (model# MX-1) at 37°C, 58% humidity and turned every two hours. Just prior to the pipping stage of development (i.e., embryonic day 26-27 for herring gulls and day 20-21 for common terns), embryos were removed from their shells and euthanized by decapitation. Each embryo was examined for physical deformities. Embryonic viability was determined as the number of viable embryos that survived to the designated embryonic day (i.e., just prior to pipping) divided by the total number of fertile eggs. Eggs that were nonviable were staged if possible (e.g., infertile; early, mid or late embryo death). Egg contents, including yolk sac, whole carcass and shell membranes, were collected in chemically-cleaned glass jars and frozen until chemical analysis for contaminants. Ten embryos were randomly selected from each colony for chemical analysis in the two study years. The one exception to this was at the Hay Point tern colony in 2011 where twenty embryos were randomly selected for analysis. This occurred because a second AOC tern colony was not available in that year for comparison purposes.

Contaminant Analyses:

Chemical analyses of individual herring gull and common tern embryos for organochlorine compounds, polybrominated diphenyl ethers (PBDEs), and mercury were conducted at NWRC. Organochlorine compounds measured included p, p'-DDE (dichlorodiphenyldichloroethylene), p, p'-DDT (dichlorodiphenyltrichloroethane), p,p'-DDD (dichlorodiphenyldichloroethane), oxychlordane, cischlordane, trans-chlordane, cis-nonachlor, trans-nonachlor, hexachlorobenzene (HCB), dieldrin, heptachlor epoxide (HE), octachlorostyrene (OCS), mirex, and PCBs. Sum chlordane is based on the sum concentrations of oxychlordane, cis-chlordane, trans-chlordane, cis-nonachlor, and trans-nonachlor. Prior to chemical analysis, thawed embryos were homogenized and then underwent neutral extraction and removal of lipids and biogenic compounds by gel permeation chromatography and Florisil column chromatography. Quantitative analysis of organochlorine compounds was performed using gas chromatography-mass selective detection (GC/MSD) operated in selected ion monitoring mode. The first and second injections were for the determinations of organochlorine pesticides and PCBs, respectively, and the third injection using GC/MSD chemical analysis in the NICI mode was for analysis of PBDEs. Sum PCBs and sum PBDEs were based on the sum concentrations of 62 individual or co-eluting PCB congeners and 15 PBDE congeners found above the limit of detection. Double-crested cormorant (Phalacrocorax auritus) egg reference material and two additional certified fish reference materials, blanks and duplicate samples were also analyzed for quality assurance purposes. Concentrations of

organochlorines and PBDEs are reported in μ g/g on a wet weight basis. The detection limit for both organochlorine compounds and PBDEs was 0.0001 μ g/g.

Embryos were also analyzed for non-*ortho* substituted PCBs, polychlorinated dibenzo-*p*-dioxins, including 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD), and polychlorinated dibenzofurans using gas chromatography-high resolution mass spectrometry (GC/HRMS) at RPC Laboratory in Fredericton, New Brunswick. Methods were based on US EPA Method 1613B and 8290A for dioxins and furans and on US EPA Method 1668C for non-*ortho* PCBs. Reference materials, blanks, and duplicates were also analyzed for quality assurance purposes. Following concerns regarding potential effects associated with elevated exposure to dioxin-like PCBs, dioxins and furans in birds in the AOC, deformed and normal embryos from St. Marys River AOC colonies were analyzed separately. A total of five deformed herring gull and common tern embryos from AOC colonies in 2011 and 2012 were analyzed as individuals. Contaminant concentrations in these individuals were then compared to concentrations in normal embryos from AOC colonies (analyzed as 2 or 3 pools) and each of the reference colonies (analyzed as a single pool). All pools consisted of five individual embryos.

Total mercury was quantified on a dry weight basis using an Advanced Mercury Analyzer (AMA-254) as described in CWS Method No. MET-CHEM-AA-03I. Certified reference materials and duplicate samples were also analyzed to ensure correct calibration, accuracy and reproducibility of test methods. Mercury concentrations in 2011 and 2012 embryos are reported in μ g/g on a wet weight basis using percent moisture content.

Stable Isotopes:

Stable isotope analyses of embryos were conducted at the University of Ottawa's G.G. Hatch Stable Isotope Laboratory in Ottawa, Ontario. Samples were weighed into tin capsules and loaded into an elemental analyser. The sample was flash combusted at about 1800° C (Dumas combustion) and the resultant gas products carried by helium through columns of oxidizing/reducing chemicals optimised for CO₂ and N₂. The gases were separated by a purge and trap adsorption column and then sent to the Delta Advantage isotope ratio mass spectrometer coupled with Conflo III. Samples were normalized to internal standards and calibrated to international standards. Stable isotope ratios are expressed in δ notation as the deviation from standards in parts per thousand (‰) according to the following relationship:

$$\delta X = (R_{sample} - R_{standard}) / R_{standard} \times 1000$$

where X is ¹⁵N or ¹³C and R is the corresponding ratio ¹⁵N/¹⁴N or ¹³C/¹²C. In this study, δ^{15} N signatures were compared to infer relative (and not absolute trophic) position at colonies.

Fatty acid analyses of embryos were also conducted to assess diet composition of egg-laying birds. These results are not yet available and, as such, will not be presented at this time.

Feather Corticosterone:

Feathers from chicks were prepared by first removing the calamus, i.e., the proximal end of the quill to where the feathers start (~ 5 to 10 mm; depending on length of feather). The remaining portion was then minced with scissors until homogenous and 1.5 ml methanol was added. The sample was vortexed, sonicated for 30 minutes at room temperature, and incubated overnight at 50° C while shaking. After centrifuging at 13,000 rpm for 20 minutes, the supernatant was transferred to a fresh tube and re-extracted with 1 ml methanol. The sample was again vortexed for 5 minutes and 0.5 ml of supernatant removed and added to the previous fraction. The methanol fraction was evaporated to dryness overnight in a fume hood and then the sample was reconstituted with 200 µl steroid diluent.

Corticosterone concentrations in the extracted feather sample were determined using a corticosterone EIA kit (Assay Designs – corticosterone enzyme immunoassay kit; product no. 900-097; 96 well kit). This is a competitive immunoassay for the quantitative determination of corticosterone in biological fluids. The kit uses a polyclonal antibody to corticosterone to bind in a competitive manner. Corticosterone in the standard or sample or an alkaline phosphatase molecule which has corticosterone covalently attached to it can be measured.

After a simultaneous incubation at room temperature, the excess reagents were washed away and substrate was added. After a short incubation time, the enzyme reaction was stopped and the sample was read on a SpectraMax 190 UV-VIS microplate reader (Molecular Devices, Sunnyvale, California, USA) at 405 nm. The intensity of the bound yellow color is inversely proportional to the concentration of corticosterone in either standards or samples. The measured optical density was used to calculate the concentration of corticosterone. The average net optical density (OD) bound for each standard and sample is calculated by subtracting the average NSB (non-specific binding) OD from the average OD bound:

Average Net OD = Average Bound OD - Average NSB OD

Then the binding of each pair of standard wells as a percentage of the maximum binding wells (Bo) was calculated using the following formula:

Percent Bound = Net OD x 100 Net Bo OD

The plot of percent bound versus concentration of corticosterone for the standards was graphed using Prism software (GraphPad, La Jolla, California, USA) and a line fitted through the points. The concentration of corticosterone in the unknowns was then determined. An in-house quality control sample was used in each plate. The sensitivity of the method was 27 pg/ml.

Thyroid Status:

Blood samples were centrifuged for 5 minutes at 14,000 rpm to separate plasma from red blood cells. Plasma was stored at -80°C and red blood cells were stored at 4°C. Concentrations of free thyroxine in the plasma of chicks sampled in 2012 were determined using a commercially available kit (AccuBind Elisa microwells product no. 1225-300 - Monobind Inc., Lake Forest, CA, 92630, USA). The method is based on a competitive enzyme immunoassay format in which a competition is set up between an immobilized antibody, the enzyme-antigen conjugate and the free thyroxine in the sample. When equilibration is reached, the unbound antigen fraction is removed and the enzyme activity in the bound fraction is measured which is inversely proportional to the concentration of free thyroxine in the sample. As per the method, standards, controls and sample plasma, as well as the enzyme reagent, were added to the microplate wells, which contained the immobilized antibody. After an incubation period, the unbound fraction was washed from the wells and the substrate added. The reaction was stopped and the plate was read at 450nm on a SpectraMax 190 UV-VIS microplate reader (Molecular Devices, Sunnyvale, California, USA). Concentrations were determined from the standard curve which was fitted using a variable slope (four parameter) method using Prism software (GraphPad, La Jolla, California, USA).

Controls used were Randox product no. HS2611 assayed human sera levels 2 & 3 (Randox Laboratories Ltd., Antrim, UK). The method gave the intra-assay precision as 10.98, 4.26 and 3.25 % coefficient of variation for low, medium and high controls, respectively. The inter-assay precision was 10.81, 6.01 and 7.90 % coefficient of variation for low, medium and high controls, respectively. The sensitivity of the assay was 0.05 ng/dl.

Thyroxine was also quantified in plasma of herring gull and common tern chicks sampled in 2011 using a different type of kit (radioimmunoassay) and, as a result, is not directly comparable to concentrations determined using the method outlined above for 2012 samples. Thyroxine was also quantified in plasma of embryos from artificially incubated eggs in 2011 but was found at concentrations below the limit of detection in all samples.

Statistical Analysis:

Contaminants and other biological endpoints were statistically analyzed using either the Student's *t*-test for between-colony comparisons or a one-way ANOVA for among-colony comparisons, which when significant, was followed by Tukey's HSD test. Data were log-transformed (log₁₀) to meet conditions of equal variance and normality for parametric analysis. If data failed these assumptions, comparisons were made using either a Mann-Whitney U non-parametric test or Kruskal-Wallis one way analysis of variance by ranks; post-hoc tests were conducted using non-parametric multiple comparison tests for unequal sample sizes. Concentrations of contaminants found below the limit of detection were given a concentration of one-half of the detection limit. Mercury concentrations in embryos were statistically analyzed on a dry weight basis however are reported on a wet weight basis for comparisons to published values and thresholds. A Spearman rank correlation analysis was performed to examine the relationship between the two stable isotopes for each species. Due to the overall low number of deformed embryos found, the Fisher exact test (one-tailed) was used to compare counts of deformed and non-deformed embryos at AOC colonies and reference colonies which were combined in the two study years. All results were considered significant at p<0.05.

Concentrations of 2,3,7,8-TCDD toxic equivalents (TEQs) were calculated for dioxin-like PCBs, furans, and dioxins and are based upon toxic equivalency factors developed by van den Berg *et al.* (1998) for birds. Dioxin-like PCBs include four non-*ortho* PCB congeners (77, 81, 126, and 169) and eight mono-*ortho* PCB

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congeners (105, 114, 118, 123, 156, 157, 167, and 189). Two of these eight mono-*ortho* PCBs (123 and 167) were not quantified as individual congeners in this study and were not included in the calculation of TEQs. For normal embryos analyzed as pools, mean mono-*ortho* PCB concentrations, quantified in the chemical analysis for organochlorines, were calculated for individuals used to create the pool (where data were available). Due to the random sampling of individuals selected for organochlorine analysis, mono-*ortho* PCB concentrations were not available for one deformed herring gull embryo from Hay Point in 2011. Total TEQ concentration is based on the sum concentration of TEQs calculated for the 4 non-*ortho* PCBs, 6 mono-*ortho* PCBs, and 17 dioxin and furan congeners. Concentrations were compared between deformed embryos and normal embryos of herring gulls from AOC colonies in 2011 and 2012 using a Student's *t*-test; a similar statistical comparison was not possible for common terns due to low sample size (N=2).

RESULTS

A) Artificial Egg Incubation Study

Embryonic Viability and Deformities:

i) Herring gulls

Embryonic viability was high in herring gulls from the two AOC colonies, Pumpkin Point and Hay Point, and ranged between 92%-100% in 2011 and 2012 (Table 1a). Embryonic viability was also high at the Double Island reference colony at 86% and 96% in 2011 and 2012, respectively. Of 52 fertile eggs examined in 2011, a total of five embryos died: one from Pumpkin Point (at developmental stage 26), one from Hay Point (stage 39) and three from Double Island (stages 19, 35, and 39). Of 50 fertile eggs examined in 2012, two embryos died, one from Pumpkin Point (stage 31) and one from Double Island (stage 29). Embryonic deformities were evident in a single embryo from both AOC colonies in at least one study year. Incidences of deformities at Pumpkin Point were 6% (1 deformed embryo/16 viable embryos) in 2011 and 8% (1/12) in 2012 and at Hay Point were 8% (1/13) in 2011 and 0% (0/12) in 2012. Of the two deformed embryos from Pumpkin Point in 2011 and 2012, one had a crossed-bill and part of the lower brain was exposed while the other individual had a split lower mandible. The one deformed embryo from Hay Point had a slightly off-centre lower mandible. No embryonic deformities were evident in incubated eggs from Double Island in either study year. No significant difference was found in the incidence of herring gull embryonic deformities between AOC colonies and the reference colony when counts for sites and years were combined.

ii) Common terns

Embryonic viability among artificially-incubated common tern eggs from the two AOC colonies, Hay Point and North Sister Rock, in 2011 and 2012 was high and ranged between 90%-100% (Table 1b). Embryonic viability at the Cousins Island reference colony was comparable at 93% in 2011 and 100% in 2012. Of the 44 fertile eggs examined in 2011, four embryos died: three from Hay Point (stages 6, 8, and 17) and one from Cousins Island (stage 17). Of the 37 fertile eggs examined in 2012, one embryo from North Sister died (stage 25). Similar to herring gulls, embryonic deformities were evident in a single embryo from each of the two AOC colonies in one study year. Incidences of embryonic deformities were 4% (1/26) at Hay Point in 2011 and 8% (1/13) at North Sister Rock in 2012. The deformed embryo from Hay Point had one eye, half a skull and the lower body was shortened while the deformed individual

Table 1. Embryonic viability and incidence of embryonic deformities in artificially-incubated eggs of herring gulls (a) and common terns (b) collected from St. Marys River AOC colonies (Hay Point, Pumpkin Point, North Sister Rock) and corresponding Lake Huron reference colonies (Double Island or Cousins Island) in 2011 and 2012.

Colony	AOC/ Ref	Year	Total No. Eggs	No. Infertile Eggs	No. Fertile Eggs	No. Viable Eggs	No. Dead Eggs	Embryonic Viability (%)	No. Deformities	Deformities (%)
Hay Point	AOC	2011	15	1	14	13	1	93%	1	8%
		2012	15	3	12	12	0	100%	0	0%
Pumpkin Point	AOC	2011	17	0	17	16	1	94%	1	6%
		2012	15	2	13	12	1	92%	1	8%
Double I.	Ref	2011	23	2	21	18	3	86%	0	0%
		2012	26	1	25	24	1	96%	0	0%

a) Herring gulls:

b) Common terns:

Site	AOC/ Ref	Year	Total No. Eggs	No. Infertile Eggs	No. Fertile Eggs	No. Viable Eggs	No. Dead Eggs	Embryonic Viability (%)	No. Deformities	Deformities (%)
Hay Point	AOC	2011	30	1	29	26	3	90%	1	4%
		2012	15	2	13	13	0	100%	0	0%
North Sister Rock	AOC	2012	15	1	14	13	1	93%	1	8%
Cousins I.	Ref	2011	15	0	15	14	1	93%	0	0%
		2012	15	5	10	10	0	100%	0	0%

from North Sister Rock had long thin limbs. No embryonic deformities were evident in incubated eggs from Cousins Island in either study year. No significant difference was found in the incidence of common tern embryonic deformities between AOC colonies and the reference colony when counts for sites and years were combined.

Contaminants:

i) Herring gulls

Concentrations of organochlorine compounds were low overall in herring gull embryos from the two AOC colonies and the reference colony in 2011 and 2012 with sum PCBs found at the highest concentrations (Table 2). Mean sum PCB concentrations ranged from 1.13 µg/g in embryos from Double Island in 2012 to 1.92 µg/g in embryos from Pumpkin Point in 2011. The maximum sum PCB concentration of 4.88 µg/g was found in an embryo from Double Island in 2011. Mean concentrations of the remaining organochlorines in embryos were below $0.4 \,\mu g/g$ at the three colonies. Sum PBDE concentrations (determined as the sum concentration of 15 PBDE congeners) were comparable to concentrations of p, p'-DDE in embryos and, based on comparisons of means, were at least six times higher than concentrations of other organochlorine pesticides. The maximum sum PBDE concentration (1.01 µg/g) was found in an embryo from Pumpkin Point in 2012. Generally, no significant differences in concentrations of organochlorines, including sum PCBs, and sum PBDEs were found in herring gull embryos among the three colonies in either 2011 or 2012. The one exception was for octachlorostyrene (OCS) in 2011 in which a significantly higher mean concentration was found in embryos from Pumpkin Point compared to those from the Hay Point AOC colony and Double Island reference colony. Concentrations of this compound, however, were among the lowest of all measured in this study. Mean percent lipid content was not significantly different among colonies within a study year and ranged from 6.5% in herring gull embryos from Double Island in 2011 to 8.4% in embryos from Pumpkin Point in 2012.

Mean concentrations of four non-ortho PCBs, 2,3,7,8-TCDD, and total TEQs were largely comparable among deformed and normal herring gull embryos from St. Marys River AOC colonies and the Double Island reference colony in 2011 and 2012 (Table 3a). Of the four non-ortho PCBs measured in deformed herring gull embryos, concentrations of PCB-126 > 77 > 81 ≈ 169. Patterns in normal embryos from AOC colonies and the reference colony were slightly different with concentrations of PCB-126 > 169 > 81 \approx 77. The mean 2,3,7,8-TCDD concentration in three deformed embryos (4.76 pg/g) was within the range of concentrations found for normal embryos from AOC colonies (3.30 pg/g) and normal embryos from the reference colony (5.62 pg/g) which was also the maximum concentration found in this study. Mean total TEQ concentrations ranged from 76.78 pg TEQ/g in normal embryos from AOC colonies to 112.76 pg TEQ/g in deformed embryos from AOC colonies. The maximum total TEQ concentration (159.17 pg TEQ/g) was found in a deformed embryo from Hay Point in 2011. Toxicity associated with non-ortho PCBs, dioxins and furans, and mono-ortho PCBs contributed 84%, 11%, and 7%, respectively, to the mean total TEQ concentration in deformed embryos from AOC colonies in 2011 and 2012. Similar contributions of toxicity associated with non- ortho PCBs, dioxins and furans, and mono-ortho PCBs to mean total TEQ concentrations were found for normal embryos from AOC colonies (78%, 12%, 10%, respectively) and the reference colony (77%, 13%, and 9%, respectively). Concentrations of non-ortho

Table 2. Mean concentrations (SD) of organochlorines and sum PBDEs in herring gull embryos (wet weights) collected from the St. Marys River AOC (Hay Point and Pumpkin Point) and Double Island in Lake Huron (reference colony) in 2011 and 2012 and incubated in the lab. All compounds are shown in $\mu g/g$. N values represent number of individual embryos analyzed. Different uppercase letters show significant differences in mean concentrations between colonies within a given year based on a one-way ANOVA.

Colony	AOC/ Ref	Year	p,p'- DDE	Sum Chlordane	НСВ	Dieldrin	HE	Mirex	OCS	PCB 1:1 ¹	Sum PCBs ²	Sum PBDEs ³
Hay Point	AOC	2011	0.376	0.037	0.015	0.024	0.013	0.026	0.002	3.435	1.807	0.244
N=10	AUC	2011	(0.173)	(0.013)	(0.005)	(0.012)	(0.004)	(0.034)	(0.001) B	(1.569)	(0.842)	(0.093)
Pumpkin	100	2011	0.304	0.036	0.018	0.022	0.012	0.013	0.004	3.036	1.922	0.237
Point N=10	AOC	2011	(0.092)	(0.007)	(0.008)	(0.009)	(0.002)	(0.018)	(0.001) A	(1.219)	(0.773)	(0.094)
Double I.	Def	2011	0.324	0.031	0.015	0.019	0.011	0.033	0.002	3.196	1.766	0.311
N=10	Ref	2011	(0.127)	(0.010)	(0.006)	(0.016)	(0.004)	(0.038)	(0.001) B	(2.218)	(1.338)	(0.198)
Hay Point	100	2012	0.248	0.035	0.009	0.009	0.012	0.010	0.002	2.824	1.433	0.333
N=10	AOC	2012	(0.125)	(0.025)	(0.005)	(0.007)	(0.015)	(0.009)	(0.003)	(1.181)	(0.671)	(0.147)
Pumpkin	100	2012	0.292	0.029	0.008	0.010	0.008	0.020	0.001	2.359	1.273	0.452
Point N=10	AOC	2012	(0.111)	(0.007)	(0.003)	(0.006)	(0.004)	(0.022)	(0.001)	(1.007)	(0.728)	(0.258)
Double I.	Def	2012	0.218	0.025	0.008	0.007	0.006	0.010	0.002	2.187	1.131	0.332
N=10	Ref	2012	(0.048)	(0.005)	(0.005)	(0.005)	(0.001)	(0.013)	(0.002)	(1.177)	(0.704)	(0.185)

¹Based on 1:1 ratio of Arochlor 1254:1260

² Based on the sum of 62 PCB congeners

³ Based on the sum of 15 PBDE congeners

Table 3. Mean concentrations (SD) of non-*ortho* PCBs, 2,3,7,8-TCDD, and 2,3,7,8-TCDD toxic equivalents (TEQ) in embryos of herring gulls (a) and common terns (b) collected from the St. Marys River (SMR) AOC and reference colonies (i.e., Double Island and Cousins Island) in 2011 and 2012 and incubated in the lab (pg/g, wet weight). Concentrations are shown for deformed embryos (analyzed as individuals) and normal embryos based on the analysis of pools consisting of five individuals. Accordingly, N represents either the number of deformed individuals or the number of pools analyzed. TEQs associated with 4 non-*ortho* PCBs, 17 dioxins and furans (PCDD/Fs), and 6 mono-*ortho* PCBs and which together comprise total TEQs are also provided.

a) Herring gulls:

Colony	AOC/	N	PCB-77	PCB-81	PCB-126	PCB-169	2,3,7,8 –	TEQ – non-	TEQ –	TEQ – mono-	Total
	Ref			100 01	1 60 120	100 100	TCDD	<i>ortho</i> PCBs	PCDD/Fs	ortho PCBs ¹	TEQs
SMR ² -	AOC	С	249.07	120.10	699.00	112.70	4.76	94.48	12.93	8.03	112.76
Deformed	AUC	5	(384.48)	(53.96)	(149.79)	(54.67)	(0.51)	(36.38)	(5.10)	(0.82)	(42.36)
SMR ² -	AOC	С	61.90	72.30	497.67	92.07	3.30	60.18	9.31	7.30	76.78
Normal	AUC	5	(71.18)	(9.52)	(70.12)	(20.57)	(1.30)	(10.78)	(4.21)	(0.96)	(15.84)
Double I	Dof	1	26.70	08 00	680.00	121 00	E 60	70.9E	12.00	0.72	102.46
Normal	Ref	T	36.70	98.90	680.00	121.00	5.62	79.85	13.89	9.73	103.46

b) Common terns:

Colony	AOC/ Ref	Ν	PCB-77	PCB-81	PCB-126	PCB-169	2,3,7,8 – TCDD	TEQ – non- <i>ortho</i> PCBs	TEQ – PCDD/Fs	TEQ – mono- <i>ortho</i> PCBs	Total TEQs
SMR ³ -	AOC	2	617.25	55.58	389.75	68.03	2.49	75.46	19.19	3.33	97.98
Deformed	AUC	Z	(135.41)	(25.49)	(259.15)	(37.16)	(1.84)	(35.27)	(11.76)	(1.94)	(48.96)
SMR ³ -	AOC	r	548.50	48.05	346.00	70.20	2.14	66.90	16.09	3.35	86.34
Normal	AOC	Z	(9.19)	(5.73)	(82.02)	(18.95)	(0.64)	(8.33)	(2.61)	(0.60)	(6.32)
Cousins I Normal	Ref	1	1000.00	75.80	455.00	85.20	3.80	103.17	33.24	2.73	139.14

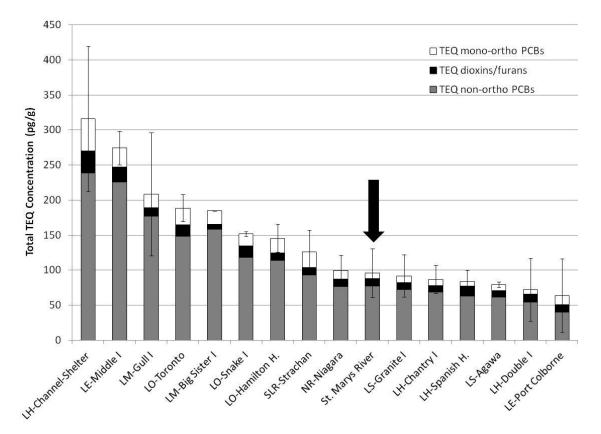
¹Since TEQ—mono-*ortho* PCBs were quantified in individual embryos (as part of the sum PCB analysis), these concentrations were determined using the mean concentration of individuals which comprised the pool (where data were available with a range of between 1-5 individuals); the mean TEQ—mono*ortho* PCB concentration for deformed herring gull embryos from SMR is based on N=2 rather than N=3 (see methods for details).

²Comprise individuals from Hay Point and Pumpkin Point

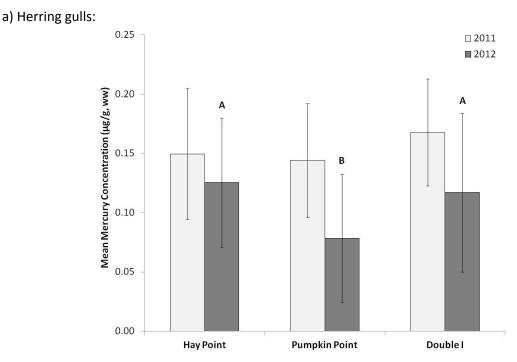
³ Comprise individuals from Hay Point and North Sister Rock

PCBs, 2,3,7,8-TCDD, and TEQs were not significantly different between deformed and normal herring gull embryos from AOC colonies (p>0.05). Overall, the mean total TEQ concentration in deformed and normal embryos from the St. Marys River AOC (N=6 samples total) was 94.77 pg TEQ/g in 2011 and 2012 and was within the range of mean TEQ concentrations in herring gull eggs from other colonies on the Great Lakes in these two years (Figure 2; EC unpublished). A full listing of mean concentrations of non-*ortho* PCBs, mono-*ortho* PCBs, dioxins and furans in deformed and/or normal herring gull embryos from St. Marys River AOC colonies and the Double Island reference colony is provided in Appendix 1.

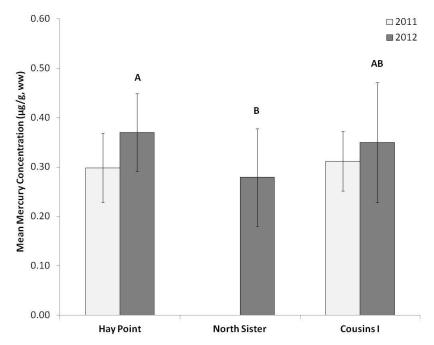
Figure 2. Mean total TEQ concentrations (SD) in deformed and normal herring gull embryos (N=6 samples total) from St. Marys River AOC (Ontario) colonies and in herring gull eggs from Great Lakes colonies in 2011 and 2012 (pg/g, wet weight). The contributions of TEQ concentrations associated with mono-*ortho* PCBs, dioxins and furans, and non-*ortho* PCBs to the total TEQ concentration are shown. Means are arranged in decreasing order from highest to lowest concentrations. Colony locations are associated with the following lakes/rivers: LH=Lake Huron, LO=Lake Ontario, LE=Lake Erie, SLR=St. Lawrence River, NR=Niagara River, LS=Lake Superior, and LM=Lake Michigan.



Mean mercury concentrations (SD) in herring gull embryos in 2011 were statistically comparable among the three study sites ranging from 0.14 (0.05) μ g/g wet weight at Pumpkin Point to 0.17 (0.05) μ g/g at Double Island (Figure 3a). In 2012, some spatial differences among colonies were evident with mean mercury concentrations (SD) ranging from 0.08 (0.05) μ g/g at Pumpkin Point to 0.13 (0.05) μ g/g at Hay Point. Mercury concentrations in embryos from Hay Point were statistically similar to those from Double Island and embryos from both colonies had significantly higher mercury concentrations than embryos Figure 3. Mean concentrations (SD) of total mercury (μ g/g, wet weight) in embryos of herring gulls (a) and common terns (b) collected from St. Marys River AOC colonies (Hay Point, Pumpkin Point, North Sister Rock) and corresponding Lake Huron reference colonies (Double Island or Cousins Island) in 2011 and 2012 (N=12-30 embryos/colony). Different uppercase letters show significant differences in mean concentrations within a study year and are based on statistical analysis of dry weight mercury concentrations. Note no data are available for common terns at North Sister Rock in 2011.



b) Common terns:



from Pumpkin Point ($F_{2,42}$ =5.54, p=0.007). The maximum mercury concentration was 0.28 µg/g in a herring gull embryo from Hay Point in 2011.

ii) Common terns

Contaminant burdens were also very low in common terns from AOC and reference colonies in 2011 and 2012 (Table 4). Sum PCBs were found at the highest concentrations relative to other organochlorines with means ranging from 0.77 μ g/g in embryos from Hay Point in 2011 to 1.43 μ g/g in embryos from Cousins Island in 2012. The maximum sum PCB concentration reported (2.58 μ g/g) was found in an embryo from the Cousins Island reference colony in 2012. Similar to herring gulls, sum PBDEs in common tern embryos at study sites were comparable in concentration to $p_{,p}$ '-DDE and higher than concentrations of other organochlorine pesticides. The maximum sum PBDE concentration (0.21 μ g/g) was found in an embryo from North Sister Rock in 2012. Some spatial differences were apparent between/among colonies in each of the two study years but in a somewhat unexpected direction. In 2011, significantly higher concentrations of sum PCBs, sum PBDEs, p,p'-DDE, sum chlordane and OCS were found in common tern embryos from the Cousins Island reference colony compared to the Hay Point AOC colony. Concentrations of HCB, dieldrin, HE and mirex were comparable between study sites. In 2012, significantly higher concentrations of p, p'-DDE, mirex and OCS were found in embryos from the Cousins Island reference colony compared to the Hay Point AOC colony while concentrations were comparable between the reference colony and the North Sister Rock AOC colony. In 2012, concentrations of other contaminants including sum PCBs and sum PBDEs were similar among study sites. Percent lipid content was not significantly different between colonies in 2011 with means in common tern embryos of 6.8% and 7.6% at Hay Point and Cousins Island, respectively. In 2012, percent lipid content was significantly higher in embryos from Cousins Island (mean=9.8%) compared to Hay Point (mean=8.8%; F_{2.27}=5.64, p=0.009). As such, the results reported in Table 3 are based on contaminants data in 2012 which were first lipid-normalized prior to statistical analysis.

Mean concentrations of four non-ortho PCBs, 2,3,7,8-TCDD, and total TEQs were comparable between deformed and normal common tern embryos from St. Marys River AOC colonies in 2011 and 2012 (Table 3b). Concentrations in tern embryos from the Cousins Island reference colony (analyzed as a pool) were generally slightly higher compared to embryos from AOC colonies and most frequently were the maximum concentration found of all embryos. Of the four non-ortho PCBs measured in common tern embryos, concentrations of PCB-77 > 126 > 169 \approx 81, a pattern which differed to that found in herring gull embryos. The mean 2,3,7,8-TCDD concentration in two deformed embryos (2.49 pg/g) was within the range of concentrations found for normal embryos from AOC colonies (2.14 pg/g) and normal embryos from the reference colony (3.80 pg/g). Mean total TEQ concentrations ranged from 86.34 pg TEQ/g in normal embryos from AOC colonies to 139.14 pg TEQ/g in normal embryos from the reference colony. Toxicity associated with non-ortho PCBs, dioxins and furans, and mono-ortho PCBs contributed 77%, 20%, and 3%, respectively, to the mean total TEQ concentration in deformed embryos from AOC colonies in 2011 and 2012. Similar contributions of toxicity associated with non-ortho PCBs, dioxins and furans, and mono-ortho PCBs to mean total TEQ concentrations were found for normal embryos from the AOC and the reference colony. Due to low sample sizes, it was not possible to statistically compare concentrations of non-ortho PCBs, 2,3,7,8-TCDD, and TEQs between deformed and normal common tern embryos from AOC colonies. An examination of mean concentrations with associated variation however

Table 4. Mean concentrations (SD) of organochlorines and sum PBDEs in common tern embryos (wet weights) collected from the St. Marys River AOC (Hay Point and North Sister Rock) and Cousins Island in Lake Huron (reference site) in 2011 and 2012 and incubated in the lab. All compounds are shown in µg/g. N values represent number of individual embryos analyzed. Different uppercase letters show significant differences in mean concentrations between colonies within a given year based on a one-way ANOVA.

Colony	Ref	Year	p,p'-DDE	Sum Chlordane	НСВ	Dieldrin	HE	Mirex	ocs	PCB 1:1 ¹	Sum PCBs ²	Sum PBDEs ³
Hay Point	AOC	2011	0.099	0.009	0.011	0.014	0.004	0.004	0.0013	1.348	0.768	0.065
N=20	AOC	2011	(0.038) B	(0.005) B	(0.003)	(0.005)	(0.002)	(0.002)	(0.0005) B	(0.590) B	(0.300) B	(0.021) B
Cousins I.	Dof	2011	0.178	0.014	0.013	0.016	0.005	0.005	0.0021	2.116	1.147	0.094
N=10	Ref	2011	(0.044) A	(0.005) A	(0.004)	(0.006)	(0.002)	(0.003)	(0.0006) A	(0.370) A	(0.165) A	(0.024) A
Hay Point	100	2012	0.146	0.009	0.013	0.017	0.006	0.003	0.0012	1.902	0.890	0.090
N=10	AOC	2012	(0.061) B	(0.006)	(0.004)	(0.006)	(0.002)	(0.001) B	(0.0005) B	(0.703)	(0.305)	(0.032)
North Sister	100	2012	0.181	0.012	0.016	0.019	0.006	0.005	0.0016	2.474	1.146	0.124
N=10	AOC	2012	(0.046) AB	(0.009)	(0.004)	(0.004)	(0.002)	(0.002) A	(0.0004) AB	(0.689)	(0.311)	(0.387)
Cousins I.	Def	2012	0.238	0.015	0.019	0.017	0.005	0.006	0.0020	3.036	1.430	0.140
N=10	Ref	2012	(0.076) A	(0.013)	(0.005)	(0.004)	(0.001)	(0.002) A	(0.0007) A	(1.262)	(0.602)	(0.032)

¹Based on 1:1 ratio of Arochlor 1254:1260

² Based on the sum of 62 PCB congeners

³ Based on the sum of 15 PBDE congeners

suggests that concentrations between these two groups are very similar. A full listing of mean concentrations of non-*ortho* PCBs, mono-*ortho* PCBs, dioxins and furans in deformed and/or normal common tern embryos from St. Marys River AOC colonies and the Cousins Island reference colony is provided in Appendix 1.

Mercury concentrations in common tern embryos in 2011 were comparable at Hay Point and Cousins Island with means (SD) of 0.30 (0.07) μ g/g and 0.31 (0.06) μ g/g, respectively (Figure 3b). In 2012, mean mercury concentrations (SD) ranged from 0.28 (0.10) μ g/g at North Sister Rock to 0.37 (0.08) μ g/g at Hay Point and, similar to that found for herring gulls, a significant spatial pattern among colonies was evident in this year (F_{2,36}=3.83, p=0.031). While mean mercury concentrations were statistically similar between each of the AOC colonies and the Cousins Island reference colony, mercury concentrations were significantly higher in common terns from Hay Point compared to those from North Sister Rock. The maximum mercury concentration was 0.59 μ g/g in a common tern embryo from Cousins Island in 2012.

Stable Isotopes:

i) Herring gulls

Significant spatial differences for mean δ^{15} N values in herring gull embryos were found among study sites in 2011 ($F_{2,42}$ =6.94, p=0.003) and 2012 ($F_{2,42}$ =4.97, p=0.013; Table 5a). As an indicator of trophic position, mean δ^{15} N values were significantly higher in gulls from the Double Island reference colony compared to one or both of the AOC colonies in both study years. In addition, mean δ^{13} C values in gull embryos from Double Island were also significantly more depleted (i.e., more negative) than mean values in embryos from the two AOC colonies in both 2011 ($F_{2,42}$ =23.23, p<0.00001) and 2012 ($F_{2,42}$ =11.22, p=0.0001). No significant correlation was found between δ^{15} N and δ^{13} C values when herring gull embryos from all colonies and years were grouped together.

ii) Common terns

Similar spatial patterns for isotopes were found in common tern embryos (Table 5b). Mean δ^{15} N values were significantly higher in common terns from the Cousins Island reference colony compared to one or both of the AOC colonies in 2011 (Mann Whitney U=450, p<0.00001) and 2012 ($F_{2,36}$ =32.63, p<0.00001). Mean δ^{13} C values in tern embryos from Cousins Island were significantly more depleted (i.e., more negative) than mean values in embryos from the AOC colonies in both 2011 (Mann Whitney U=91, p=0.001) and 2012 (Kruskal Wallis test H=9.92, p=0.007). Unlike that found for herring gulls, a highly significant negative correlation was found between δ^{15} N and δ^{13} C values when common tern embryos from all colonies and years were grouped together (r_s =0.72, p<0.00001).

Table 5. Mean (SD) values for δ^{15} N and δ^{13} C in embryos of herring gulls (a) and common terns (b) collected from St. Marys River AOC colonies (Hay Point, Pumpkin Point, North Sister Rock) and corresponding Lake Huron Double Island reference colonies (Double Island or Cousins Island) in 2011 and 2012 (N=12-30 embryos/colony). Different uppercase letters indicate significant differences between colonies within a study year.

Colony	AOC/Ref	δ	¹⁵ N	δ1	³ C
		2011	2012	2011	2012
Hay Point	AOC	9.00 (0.82) B	8.61 (0.44) B	-20.40 (1.00) A	-21.32 (1.11) A
Pumpkin Point	AOC	8.94 (0.39) B	8.77 (0.47) AB	-20.75 (0.87) A	-21.49 (1.20) A
Double I.	Ref	9.80 (0.82) A	9.29 (0.88) A	-22.49 (0.82) B	-23.05 (1.00) B

a) Herring gulls:

b) Common terns:

Colony	AOC/Ref	δ1	δ ¹⁵ N		³ C
		2011	2012	2011	2012
Hay Point	AOC	10.57 (0.61) B	10.53 (0.69) B	-25.16 (1.09) A	-25.61 (0.88) A
North Sister Rock	AOC	NA	11.07 (0.56) B	NA	-25.32 (1.48) A
Cousins I.	Ref	12.12 (0.32) A	12.29 (0.33) A	-25.95 (1.02) B	-26.63 (0.50) B

B) Field Study

Egg Size Parameters:

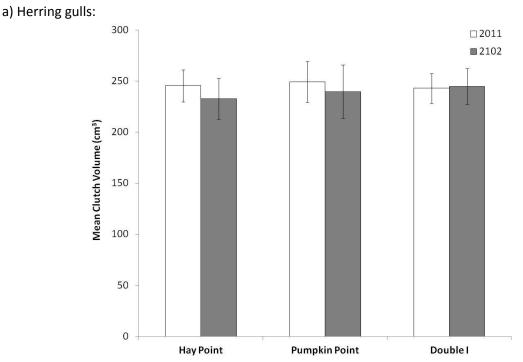
i) Herring gulls

To assess potential food stress in birds, total clutch volume and intraclutch variation in egg size were examined in 3-egg clutches. Mean total clutch volumes (SD) for herring gulls from St. Marys River AOC colonies ranged from 232.8 (19.9) cm³ at Hay Point in 2012 to 249.2 (19.9) cm³ at Pumpkin Point in 2011 and overall were not significantly different from total clutch volumes in gulls from the Double Island reference colony in either of the two study years (Figure 4a). Mean intraclutch variation (SD) in egg size from AOC colonies in the two study years ranged from 9.5 (5.5)% at Pumpkin Point in 2011 to 12.0 (7.2)% at Hay Point in 2012 and were also statistically comparable to means (SD) at the Double Island reference colony of 11.1 (3.3)% and 7.3 (4.4)% in each of 2011 and 2012, respectively.

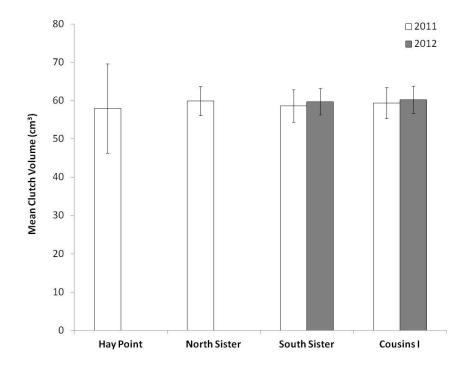
ii) Common terns

Mean total clutch volume (SD) in 3-egg clutches of common terns at three AOC colonies in 2011 ranged from 58.0 (11.6) cm³ at Hay Point to 60.0 (3.8) cm³ at North Sister Rock and were not significantly different from mean clutch volume at Cousins Island (59.4 (4.0) cm³; Figure 4b). Similarly in 2012, mean total clutch volume (SD) was not significantly different between the South Sister Rock AOC colony (59.8 (3.5) cm³) and the Cousins Island colony (60.3 (3.6) cm³). Mean intraclutch variation (SD) in egg size at AOC tern colonies in the two years ranged from 6.8 (3.2)% at South Sister Rock in 2011 to 9.5 (7.6)% at Hay Point in 2011. Despite relatively lower estimates of mean intraclutch variation (SD) in egg size from Cousins Island of 6.2 (2.8)% and 6.0 (2.6)% in 2011 and 2012, respectively, no significant differences in intraclutch variation were found between AOC tern colonies and the reference colony in either of the two study years.

Figure 4. Mean clutch volume (SD) in 3-egg clutches of herring gulls (a) and common terns (b) from St. Marys River AOC colonies (Hay Point, Pumpkin Point, North Sister Rock, South Sister Rock) and corresponding Lake Huron reference colonies (Double Island or Cousins Island) in 2011 and 2012. Numbers of 3-egg clutches ranged from 8-38 for both species with the exception of the Hay Point common tern colony in 2011 where the mean is based on three clutches of eggs. Note no data are available for common terns at Hay Point and North Sister Rock in 2012.



b) Common terns:

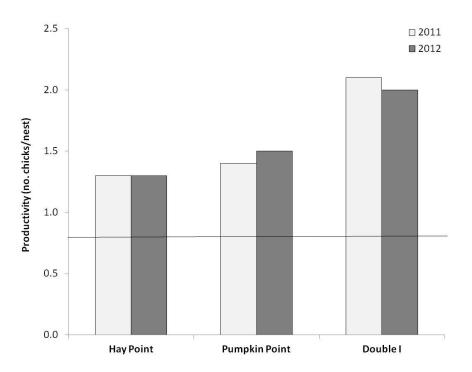


Productivity & Prevalence of Deformities in Wild Populations:

i) Herring gulls

Herring gull productivity, defined as the number of \geq 21-day-old chicks produced per nest, was equal to 1.3 chicks per nest at Hay Point (N=7 enclosed nests), 1.4 chicks per nest at Pumpkin Point (N=10 nests), and 2.1 chicks per nest at Double Island (N=9 nests) in 2011 (Figure 5). Similarly, productivity was high in 2012 with estimates equal to 1.3 chicks per nest at Hay Point (N=12 nests), 1.5 chicks per nest at Pumpkin Point (N=10 nests), and 2.0 chicks per nest at Double Island (N=7 nests). Productivity estimates at AOC colonies in both study years were well within the range of productivity levels required to maintain a stable population (0.8-1.4 chicks per nest; Kadlec and Drury 1968).

Figure 5. Herring gull productivity, as the number of \geq 21-day-old chicks produced per nest, at St. Marys River AOC colonies (Hay Point and Pumpkin Point) and the Double Island reference colony in 2011 and 2012. The solid line indicates the minimum productivity level of 0.8 chicks per nest associated with maintaining a stable herring gull population (range in levels=0.8-1.4 chicks per nest; Kadlec and Drury 1968).



In 2011, no morphological deformities were found in herring gull chicks from the two AOC colonies or the reference colony (N=39-76 chicks). This count included chicks from inside and outside of the enclosures (Table 6). Similarly, no deformities were found in gull chicks from enclosed nests at the three colonies in 2012 where numbers of chicks examined ranged from 14-16 per colony.

Table 6. Prevalence of morphological deformities (%) in herring gull chicks examined in enclosed nests and non-enclosed nests from St. Marys River AOC colonies (Hay Point, Pumpkin Point) and the Double Island reference colony in 2011 and 2012.

Colony	Year	No. Chick	% Deformities	
		Enclosed	Non-enclosed	
		Nests	Nests	
Hay Point	2011	19	20	0%
	2012	16	-	0%
Pumpkin Point	2011	23	40	0%
	2012	15	-	0%
Double I.	2011	31	45	0%
	2012	14	-	0%

ii) Common terns

Common tern productivity, also defined as the number of \geq 21-day-old chicks produced per nest, was equal to 0.8 chicks per nest at North Sister Rock in 2011 (N=10 enclosed nests; D. Moore and D.V. Weseloh 2012). This productivity estimate was higher than the estimate of 0.3 chicks produced per nest at the Cousins Island reference colony (N=20 nests). In 2012, tern productivity at 15 enclosed nests was equal to 0 at North Sister Rock as a result of predation of eggs in enclosures. There was however some evidence of renesting and successful nests later in the season at this colony (D. Moore, pers. comm.) Although considered low, productivity at Cousins Island was relatively higher in 2012 at 0.6 chicks per nest (N=16 nests). Productivity estimates for all study colonies were well below the threshold of 1.1 common tern chicks per nest associated with a stable population (DiCostanzo 1980).

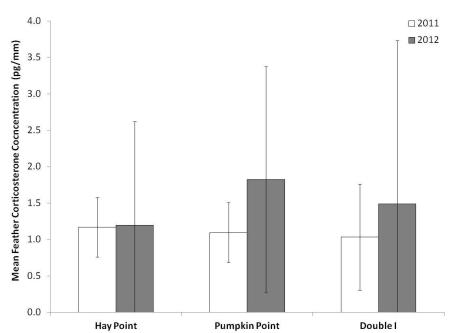
No deformities were observed in common tern chicks examined at non-enclosed nests at North Sister Rock and the Cousins Island reference colony in 2011 (N=13 and 10 chicks respectively). Since common tern colonies were not visited a second time (i.e., when chicks were \geq 21 days old) in 2012, deformity estimates are not available.

Corticosterone in Feathers:

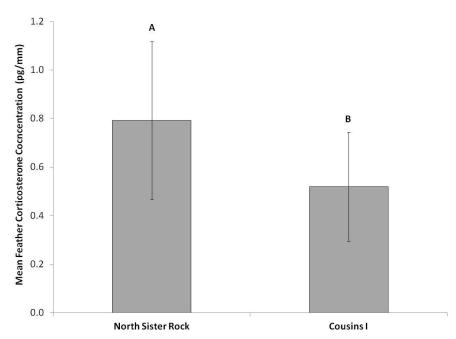
Mean corticosterone concentrations (SD) in herring gull chick feathers from the AOC colonies ranged from 1.1 (0.41) pg/mm to 1.8 (1.6) pg/mm at Pumpkin Point in 2011 and 2012, respectively, and overall were not significantly different from concentrations at the Double Island reference colony in either of the two study years (Figure 6a). However, a significant difference was found for common terns in 2011 (Figure 6b). Chicks from North Sister Rock had a significantly higher mean feather corticosterone concentration compared to those from Cousins Island (t_{21} =2.26; p=0.034). No data are available for common tern chicks in 2012.

Figure 6. Mean corticosterone concentrations (SD), expressed as picograms of corticosterone per millimetre of feather, in chicks of herring gulls (a) and common terns (b) from St. Marys River AOC colonies (Hay Point, Pumpkin Point, North Sister Rock) and corresponding Lake Huron reference colonies (Double Island or Cousins Island) in 2011 and/or 2012. Data for common terns are for 2011 only. Numbers of feathers ranged from 10-20 per colony for both species. Different uppercase letters show significant differences in mean concentrations within a study year.

a) Herring gulls:



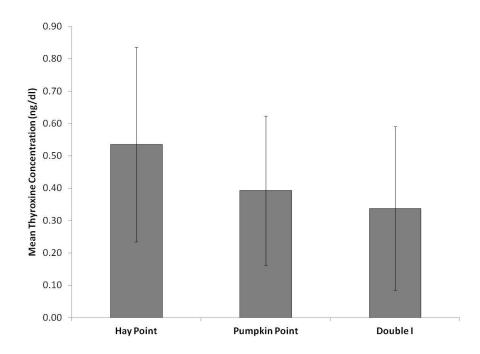
b) Common terns:



Thyroxine in Plasma of Herring Gull Chicks:

Mean thyroxine concentrations (SD) were 0.54 (0.30) ng/dl and 0.39 (0.23) ng/dl in plasma of herring gull chicks from Hay Point and Pumpkin Point, respectively, and 0.34 (0.25) ng/dl in chicks from Double Island in 2012 (Figure 7). No significant difference in mean concentrations was found among colonies. Similarly, no significant differences in thyroxine concentrations were found between chicks from AOC colonies and corresponding reference colonies for herring gulls or common terns in 2011 (data not shown).

Figure 7. Mean thyroxine concentrations (SD) in plasma of herring gull chicks from St. Marys River AOC colonies (Hay Point and Pumpkin Point) and the reference colony (Double Island) in 2012. Numbers of blood samples collected ranged from 13-15 per colony.



DISCUSSION

Based on the low overall contaminant burdens reported in embryos in 2011 and 2012, concentrations of PCBs, other organochlorines and PBDEs were not sufficiently elevated to adversely impact the reproductive success of herring gulls and common terns foraging in the St. Marys River AOC. In a broad literature review of PCB effects in birds, Hoffman *et al.* (1996) concluded that sum PCB concentrations in the range of 8 to 25 μ g/g in eggs were associated with decreased hatching success for terns and cormorants. Sum PCB concentrations in all embryos of both species were well below the 8 μ g/g threshold. Similarly, concentrations of *p*,*p*'-DDE in eggs were well below threshold levels associated with significant effects on reproductive success as reported in black-crowned night-herons (8 μ g/g; Henny *et al.* 1984) and cormorants (10 μ g/g; Pearce *et al.* 1979). These findings concur with those found in herring gull eggs from two U.S. colonies in the St. Marys River AOC (Michigan) between 2002-2006

where concentrations of both compounds were below recommended threshold levels associated with adverse reproductive effects (MDEQ 2012). Comparatively, median PCB and p,p'-DDE concentrations in gulls from these U.S. colonies were approximately two to three times higher than median concentrations in gulls from St. Marys River (Ontario) in 2011 and 2012. Sum PBDE concentrations were also low and well below the lowest-observed effect level on pipping and hatching success in American kestrels (*Falco sparverius*) equal to 1.8 μ g/g in eggs (McKernan *et al.* 2009). Overall, concentrations of all compounds in both species were largely comparable between the AOC colonies and the reference colonies and were in fact significantly lower for some compounds (e.g., PCBs and PBDEs in terns) at an AOC colony than the reference colony.

Concentrations of 2,3,7,8-TCDD, dioxin-like PCBs and total TEQs in embryos of herring gulls and common terns from the St. Marys River AOC (Ontario) were not notably elevated relative to birds from the respective reference colonies in 2011 and 2012. Concentrations of these compounds in terns from AOC colonies were also below levels found in a previous study of common terns at a colony at Lime Island, on the U.S. side of the St. Marys River, and which is within 1 km of the Hay Point colony (Senthilkumar et al. 2003). In that study, ten unhatched eggs collected in 1999 had concentrations of 2,3,7,8-TCDD (range=1.7-6.6 pg/g), dioxin-like PCBs (range in total concentrations of non-ortho PCBs and mono-ortho PCBs=161-612 ng/g), and total TEQs (91-219 pg TEQ/g) which were considered to be elevated and which also coincided with the collapse of the colony in that year. Tern embryos from AOC colonies in 2011 and 2012 were below concentrations at the Lime Island colony in 1999 with corresponding concentrations of 2,3,7,8-TCDD (range=1.2-3.8 pg/g), dioxin-like PCBs (range=44-110 ng/g), and total TEQs (range=63-133 pg TEQ/g). Median total TEQs in herring gull embryos in this study (85 pg TEQ/g) were less than one-half of median total TEQs (222 pg TEQ/g) in herring gull eggs from colonies on the U.S. side of the St. Marys River between 2002-2006 (MDEQ 2012). Total TEQs in herring gulls from the St. Marys River AOC were within the range of mean TEQ concentrations in herring gull eggs from other colonies on the Great Lakes in 2011 and 2012 (Figure 2).

With respect to concentrations associated with toxicity, concentrations of 2,3,7,8-TCDD (<6 pg/g) and total TEQs (<160 pg TEQ/g) in herring gull and common tern embryos from colonies in the St. Marys River AOC were below concentrations associated with effects on reproduction in avian species. Median concentrations of 2,3,7,8-TCDD (37 pg/g) and total TEQs (2175 pg TEQ/g) in eggs of Forster's tern (*Sterna forsteri*) from Lake Michigan were associated with a significant reduction in hatching success while no effect on hatching success was found at relatively lower median concentrations of 2,3,7,8-TCDD (201 pg TEQ/g) in eggs from the reference colony in 1983 (TEQs based on concentrations of 2,3,7,8-TCDD and dioxin-like PCBs only; Kubiak *et al.* 1989). Concentrations of PCB-126 in common terns embryos from the AOC were two orders of magnitude below the lethal dose associated with 50% embryonic mortality in common terns (104 ng/g based on egg injection; Hoffman *et al.* 1998). In addition, 2,3,7,8-TCDD concentrations were below those associated with decreased embryonic growth and edema in herons (150-250 pg/g; Hoffman *et al.* 1996).

Exposure to high concentrations of mercury can have significant impacts on reproductive success in birds and can also result in teratogenic effects in avian embryos, as demonstrated in egg injection studies with methylmercury in the laboratory (Fimreite 1974; Hill *et al.* 2008; Heinz *et al.* 2011). Overall,

mercury concentrations in all embryos were below the predicted threshold of 0.6 μ g/g (wet weight) in eggs as being protective against adverse reproductive effects for 95% of non-marine avian species (Shore et al. 2011). Two common tern embryos from North Sister Rock and the Cousins Island approached this concentration in 2012 (0.57 μ g/g and 0.59 μ g/g, respectively). Common terns however may be less sensitive to the effects of mercury exposure relative to other species. Based on the results of egg injection studies with methylmercury, both common terns and herring gulls exhibited medium sensitivity to mercury compared to 24 other avian species where embryo survival was examined (Heinz et al. 2009); other studies have also demonstrated species-specific differences in mercury sensitivity (e.g., Braune et al. 2012). Fimreite (1974) found no effect on hatching success in common terns at a colony where the mean mercury concentration in eggs $(1 \mu g/g)$ was at least twice those found in this study. While no deformities were found in these two embryos, sublethal effects that impair chick growth, behavior or survival cannot be ruled out however and these may be apparent in birds with relatively higher mercury burdens. Relative to mercury concentrations in herring gull eggs from other Great Lakes colonies in 2011 and 2012, mercury concentrations in gull eggs from the AOC colonies ranked 10th out of 15 herring gull colonies monitored (ranked as means from highest to lowest; EC unpublished). For both herring gulls and common terns, there was no evidence of significantly higher mercury concentrations in embryos from the St. Marys River AOC compared to reference colonies. These results suggest that current mercury concentrations would unlikely impact reproduction of breeding herring gulls and common terns in the St. Marys River AOC.

Elevated concentrations of PCBs and dioxins in eggs were associated with increased incidences of morphological deformities observed in wild populations of colonial waterbirds in the 1970s-1990s (Gilbertson et al. 1976; Ludwig et al. 1996). In 1998, Michigan State University researchers found three cross-billed common tern chicks out of 120 birds at the colony on Lime Island (EC et al. 2002). In this study, surveys of herring gull and common tern chicks at both AOC and reference colonies revealed no evidence of morphological deformities in both study years, a finding which is consistent with low organochlorine and dioxin burdens found in embryos from these sites. In the artificial incubation study however, embryonic deformities were detected in a single embryo from one or more AOC colonies per year, but out of a small sample size of viable embryos. The detection of a limited number of deformed embryos, but not free-living chicks, may not be surprising since affected individuals in wild populations would likely die before hatching or shortly thereafter and therefore not be captured in a deformity survey. It is somewhat puzzling however that deformed embryos were found only at AOC colonies and not at reference colonies for both species in both years. Specifically, three deformed herring gull embryos were found out of 53 viable eggs (6%) and two deformed common tern embryos were found out of 52 viable eggs (4%) when counts for AOC colonies and study years were combined. No deformed embryos were observed out of 46 viable herring gull eggs and 25 viable common tern eggs examined at the reference colonies, Double Island and Cousins Island, in the two years, respectively. Similar concentrations of non-ortho PCBs, 2,3,7,8-TCDD, and TEQs between deformed and normal embryos from AOC colonies of both herring gulls and common terns rule out the possibility that embryonic deformities were associated with exposure to dioxin-like PCBs and dioxins. A third year of egg collection and artificial incubation of eggs in the laboratory will be conducted in 2014 to examine this pattern further at both AOC and reference colonies. Nonetheless, given the low burdens of PCBs, other

organochlorines, dioxins and furans, and mercury found in embryos, it is unlikely that this is a contaminant-related issue.

Reproduction based on productivity values for these two species in the St. Marys River AOC was considered good for herring gulls and was within a range considered typical for common terns in this region. Productivity of herring gulls at all study colonies was consistently high and above 1.3 chicks per nest in both study years. This was in the upper range of productivity levels required to maintain a stable herring gull population (range=0.8-1.4 chicks per nest; Kadlec and Drury 1968). Overall productivity of common terns at both the AOC colony and reference colony was low with comparable estimates of 0.40 and 0.45 chicks produced per nest, respectively, in the two study years. These rates are well below those required to maintain a population (1.1 chicks per nest; DiCostanzo 1980). However, these rates are consistent with low productivity observed as part of an intensive study of site tenacity and productivity in common terns conducted by EC throughout the North Channel in 2010-2012 (Moore and Weseloh 2012). In that study, the overall mean annual productivity of common terns in the region was 0.34 chicks per nest with a range in rates at six colonies in three study years from 0 - 1.0 chicks produced per nest. Productivity estimates for North Sister Rock and Cousins Island cited in this report are included in this overall mean annual productivity value. Low productivity in the North Channel was attributed primarily to egg loss and nestling loss caused by severe weather events (e.g. flooding) and predation by birds and mammals. The latter was evident at Hay Point in 2011 where significant predation led to abandonment of this as a nesting site. These results suggest that low productivity observed for common terns in 2011 and 2012 were not localized to the St. Marys River AOC but rather were consistent with low productivity observed in terns in the North Channel during this period. In an earlier study in the St. Marys River, Michigan in 1980, mean productivity of common terns at 12 colonies was also low at 0.36 chicks fledged per nest (Scharf 1981). Habitat loss associated with high water levels and flooding, erosion of natural and made-made islands, and encroachment by vegetation at nesting sites in the St. Marys River were cited as factors contributing to reduced reproductive success at those nesting sites. Overall, external stressors appear to be largely limiting productivity of terns in the AOC and the region. As effectively demonstrated in the artificial incubation study, intrinsic factors (e.g., contaminants) did not appear to impact hatchability of eggs where embryonic viability was high for both terns and gulls at study colonies in the two years.

Stable isotopes of nitrogen and carbon are used to provide information on trophic position and carbon source in the food web, respectively (Hobson 1999). For both herring gulls and common terns, the results of these analyses indicate that there were distinct and consistent differences in diet between birds from AOC colonies and those from reference colonies in the two study years. Significantly higher δ^{15} N values in embryos from the reference colonies in 2011 and 2012 suggest that the breeding birds from the reference colonies occupied a higher trophic level compared to birds from AOC colonies. Specifically, gulls and terns at the reference colonies may have fed more on fish (or larger fish) compared to birds from AOC colonies which fed at a relatively lower trophic level. For gulls, this could include terrestrial food sources such as small mammals, refuse and plant material (Fox *et al.* 1990). This apparent difference in trophic level between the colonies may have contributed to higher concentrations of some compounds in terns from the reference colony compared to an AOC colony in 2011 and 2012. Based on the isotopic signatures for δ^{13} C, gulls from reference colonies may have fed

more on aquatic-based prey types (with more depleted δ^{13} C signatures) compared to gulls from AOC colonies which fed more on terrestrial-based prey types (with more enriched δ^{13} C signatures). This is consistent with the spatial pattern evident for δ^{15} N in gulls. In the case of piscivorous common terns, birds from the reference colony may have fed more on fish from offshore areas compared to terns from AOC colonies which fed on fish from more inshore areas. Determinations of total clutch volume and intraclutch variation in 3-egg clutches for both species were informative for evaluating potential food stress during the egg production period in the two study years. Similarities for these endpoints between AOC and reference colonies suggest that food availability was likely not limited for laying females.

Two additional endpoints relating to growth and development of chicks were also measured in this study. Corticosterone deposited in growing feathers provides important insight into the physiology of stress during the period of feather growth (Bortolotti *et al.* 2009). As an indicator of stress over time, comparable corticosterone concentrations were found in feathers of herring gull chicks between AOC colonies and the reference colony in both years. However, common tern chicks from the AOC colony had a significantly higher mean concentration of corticosterone in feathers compared to reference colony chicks in 2011. Possible stressors could include low food availability, inclement weather, and/or threats of predation that may have been more prevalent at the AOC colony. Food availability may not have been an issue for terns in the AOC based on preliminary analyses of body condition of tern chicks and further supported by the clutch data for breeding females during the egg-laying period. Thyroxine concentrations in plasma of herring gull chicks were comparable among study sites in 2011 suggesting that there were no differences in chick growth and development or possible endocrine disruption among colonies.

In conclusion, the results of this two-year study suggest that there is little evidence of impaired reproduction or deformities in colonial waterbirds attributable to local contamination effects within the St. Marys River AOC (Ontario). Embryonic viability of herring gulls and common terns was high at AOC colonies. Herring gull productivity at AOC colonies was sufficiently high to maintain a stable population. While considered low for common terns, productivity at AOC colonies was consistent with that in the region and was largely attributable to external stressors, such as predation and severe weather events. No morphological deformities were found in ≥ 21-day-old chicks of either species. While there was some evidence of deformities in a limited number of embryos from AOC colonies, the significance of this is unknown and further investigation of this will be conducted in 2014. Importantly, contaminant burdens (e.g., PCBs, 2,3,7,8-TCDD, and mercury) in gull and tern embryos from the St. Marys River AOC (Ontario) were not notably elevated and were comparable to burdens at respective reference colonies in 2011 and 2012. Concentrations of PCBs, other organochlorine compounds, PBDEs, dioxins and furans and mercury were not sufficiently elevated to adversely impact the reproductive success and development of herring gulls and common terns foraging in the St. Marys River AOC.

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REFERENCES

Bortolotti, G.R., T. Marchant, J. Blas, and S. Cabezas. 2009. Tracking stress: localisation, deposition and stability of corticosterone in feathers. *J. Exper. Biol.* 212: 1477-1482.

Braune, B.M., A.M. Scheuhammer, D. Crump, S. Jones, E. Porter, and D. Bond. 2012. Toxicity of methylmercury injected into eggs of thick-billed murres and arctic terns. *Ecotoxicology* 21: 2143-2152.

DiConstanzo, J. 1980. Population dynamics of a common tern colony. J. Field Ornithol. 51: 229-243.

Environment Canada and Ontario Ministry of the Environment. 2011. *St. Marys River Area of Concern Canadian section. Status of beneficial use impairments September 2010*. Queen's Printer for Ontario. 8 pp.

Environment Canada (EC), United States Environmental Protection Agency, Ontario Ministry of the Environment, and Michigan Department of Environmental Quality. 2002. *The St. Marys River Area of Concern Remedial Strategies for Ecosystem Restoration*. 140 pp.

Fimreite, N. 1974. Mercury contamination in aquatic birds in northwestern Ontario. *J. Wildl. Manage*. 38: 120-131.

Fox, G.A. and D.V. Weseloh. 1987. Colonial waterbirds as bio-indicators of environmental contamination in the Great Lakes. In: *The value of birds*, eds. A.W. Diamond and F.L. Fillion. International Council for Bird Conservation Technical Publication No. 6, Cambridge, U.K.

Fox, G.A. L.J. Allan, D.V. Weseloh, and P. Mineau. 1990. The diet of herring gulls during the nesting period in Canadian waters of the Great Lakes. *Can. J. Zoology* 68: 1075-1085.

Gilbertson, M., R.D. Morris, and R.A. Hunter. 1976. Abnormal chicks and PCB residue levels in eggs of colonial birds on the lower Great Lakes (1971-73). *The Auk*. 93: 434-442.

Heinz, G.H., D.J. Hoffman, J.D. Klimstra, K.R. Stebbins, S.L. Kondrad, and C.A. Erwin. 2009. Species differences in the sensitivity of avian embryos to methylmercury. *Arch. Environ. Contam. Toxicol.* 56: 129-138.

Heinz, G.H., D.J. Hoffman, J.D. Klimstra, K.R. Stebbins, S.L. Kondrad, and C.A. Erwin. 2011. Teratogenic effects of injected methylmercury on avian embryos. *Environ. Toxicol. Chem.* 30(7): 1593-1598.

Henny, C.J., L.J. Blus, A.J. Krynitsky, and C.M. Bunck. 1984. Current impact of DDE on black-crowned night-herons in the intermountain west. *J. Wildl. Manage*. 48(1): 1-13.

Hill, E.F., C.J. Henny, and R.A. Grove. 2008. Mercury and drought along the lower Carson River, Nevada: II. Snowy egret and black-crowned night-heron reproduction on Lahontan Reservoir, 1997-2006. *Ecotoxicology* 17: 117-131.

Hobson, K.A. 1999. Tracing origins and migration of wildlife using stable isotopes: a review. *Oecologia* 120: 314-326.

Hoffman, D. J., C.P. Rice, and T.J. Kubiak. 1996. PCBs and dioxins in birds. In: *Environmental contaminants in wildlife: interpreting tissue concentrations*, eds. W.N. Beyer, G.H. Heinz and A.W Redmon-Norwood, pp. 165-207. SETAC Special Publications, Lewis Publishers, Boca Raton, FL.

Hoffman, D.J., M.J. Melancon, P.N. Klein, J.D. Eisemann, and J.W. Spann. 1998. Comparative developmental toxicity of planar polychlorinated biphenyl congeners in chicken, American kestrels, and common terns. *Environ. Toxicol. Chem.* 17: 747-757.

Kadlec, J.A. and W.H. Drury. 1968. Structure of the New England herring gull population. *Ecology* 49: 644-676.

Kubiak, T.J., H.J. Harris, L.M. Smith, T.R. Schwartz, D.L. Stalling, J.A. Trick, L. Sileo, D.E. Docherty, and T.C. Erdman. 1989. Microcontaminants and reproductive impairment of Forster's tern on Green Bay, Lake Michigan – 1983. *Arch. Environ. Contam. Toxicol.* 18: 706-727.

Ludwig, J.P., H. Kurita-Matsuba, H.J. Auman, M.E. Ludwig, C.L. Summer, J.P. Giesy, D.E. Tillitt, and P.D. Jones. 1996. Deformities, PCBs, and TCDD-equivalents in double-crested cormorants (*Phalacrocorax auritus*) and Caspian terns (*Hydroprogne caspia*) of the upper Great Lakes 1986-1991: testing a cause-effect hypothesis. *J. Great Lakes Res.* 22: 172-197.

McKernan, M.A., B.A. Rattner, R.C. Hale, and M.A. Ottinger. 2009. Toxicity of polybrominated diphenyl ethers (DE-71) in chicken (*Gallus gallus*), mallard (*Anas platyrhynchos*), and American kestrel (*Falco sparverius*) embryos and hatchlings. *Environ. Toxicol. Chem.* 28: 1007-1017.

Michigan Department of Environmental Quality (MDEQ). 2012. Assessment of the bird or animal deformities or reproductive problems beneficial use impairment in Michigan's Great Lakes Areas of Concern. MI/DEQ/WRD-12-032.

Moore, D. and C. Weseloh. 2012. Breeding site tenacity and productivity of common terns nesting on the North Channel of Lake Huron. Poster presented at North American Ornithological Conference Vancouver, Canada.

Nisbet, I. C. 2002. Common tern (*Sterna hirundo*). In: *The Birds of North America Online*, eds. A. Poole. Ithaca: Cornell Lab of Ornithology. Retrieved from the Birds of North America Online: <u>http://bna.birds.cornell.edu/bna/species/618</u>

Ontario Ministry of the Environment (OMOE) and Michigan Department of Natural Resources (MDNR). 1992. *The St. Marys River Area of Concern: Environmental Conditions and Problem Definitions. Remedial Action Plan Stage* 1. 444 pp.

Pearce, P.A., D.B. Peakall, and L.M. Reynolds. 1979. Shell thinning and residues of organochlorines and mercury in seabird eggs, eastern Canada, 1970-1976. *Pest. Monit. J.* 13(2): 61-67.

Scharf, W.C. 1981. The significance of deteriorating man-made island habitats to common terns and ring-billed gulls in the St. Mary's River, Michigan. *Colonial Waterbirds* 4: 155-159.

Senthilkumar, K., W.W. Bowerman, K.F. Millenbah, D.A. Best, T. Takasuga and S. Masunaga. 2003. Polychlorinated-dibenzo-*p*-dioxins/furans and -dioxin-like biphenyls in eggs of common terns from Lime Island, St. Mary's River, Michigan, USA. *Toxicol. Environ. Chem.* 85: 221-232.

Shore, R.F., M.G. Pereira, L.A. Walker, and D.R. Thompson. 2011. Mercury in nonmarine birds and mammals. In: *Environmental contaminants in biota (2nd edition)*, eds. W.N. Beyer and J.P. Meador, pp. 609-624. CRC Press, Boca Raton, FL.

Van den Berg, M., L. Birnbaum, A.T.C. Bosveld, B. Brunström, P. Cook, M. Feeley, J.P. Giesy, A. Hanberg, R. Hasegawa, S.W. Kennedy, T. Kubiak, J.C. Larsen, F.X.R. van Leeuwen, A.K. Dijen Liem, C. Nolt, R.E. Peterson, L. Poellinger, S. Safe, D. Schrenk, D. Tillitt, M. Tysklind, M. Younes, F. Waern, and T. Zacharewski. 1998. Toxic equivalency factors (TEFs) for PCBs, PCDDs, PCDFs for humans and wildlife. *Environ. Health Perspect.* 106: 775-792.

Appendix 1. Mean concentrations (SD) of 4 non-*ortho* PCBs, 6 mono-*ortho* PCBs, and 17 furans and dioxins in embryos of herring gulls (HERG) and common terns (COTE) collected from the St. Marys River (SMR) AOC and reference colonies (i.e., Double Island and Cousins Island) in 2011 and 2012 and incubated in the lab (pg/g, wet weight). Concentrations are shown for deformed embryos (analyzed as individuals) and normal embryos based on the analysis of pools consisting of five individuals. Accordingly, N represents either the number of deformed individuals or the number of pools analyzed.

Compound	HERG-SI	MR AOC	HERG-Double I	COTE-SN	IR AOC	COTE-Cousins I
	Deformed N=3	Normal N=3	Normal N=1	Deformed N=2	Normal N=2	Normal N=1
PCB-77	249.07 (384.48)	61.90 (71.18)	36.70	617.25 (135.41)	548.50 (9.19)	1000.00
PCB-81	120.10 (53.96)	72.30 (9.52)	98.90	55.58 (25.49)	48.05 (5.73)	75.80
PCB-126	699.00 (149.79)	497.67(70.12)	680.00	389.75 (259.15)	346.00 (82.02)	455.00
PCB-169	112.70 (54.67)	92.07 (20.57)	121.00	68.03 (37.16)	70.20 (18.95)	85.20
PCB-105	48102 (5660)	42113 (8771)	59149	18067 (8249)	17970 (279)	16877
PCB-114	3273 (245)	2754 (116)	3150	1045 (502)	974 (131)	713
PCB-118	109721 (12333)	94632 (5426)	116622	45638 (28654)	47239 (10551)	34511
PCB-156	15096 (1125)	15201 (285)	19236	7543 (6161)	7803 (3437)	4891
PCB-157	2609 (129)	3149 (299)	3732	1931 (1512)	1906 (1029)	1251
PCB-189	2468 (945)	2724 (389)	3299	1531 (663)	1716 (261)	1478
2378-TCDD	4.76 (0.51)	3.30 (1.30)	5.62	2.49 (1.84)	2.14 (0.64)	3.80
12378-PeCDD	4.15 (1.93)	3.13 (0.94)	4.49	4.49 (3.24)	4.37 (1.75)	5.87
123478-HxCDD	0.39 (0.25)	0.27 (0.22)	NDR	1.09 (0.53)	0.46 (0.64)	0.95
123678-HxCDD	5.78 (1.40)	4.85 (0.72)	5.26	3.38 (2.10)	3.99 (0.43)	3.68
123789-HxCDD	1.28 (0.50)	0.82 (0.16)	1.15	1.22 (0.45)	1.10 (0.34)	1.00
1234678-HpCDD	3.42 (0.86)	7.64 (3.44)	5.63	2.56 (0.33)	1.85 (0.66)	2.59
12346789-OCDD	8.07 (1.62)	16.26 (8.28)	11.6	5.03 (3.37)	2.33 (0.72)	2.57
2378-TCDF	0.55 (0.95)	۸	۸	6.55 (2.79)	4.51 (6.37)	18.10
12378-PeCDF	0.46 (0.80)	0.03 (0.04)	۸	0.17 (0.23)	0.67 (0.94)	۸
23478-PeCDF	2.92 (2.96)	2.44 (2.14)	3.17	5.11 (3.62)	4.54 (1.34)	4.95
123478-HxCDF	1.07 (0.43)	0.99 (0.46)	1.05	0.97 (0.61)	0.83 (0.32)	1.01
123678-HxCDF	1.43 (0.20)	0.96 (0.30)	1.64	1.17 (0.89)	1.07 (0.21)	1.11
234678-HxCDF	0.82 (0.57)	0.66 (0.19)	0.97	0.90 (0.37)	0.87 (0.06)	1.04
123789-HxCDF	NDR	0.09 (0.02)	0.21	0.17 (0.09)	0.11 (0.06)	0.12
1234678-HpCDF	NDR	NDR	5.05	0.13 (0.18)	0.04 (0.06)	0.12
1234789-HpCDF	0.18 (0.08)	0.09 (0.08)	NDR	0.07 (0.03)	0.19 (0.10)	0.16
12346789-OCDF	1.13 (0.51)	1.04 (0.52)	1.16	0.58 (0.21)	0.52 (0.05)	0.53

NDR = Not detected due to incorrect isotope ratio; ^ Possible chlorinated diphenyl interference

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